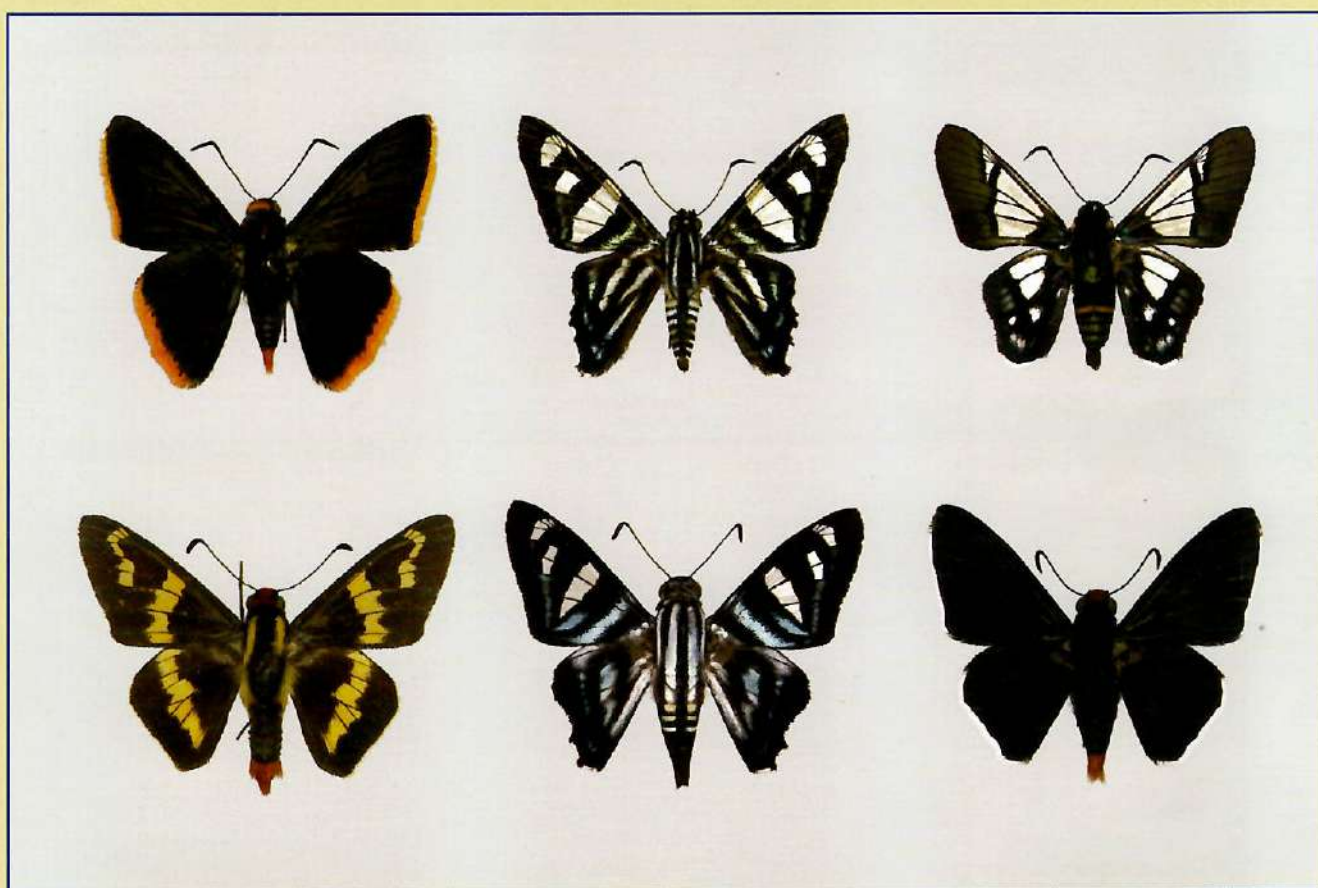


Butterflies of the world

est. by Erich Bauer & Thomas Frankenbach (†)



Part 49

O. H. H. Mielke, E. Brockmann & C. G. C. Mielke

Hesperiidae II

New World Pyrrhopyginae, detailed text

Butterflies of the World 49

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Neotropical Hesperiidae: Pyrrhopyginae. Part 1

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ABSTRACT

All Pyrrhopyginae taxa (Hesperiidae) are illustrated in dorsal and ventral views in both sexes wherever possible, followed by their geographical distributions. One new genus and some new species and subspecies are described, and new combinations and new, revalidated and reinstated stati are presented. Primary types are identified and mentioned with their labels whenever possible. Lectotypes and neotypes are designated to stabilise correct identification of taxa with one exception: *Mimoniades minthe* var. *egena* Mabille & Boulet, 1908, due to the lack of suitable specimen. In total, 50 genera (one new), 220 species (20 new) and 142 subspecies (18 new) are recognized as valid. New synonymies are proposed for 17 species-group names.

INTRODUCTION

The book series “Butterflies of the World” edited by Erich Bauer and Thomas Frankenbach aims to figure all taxa of a given taxonomic group, presenting their geographical distribution, describing new taxa and ensuring correct identifications by designating lectotypes and neotypes when necessary succinctly.

MATERIAL AND METHODS

Evans (1937, 1949, 1951, 1952, 1953, 1955) was the most important researcher on the Hesperiidae and we follow him, whenever possible. His publications are the base of our work, and he is cited frequently. Some other more recent authors are also cited (e.g. O. Mielke 1995, O. Mielke 2002, Warren *et al.* 2008, 2009, Orellana [2010], Li *et al.* 2019).

The geographical distributions include all references cited in the “Catalogue of the American Hesperioidea, Vol. 1 Pyrrhopyginae” (O. Mielke 2005) and from its updated version until 2020, available in the site of the “Laboratório de Estudos de Lepidoptera Neotropical”: <https://sites.google.com/site/lablepneo/home>, from the “Butterflies of America” (BoA): <http://butterfliesofamerica.com/> and from specimens deposited in several collections listed below. Only references considered to have correct identifications are listed.

Some subspecies of the same species are mentioned as sympatric, a fact that is probably due to errors of identification in the literature, mislabelling, lack of detailed study and/or lack of specimens for clarification.

Bogotá, Cundinamarca, Colombia, a common locality in old references, is unclear; probably this locality refers to a place near Bogotá, at a lower altitude (Fassl 1914: 98).

DNA barcode follows the methodology of Hebert *et al.* (2003), Hajibabaei *et al.* (2006), Ratnasingham & Hebert (2007), Cong & Grishin (2014) and Zhang *et al.* (2019).

For the dissection of male and female genitalia, the abdomen was removed, immersed in water for about 24 hours in natural conditions and then placed in a petri dish with paraffin at the bottom, where the abdomen was fixed with one insect pin. The pleura was cut, the abdomen opened and the genitalia removed. The genitalia was then soaked in a tube with heated potassium hydroxide solution (KOH) for a few minutes. Finally, the genitalia was dissected and preserved in microvials with glycerol along with the specimen. The abdomen

was dried and glued on the specimen, preserving its structure and colours.

For each taxon, the labels of the holotype, lectotype or neotype are given verbatim whenever possible, separated by forward slashes. Some were designated here to guarantee a correct identification, considering that some taxa have similar species or subspecies, which can lead to misidentifications. For each neotype designation, we provide a justification, a bibliographic reference that correctly identifies the taxon, affirmation of the absence of a primary type (holotype, lectotype), the collection depository, the collecting locality (the closest to the lost type), and the illustration of the specimen. We believe that each taxon must have a specimen that represents it, that is, a holotype, lectotype or neotype. The unique exception is for *Mimoniades minthe* var. *egena* Mabille & Boulet, 1908, for which neither a primary type nor a specimen compatible with the original description was found.

Authorship of some newly described taxa are of authors who do not co-author the present book. They are: Mirna M. Casagrande (Curitiba) and Gerardo Lamas (Lima).

The figures that complement the descriptions of the new species and subspecies are found in the Part 2, published along with this part.

Finally, we list all examined specimens with barcodes along with their voucher numbers in BOLD System (<http://www.boldsystems.org/>) or in Nick Grishin's lab and in GenBank.

Abbreviations of the collections (curator(s) between parentheses).

AMNH	American Museum of Natural History, New York, USA (the late Frederick Rindge)
AME	The Allyn Museum of Entomology, Sarasota, Florida, USA – transferred to MGCL
ANSP	Academy of Natural Sciences, Philadelphia, Pennsylvania, USA – transferred to CMP (John Rawlins)
AO	Collection Andrés Orellana, Mérida, Venezuela
BH	Collection Bernard Hermier, Rémire-Montjoly, Cayenne, French Guiana
CB	Collection Christian Brévignon, Matoury, Cayenne, French Guiana
CFC	Collection family Constantino, Cali, Valle del Cauca, Colombia
CFR	Collection family Romero, Maracay, Aragua, Venezuela
CJS	Collection Julián Salazar, Manizales, Caldas, Colombia
CJU	Collection Jiro Uehara, Yokohama, Japan
CMP	Carnegie Museum, Pittsburgh, Pennsylvania, USA (John Rawlins)
CORN	Cornell University, Ithaca, New York, USA (John G. Franclemont)
DEI	Deutsches Entomologisches Institut, Münchenberg, Germany (Reinhard Gaedike)
DZUP	Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil (Olaf H. H. Mielke, Mirna M. Casagrande)
EB	Collection Ernst Brockmann, Lich, Germany. To be transferred to Museum Senckenberg, Frankfurt am Main, Germany
FMNH	Field Museum of Natural History, Chicago, Illinois, USA

HMUG	Hunterian Museum, University of Glasgow, Glasgow, UK
IML	Instituto Miguel Lillo, Tucumán, Argentina (Kenneth J. Hayward)
IN	Collection Ichiro Nakamura, Williamsville, New York, USA
IOC	Instituto Oswaldo Cruz, Rio de Janeiro, Rio de Janeiro, Brazil (Lauro P. Travassos, Sebastião Oliveira)
IRSC	Institut Royal de Sciences Naturelles, Brussels, Belgium (Patrick Grootaert)
LACM	Los Angeles County Museum of Natural History, Los Angeles, California, USA
LCRO	Collection Jean Francois Le Crom, Bogotá, Colombia
LSL	Linnean Society of London, London, UK
MABR	Museo Argentino de Ciencias Naturales “Bernadino Rivadavia”, Buenos Aires, Argentina (Manuel Vianna)
MCZ	Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, USA
MfN	Museum für Naturkunde, Berlin, Germany (Hanz-Joachim Hannemann, Wolfram Mey, Théo Léger)
MGCL	McGuire Center for Lepidoptera and Biodiversity, University of Florida, Gainesville, Florida, USA (the late Thomas C. Emmel, George Austin, Jacqueline Miller, Keith R. Willmott, Andrew D. Warren)
MIZA	Museo del Instituto de Zoología Agrícola, Universidad Central de Venezuela, Maracay, Aragua, Venezuela (Francisco F. Yépez, José Clavijo)
MLP	Museu de La Plata, La Plata, Argentina (Luiz de Santis)
MNHN	Muséum national d'Histoire naturelle, Paris, France (the late Pierre Viette, the late Georges Bernardi, Jacques Pierre, Rodolphe Rougerie)
MNRJ	Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil (Alexandre Soares). Destroyed by fire at September 2 nd , 2018
MSFR	Museum Senckenberg, Frankfurt, Germany (Heinz Schröder, Wolfgang Nässig)
MUSM	Museo de Historia Natural, Universidad Nacional Mayor de San Marcos, Lima, Peru (Gerardo Lamas)
NBC	Naturalis Biodiversity Center, Leiden, The Netherlands (Rienk de Jong, Behnaz Bekkum-Ansari, Eulàlia Gassó)
NHMUK	The Natural History Museum, London, U.K. (Richard Vane-Wright, Phillip R. Ackery)
NMI	National Museum of Ireland, Dublin, Ireland
OM-DZUP	Collection Olaf H. H. Mielke, Curitiba, Paraná, Brazil, deposited in DZUP
OUMNH	University Museum, Oxford University, Oxford, UK
SF	Collection Serge Fernandez, Rémire-Montjoly, Cayenne, French Guiana
SMT	Senckenberg Museum für Tierkunde, Dresden, Germany (Matthias Nuss)
SMW	Staatliches Museum Wiesbaden, Wiesbaden, Germany (Michael Geisthardt)
USNM	National Museum of Natural History, Washington, D.C., USA (John Burns, Robert K. Robbins)
UZI	Universitets Zoologiska Institut, Uppsala, Sweden (Sten Jonsson)
ZfB	Zentrum für Biodokumentation, Landsweiler-Reden, Germany (includes the Hesperiidae collection H. Fassl) (Andreas Werno)
ZISP	Zoological Institute, Russian Academy of Sciences, St. Petersburg, Russia (Aleksey Zinovjev, Vladimir Kuznetzov)
ZMUC	Zoologisk Museum, Universitets Copenhagen, Copenhagen, Denmark (Niels P. Kristensen)
ZSM	Zoologische Staatssammlung München, München, Germany (Walter Forster, Wolfgang Dierl, Axel Hausmann)

Hesperiidae (Latreille, 1809) Stephens, 1828

The Hesperiidae, commonly called skippers, form a monophyletic group based on some morphologic apomorphies: the distance between the base of each antenna is at least twice the width of the scape; basal ring of the eyes formed by very reduced ommatidia; third axillary sclerite of forewing very wide and indented anteriorly, narrow and curved posteriorly, forming a very irregular Y-shaped structure; vein R of hindwing apparently simple, touching Sc at a short distance from base to form a narrow cell and R completely fused with Sc, indiscernible as a separate vein; area of very small, specialized scales on dorsal hindwing, covering basal part of R (Ackery *et al.* 1998). Some other characters may be mentioned: the recurved apiculus of the antennae and the forewing venation normally with 12 free veins, but there are some exceptions in Pyrrhopyginae: *Olafia roscius* (Hopffer, 1874) lacks Sc; *Oxynetra* C. & R. Felder, 1862 and *Dis* Mabilie, 1889 species have a reduced Sc, and some other species have anastomosed veins on the fore and hindwings.

The family is divided into 13 currently recognised subfamilies: Barcinae Grishin, 2019, Coeliadinae Evans, 1937, Chamundinae Grishin, 2019, Eudaminae (Burmeister, 1878) Pelham, 2008, Euschemoninae (Kirby, 1897) Mabilie & Boulet, 1908, Hesperinae (Latreille, 1809) Watson, 1893, Heteropterinae Aurivillius, 1925, Katreinae Grishin, 2019, Malazinae Lees & Grishin, 2020, Pyrginae (Burmeister, 1878) Speyer, 1879, Pyrrhopyginae (Mabilie, 1877) Watson, 1893, Tagiadinae (Mabilie, 1878) Li *et al.*, 2019 and Trapezitinae Waterhouse & Lyell, 1914. Six subfamilies occur in the Neotropics: Eudaminae, Hesperinae, Heteropterinae, Pyrginae, Tagiadinae and Pyrrhopyginae, only the latter is treated in this book.

Distribution is worldwide, from sea level to above 5000 m; flight normally very fast and erratic, quite different from all other Papilionoidea.

The following plant families are used as foodplant: Anacardiaceae, Annonaceae, Berberidaceae, Clusiaceae, Combretaceae, Cunoniaceae, Escalloniaceae, Euphorbiaceae, Fabaceae, Fagaceae, Flacourtiaceae, Lauraceae, Malpighiaceae, Melastomataceae, Meliaceae, Myristicaceae, Myrtaceae, Pinaceae, Rutaceae, Sapindaceae and Sterculiaceae (Santin 2004, Beccaloni *et al.* 2008).

Pyrrhopyginae (Mabilie, 1877) Watson, 1893

The Pyrrhopyginae was considered by recent authors (Warren *et al.* 2008, 2009, Sahoo *et al.* 2016, 2017, Toussaint *et al.* 2018) as a tribe of Pyrginae, however Li *et al.* (26-III-2019) based on a genomic basis, confirmed its subfamily status: Pyrrhopyginae.

Li *et al.* (2019) also noted that the Pyrrhopyginae has two tribes: Pyrrhopygini with four subtribes (Passovina, Zoniina, Oxynetrina and Pyrrhopygina) and Jerini Grishin in Li *et al.* (2019), Grishin (2019). The latter only includes the genus *Jera* Lindsey, 1925 with one species: *Jera tricuspidata* (Mabilie, 1902). Zhang *et al.* (25-V-2019) presented different conclusions, although the team of authors is partially the same and led by Nick Grishin (the last author in both articles), with Pyrrhopyginae comprising five tribes: Azonaxini Grishin, described herein, Zoniini, Passovini, Pyrrhopygini, and Oxynetrini, so that, Jerini was no longer considered, but confirmed by Grishin (pers. comm.). In addition, Zhang *et al.* (25-V-2019) provided a systematic view of the entire subfamily with three different results based on the taxa divergence time: i) 15 million years, ii) 10 million years, and iii) 5 million years, being that of 10 millions of years which received more emphasis and was considered to propose some taxonomic acts within Pyrrhopygini: i) three subtribes described (Apyrrothrixina Grishin, Mimoniadina Grishin, and Microcerisina Grishin); ii) 10 subgenera described: *Aesculapyge* Grishin, 2019, *Sarbienna* Grishin, 2019, *Santea* Grishin, 2019, *Mimadia* Grishin, 2019, *Jember* Grishin, 2019,

Jematus Grishin, 2019, *Jemasonia* Grishin, 2019, *Merobella* Grishin, 2019, *Blubella* Grishin, 2019, and *Apatiella* Grishin, 2019; iv) four genera synonymised (*Cyanopyge* O. Mielke, 2002 with *Melanopyge* O. Mielke, 2002, *Mimardaris* O. Mielke, 2002 with *Ardaris* Watson, 1893, *Metardaris* Mabilite, 1903 with *Sarbia* Watson, 1893, and *Elbella* Evans, 1951 with *Microceris* Watson, 1893); v) one genus revalidated (*Agara* Mabilite & Boulet, 1908) and finally; vi) 12 genera revised to subgeneric status (*Melanopyge* O. Mielke, 2002, *Chalpyge* O. Mielke, 2002, *Mysarbia* O. Mielke, 2002, *Mysoria* Watson, 1893, *Amyrsoria* O. Mielke, 2002, *Amenis* Watson, 1893, *Mabotis* Watson, 1893, *Ochropyge* O. Mielke, 2002, *Pseudocroniades* O. Mielke, 2002, *Hegesippe* Evans, 1951, *Olafia* Nemésio, 2005 and *Dis* Mabilite, 1889). The generic phylogenetic divergence treatment of approximately 5 million years is adopted here, since it causes the least systematic change in the group (O. Mielke 2005); in fact, this is a recommendation by the authors themselves (Zhang *et al.* (25-V-2019, S1 Appendix: 17)). Some modifications of this treatment are considered and explained in the appropriate genus, as a result of complementary morphological considerations.

Pyrrhopyginae is a monophyletic group with a characteristic phenotype (Li *et al.* (26-III-2019)). In particular the antenna with its unique shape which enables immediate identification of whether a specimen belongs to this group or not, including the more or less perfect coloured mimetic species of other subfamilies as explained below. But, there is an exception, *Jera tricuspidata*, a very strange species, included in Pyrrhopyginae as discussed above.

The distribution of the species of Pyrrhopyginae is almost completely restricted to the Neotropical region, with only one exception – *Apyrrothrix arizonae* (Godman, 1893), which reaches the south of the Nearctic region (USA – Arizona, New Mexico and Texas). The remainder occur from northern Mexico to Bolivia, northern and northwestern Argentina and Uruguay, showing a higher diversity at the Andes zone, being absent in the main Caribbean islands.

The mimicry rings

It is interesting to note a number of notable cases of probable mimicry, until now not explained, either between several taxa within Pyrrhopyginae or even between this and other subfamilies of Hesperidae. This fact was first mentioned by Bell (1931) and later by Evans (1941, 1951). Bell concluded that the various similar phenotypes present in *Pyrrhopyge* species represented distinct species, as the male genitalia were completely different. Evans (1941) defined four phenotypes as follows: “types of facies, viz.: *A. phidias*: plain dark blue above and below. *B. bixae*: resembling *phidias* above, but with a well-marked basal white band on the hind-wing underside. *C. hyperici*: resembling *bixae* below, but with a large pale blue area on the upperside of the hind-wing. *D. sergius*: resembling *phidias* above, but with a broad submarginal white band on the hind-wing underside”. Later, Evans (1951) called these phenotypes “*phidias* form”, “*bixae* form”, “*hyperici* form” and “*sergius* form” respectively and characterized the subspecies with these names where appropriate.

The first group, “*phidias* form”, comprises several species similar to *Pyrrhopyge phidias* (Linnaeus, 1758), *Elbella intersecta* (Herrich-Schäffer, 1869) (Pyrrhopygini) and *Passova passova* (Hewitson, 1866) (Passovini). Thus, there are two different tribes in which the four phenotypes occur.

Only *Pyrrhopyge phidias* shows all four phenotypes in its subspecies, while the other species contain only one, two or three, but interestingly subspecies with the same phenotypes have the same geographic distribution. The “*hyperici* form” subspecies all occur in the state of Pará, Brazil with two exceptions: *P. aziza aziza* Hewitson, 1866, in Ecuador and *Passova passova gortyna* (Hewitson, 1866) in Tefé, Amazonas, Brazil (type-locality), although it is present in Pará as well. The subspecies of the “*sergius* form” only occur on the eastern side of the Andes between Ecuador and Bolivia, while the species or subspecies of “*phidias* and *bixae* forms” are only present in the Amazon, where they are sympatric. The “*phidias* form” also extends its distribution towards central and southern Brazil. The “*phidias* form” and “*bixae* form” phenotypes are also present in a species of Hesperinae

(*Pyrrhopygopsis socrates* (Ménétriés, 1855)) with distribution ranging from Colombia to Bolivia, Paraguay, northern Argentina and southern Brazil. Curiously, the “*phidias* form” only occurs in the southern part of its distribution.

A second, very complex mimicry assemblage occurs between species of *Jemadia* Watson, 1893, *Jemasonia*, *Jematus*, *Jember*, *Nosphistia* Mabilite & Boulet, 1908, *Merobella*, *Parelbella* O. Mielke, 1995, and *Protellabella* O. Mielke, 1995 (Pyrrhopygini). The species of these genera, together with the female of *Croniades auraria* H. H. Druce, 1908 (Pyrrhopygini), *Granila paseas* (Hewitson, 1857) (Passovini) and *Zonia zonia* Evans, 1951 (Zoniini) belong to three tribes, all of which display blue lines on both dorsal and ventral sides of the wings and three sets of semi-hyaline spots on the forewing. In addition, species of *Granila* Mabilite, 1903 and *Zonia* Evans, 1951 bear a similar elongated spot in the discal cell of the hindwings. The distribution ranges from Mexico to Bolivia, Paraguay, northern Argentina and southern Brazil (Rio Grande do Sul). This mimetic ring includes members of another subfamily (Eudaminae), such as the species of *Phocides* Hübner, [1819], similar to *Phocides pigmalion* (Cramer, 1779), and *Tarsoctenus papias* (Hewitson, 1857).

A third case concerns the phenotypic similarity of two species of *Hegesippe* (*H. luteizona* (Mabilite, 1877) and *H. hegesippe* (Mabilite & Boulet, 1908)) with several species of *Sarbia* Watson, 1893, with overlapping distributions. A further two species of Hesperinae are included with overlapped distribution: *Pseudosarbia* Berg, 1897 (*P. phoenicicola* Berg, 1897 and *P. flavofasciata* Skinner, 1921).

A fourth and final case of mimicry occurs between two similar species belonging to distinct genera: *Mimoniades* Hübner, 1823 (*M. ocyalus* Hübner, 1823) and *Apatiella* (*A. iphinous iphinous* (Latreille, [1824])). Both taxa have three sets of similar yellow spots on the forewing with the median spot in *M. ocyalus* occupying two spaces M_2 - M_3 and M_3 - CuA_1 and in *A. iphinous iphinous* occupying only one space M_3 - CuA_1 .

JERINI Grishin, 2019

One genus.

TAXONOMY – The tribe is characterised by the genome and by some morphological characters: forewing with outer margin concave in the space M_3 - CuA_1 ; hindwing produced with a short and stout bifurcated tail extending in space CuA_1 - CuA_2 ; a tornal lobe; antennal club blunt without apiculus; palpi long, and forewing cell longer than 3/5 of costa (Grishin *in* Li *et al.* 2019, Grishin 2019).

Jera Lindsey, 1925

One species.

DISTRIBUTION – Northwestern and central Colombia to central-eastern Peru.

Jera tricuspidata (Mabilite, 1902)

Pl. 1, Figs. 1–2 (♂) (d, v)

DISTRIBUTION – Northwestern and central Colombia to central-eastern Peru.

Colombia: Antioquia (*Medellín*); Caldas (*Manizales*). **Ecuador:** Pastaza (*Canelos*); Tungurahua (*Ambato*, *Cruz Loma*, *Yunguilla*). **Peru:** Pasco (*Huancabamba*); San Martín (*Rioja – Nascientes del Río Negro*).

REMARKS – Poorly represented in collections. The female remains unknown.

TYPE:

Dichelura tricuspidata Mabilite, 1902. Holotype (one female [recte male] is mentioned) with the following labels: / Type H T/ Ecuador/ Type Mabilite/ Ex musaeo P. Mabilite 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

AZONAXINI Grishin, 2019

One genus.

TAXONOMY – The tribe is characterized by the genome (Grishin in Zhang *et al.* 2019) and by some morphological characters: uncus bifid; gnathos well developed as a transversal plate with a median distal projection; harpe elongated and directed dorsally; ampulla absent; aedeagus thin and longer than valva; vesica with a few long cornuti or a small area with very small cornuti; male with dense brushes on internal side of femora and tibiae of the hindlegs (as in *Agara*, *Granila*, *Myscelus* and *Passova* species), and forewing apex truncate and excavate.

Azonax Godman & Salvin, 1893

Three species.

DISTRIBUTION – Southern Mexico to northern Brazil and French Guiana.

REMARKS – All species of *Azonax* are very similar and identification is best confirmed by genitalic examination (female genitalia unknown) or DNA.

Azonax guiana O. Mielke, Brockmann & C.

Mielke, *sp. n.*

Pl. 1, Figs. 3 (allotype ♀), 4 (holotype ♂) (d, v); Text-Fig. 1 (♂ genitalia)

DISTRIBUTION – Guyana and French Guiana.

Guyana. French Guiana: Cayenne (Dégrad Lalanne, Galion, Montagne des Chevaux, Montagne de Kaw).

The citation of *Azonax typhaon* in Guyana (Evans 1952) probably refers to this species.

DESCRIPTION – Male: forewing length 25 mm. Very similar to *Azonax typhaon*, but distinguished dorsally by a little more darker greyish ground colour, more yellowish in *A. typhaon*; less falcate forewing apex; yellowish semi-hyaline spots: R_2 – R_3 larger than R_1 – R_2 , R_5 – M_1 reaches R_2 – R_3 , R_3 – R_4 do not partially overlap, R_4 – R_5 , M_2 – M_3 narrow only in the under half of the space, M_3 – CuA_1 with proximal and distal margins straight, CuA_1 – CuA_2 with posterior margin markedly narrower than anterior margin, and spot below spot in CuA_1 – CuA_2 , in CuA_2 –2A, absent. Hindwing with the semi-hyaline white spot in discal cell rectangular, with anterior margin projected, whereas in *A. typhaon* more pentagonal; ventrally, the yellowish spot in CuA_2 –2A, posterior to the semi-hyaline white cell spot, squared (rectangular in *A. typhaon*).

Tegumen and uncus, in dorsal view, narrowing only in the distal part; uncus bifid with arms well apart and divergent forming a wide V; gnathos with a broad median distal projection; valva with harpe elongated, sickle-shaped and slightly bent upwards; aedeagus with a cornutus as a diffuse sclerotised spot.

Female: forewing enlarged when compared to male and with outer margin more rounded, apex falcate; all semi-hyaline spots whitish, in Sc – R_1 and M_1 – M_2 more hour glass-shaped. Hindwing expanded at M_3 .

TAXONOMY – Best confirmed by genitalic examination (female genitalia unknown) or DNA.

ETYMOLOGY – The specific name refers to the type locality – French Guiana, where the holotype was collected.

REMARKS – Locally common during March and April in French Guiana.

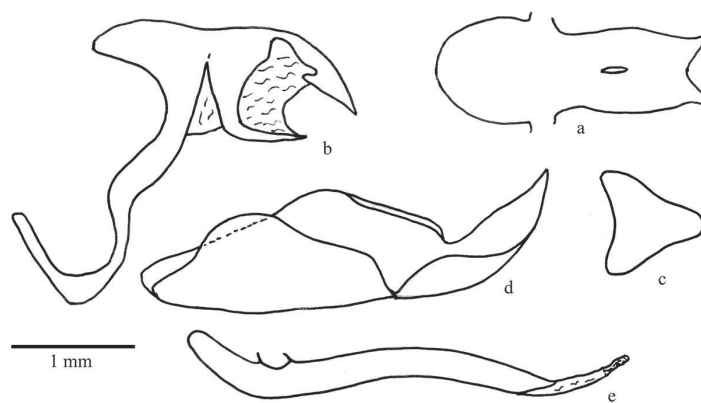
TYPES:

Holotype male with the following labels: / Holotypus/ 9-IV-1993 Dégrad Lalanne, Montagne de Kaw, [Cayenne] Guiana Francesa Gallard leg./ ♂/ gen[itália]. prep[arada]., [O.] Mielke 2018/ Holotypus *Azonax guiana* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/ OM 33.689/ BC-OM/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ 60/

Montagne des Chevaux, 4°43'30" N, 52°24'30" W, 11.V.1995, J.-Y. Gallard [leg.]/ Hermier n° 7204 in cop[ula]. 7203/ ex coll. Bernard Hermier, Montjoly, [Cayenne,] Fr. Guiana, IX-2017/. EB.

Paratypes: 1 ♂, 9-IV-1993, Dégrad Lalanne, Cayenne, French Guiana, 4°34' N, 52°13' W, J.-Y. Gallard leg.; 1 ♂, 3-IV-1988, B. Hermier leg., 4 ♂, 17-IV-1988, 2 ♂, 2-IV-1889, 1 ♂, 30-III-1991, 1 ♂, 8-V-1992, 1 ♂ 1 ♀, 22-V-1992 (*in copula*), 1 ♂, 18-I-1993, 1 ♂, 10-IV-1994, 1 ♂, 29-IV-1995, 1 ♂, 11-V-1996, 1 ♂ 1 ♀, 19-IV-1997 (*in copula*), 1 ♂, 4-V-2002, J.-Y. Gallard leg., Montagne des Chevaux, Cayenne, French Guiana, 4°43'30" N, 52°24'30" W, nrs. 4705, 1499, 1500, 1501, 1502, 1503, 1957, 1958, 2687, 3328, 3311, 3312, 5737, 6095, 7205, 10066, 12428, 12429, 20094 (BH); 1 ♂, 17-III-2008, PK 21, Route de Kaw, Roura, Cayenne, French Guiana, nr. 100 (SF); 3 ♂, 29-III-1988, 4-IV-1988, 11-IV-1989, 1 ♀, 11-IV-1989, Galion, Cayenne, French Guiana, Brévignon leg., nrs. 731, 11, 838, 839 (CB); 2 ♂, 12-IV-1989, PK19 D5, Montsinery, Cayenne, French Guiana, nrs. 12, 13 (CB); 1 ♂, 9-IV-1993, Dégrad Lalanne, Montagne de Kaw, Cayenne, French Guiana, 4°34' N, 52°13'30" W, J.-Y. Gallard leg., Hermier nr. 4734 (EB); 1 ♂, 18-IV-1994, 1 ♂ 1 ♀, 11-V-1995 (*in copula*), Montagne des Chevaux, Cayenne, French Guiana, 4°43'30" N, 52°24'30" W, J.-Y. Gallard leg., Hermier nrs. 6096, 7203, 7204 (EB); 1 ♂, 20-III-1988, Montagne des Chevaux, Cayenne, French Guiana, 4°43'30" N, 52°24'30" W, J.-Y. Gallard leg., Hermier nr. 1476 (IN); 1 ♂, 16-IV-1989, Montagne des Chevaux, Cayenne, French Guiana, 4°43'30" N, 52°24'30" W, J.-Y. Gallard leg., Hermier nr. 1999 (AMNH); 1 ♂, 9-IV-1993, Montagne des Chevaux, Cayenne, French Guiana, 4°43'30" N, 52°24'30" W, J.-Y. Gallard leg., Hermier nr. 4706 (CORN); 1 ♂, 7-IV-1991, Montagne des Chevaux, Cayenne, French Guiana, 4°43'30" N, 52°24'30" W, J.-Y. Gallard leg., Hermier nr. 2686 (MNH).



Text-Fig. 1. *Azonax guiana* *sp. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) gnathos, ventral view; d) right valva, internal view; e) aedeagus, lateral left view (holotype).

Azonax typhaon (Hewitson, 1877)

Pl. 1, Figs. 5 (lectotype ♂) (d, v); Text-Fig. 2 (♂ genitalia)

DISTRIBUTION – Southern Mexico, Nicaragua and eastern Colombia.

Mexico: Chiapas; Oaxaca (*Chimalapa*). **El Salvador** (?).

Nicaragua. Colombia: Meta (*Sierra de la Macarena* – *Río Cano Cristales*).

TAXONOMY – The male genitalia are illustrated for comparison with congeneric species. The tegumen and uncus, in dorsal view, narrowing gradually; uncus bifid with arms slightly apart and slightly converging; gnathos with median distal projection thin (see Evans 1951); valva with harpe bent dorsally at right angle; aedeagus with bristles like cornuti.

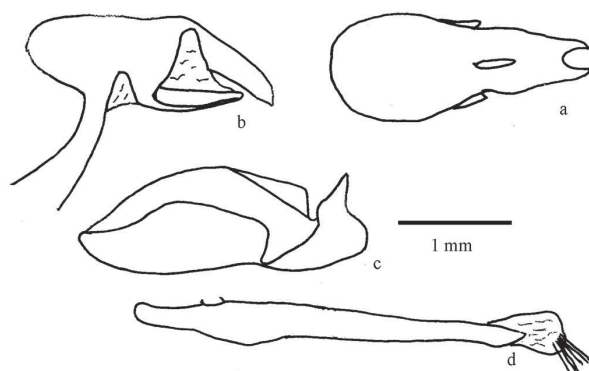
Azonax typhaon has a ground colour more yellow dorsally, while *A. guiana* *sp. n.* darker greyish, and *A. urupa* *sp. n.* less intense yellowish; *A. typhaon* and *A. urupa* *sp. n.* have a more falcate forewing apex; *A. guiana* *sp. n.* has the external margin of the forewing less concave; *A. urupa* *sp. n.* has the apical semi-hyaline yellowish spots on the forewing a little larger; in *A. guiana* *sp. n.* the semi-hyaline yellowish

spot in M_2 – M_3 on the forewing only in the lower part of the space; in *A. urupa* **sp. n.** the lower part of the semi-hyaline yellowish cell spot on the forewing more expanded distally; *A. urupa* **sp. n.** with a small whitish spot on the dorsal forewing in CuA_2 – $2A$ below the semi-hyaline spot in CuA_1 – CuA_2 ; in *A. urupa* **sp. n.** the outer margins, mainly on the dorsal hindwing, greyer, and there are a fine dark marginal line; in *A. urupa* **sp. n.** forewing ventrally with the area between the yellowish semi-hyaline spots of the costa, apex, discal cell and M_2 – M_3 yellowish; *A. urupa* **sp. n.** with submarginal and inner margins areas of the ventral hindwing light grey. Only the holotypes of *A. guiana* **sp. n.** and *A. urupa* **sp. n.** and the lectotype of *A. typhaon* (herein designated) were examined. *Azonax typhaon* is only known from a few specimens in official collections.

REMARKS – Poorly represented in collections.

TYPE:

Myscelus typhaon Hewitson, 1877. The original description has no data. To ensure the correct identification of *Myscelus typhaon* Hewitson, 1877, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H. 83/ Nicg/ *typhaon*/ Nicaragua Hewitson Coll. 79-69/ *Myscelus typhaon*, Hew/ B.M.(N.H.) Rhopalocera slide No. 13015/ B.M.(N.H.) Rhopalocera slide No. 13015/ NHMUK(E)#810373/; two other labels will be added: / Lectotypus/ Lectotypus *Myscelus typhaon* Hewitson, 1877, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.



Text-Fig. 2. *Azonax typhaon*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

Azonax urupa O. Mielke, Brockmann & C. Mielke, **sp. n.**

Pl. 1, Figs. 6 (holotype ♂) (d, v); Text-Fig. 3 (♂ genitalia)

DISTRIBUTION – Northern Brazil (Rondônia).

Brazil: Rondônia (Candeias do Jamari – Fazenda Urupá).

DESCRIPTION – Male: forewing length 27 mm. Very similar to its congeneric species, but distinguished dorsally by a less intense yellowish ground colour, more greyish outer margins, more compact and a little larger yellowish semi-hyaline spots on the apex, and the yellowish semi-hyaline discal cell spot with the anterior half slightly recessed proximally and the posterior half slightly projected distally under the anterior half; ventrally with the area between the yellowish semi-hyaline spots of the costa, apex, discal cell and M_2 – M_3 yellowish.

Hindwing dorsally with the outer margin narrowly chestnut; submarginal area lighter and greyish; chestnut spots in R_s – M_2 with white semi-hyaline centre and the two spots in M_2 – CuA_1 with only a few white scales; semi-hyaline white spot in discal cell slightly squared; ventrally with submarginal and inner margin areas light grey, and the yellowish spot in CuA_2 – $2A$, posteriorly to the semi-hyaline white cell spot, squared, as in *A. guiana* **sp. n.** rectangular in *A. typhaon*.

Tegumen and uncus, in dorsal view, narrowing only in the distal part; uncus bifid with arms well apart by a rounded cavity and slightly convergent; gnathos with the median distal projection thin; valva with elongated harpe, uniformly broad and slightly bent upwards; aedeagus with a cornutus as a diffuse sclerotised spot.

Female: unknown.

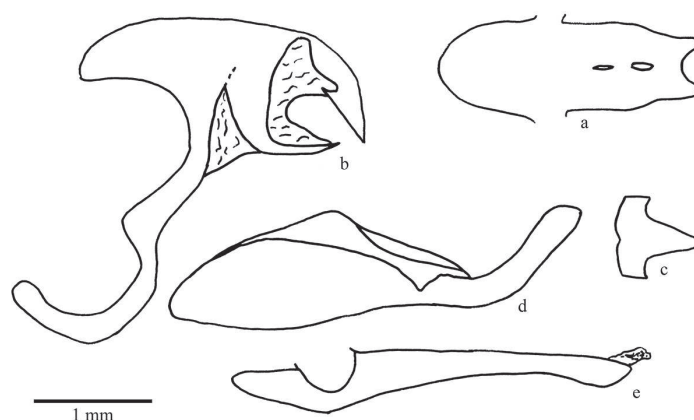
TAXONOMY – Very similar species to its congeners, only certainly separable by the male genitalia.

ETYMOLOGY – The specific name refers to the farm, Fazenda Urupá, where the holotype was collected.

REMARKS – Only known from the holotype, collected on highest flower of an *Eupatorium* bush (Asteraceae) in July during the dry period.

TYPE:

Holotype male with the following labels: / Holotypus/ 23-25-VII-1991 Faz[enda] Urupá, Banho do Rio Preto, 40 KM SE P[orto]. Velho [Candeias do Jamari], RO. [Rondônia, Brazil] C. Mielke & Miers/ gen[itália]. prep[arada]., [O.] Mielke 2018/ Holotypus *Azonax urupa* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke 2019/ BC-OM/ OM 27.118/. OM-DZUP.



Text-Fig. 3. *Azonax urupa* **sp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) gnathos, ventral view; d) right valva, internal view; e) aedeagus, lateral left view (holotype).

PASSOVINI O. Mielke, 2001

Five genera.

TAXONOMY – The tribe is characterized by the genome (Zhang *et al.* 2019) and by some morphological characters: gnathos as a transverse band, with or without a median distal projection; valva long, with the ampulla of the same size or larger than the harpe, and the ventral part of the valva (harpe), forming a distinct lobe; aedeagus longer than valva; vesica with at least one spiniform cornutus, similar in size or larger than the opening of the aedeagus, or, if smaller, than with an area of sclerotised dots; fultura inferior thin as a transversal bar and dorsally concave to support the aedeagus; sterigma laminate, with the distal point bifid; lamella postvaginalis nearly triangular.

Agara Mabilie & Boulet, 1908

Ten species.

DISTRIBUTION – Mexico to southern Brazil, Paraguay and northeastern Argentina.

REMARKS – Some species included in *Agara* were formerly included in *Myscelus* Hübner.

Agara gallardi O. Mielke, Brockmann & C. Mielke, **sp. n.**

Pl. 1, Figs. 7 (holotype ♂), 8 (♀ allotype) (d, v); Text-Fig. 4 (♂ genitalia)

DISTRIBUTION – French Guiana.

French Guiana: Cayenne (*Kourou*); Saint-Laurent-du-Maroni (*Saül – Montagne Boeuf Mort, Point de Vue du Belvédère*).

DESCRIPTION – Male: head dorsally with transverse yellow lines at the frons, before and between the antennae, on the posterior portion of the vertex, and on the collar. Thorax dorsally and tegulae dark yellow to rusty. Abdomen dorsally black with the base and distal margins of each segment yellow, mesally darker. Ventrally, labial palpi light yellow; genae, thorax, legs partially, basal portion of the abdomen and distal portions of the abdominal segments yellow, as the wing base.

Forewing length 19–21 mm. The two available specimens show colour variation; while the ground colour of the holotype is blackish-brown and the dorsal sides of the wings are ornamented with light yellow drawings, at the paratype the ground colour is rusty-brown and the dorsal sides of the wings are ornamented with rusty-yellow drawings. Forewing dorsally blackish-brown to rusty-brown, with markings yellow or rusty to various degrees, and hyaline spots. Hyaline spot in the discal cell subsquared, slightly yellowish, anterior margin slightly smaller than the inferior margin. Hyaline spots in the spaces between R_3 to $2A$; in R_3 – R_4 and R_4 – R_5 oval and the smallest, in R_5 – M_1 subrectangular, in M_1 – M_2 deeply concave distally (these four spots white), in M_2 – M_3 narrower at M_2 than at M_3 , in M_3 – CuA_1 subrectangular, in CuA_1 – CuA_2 similar but double sized and with the basal 1/3 contiguous to the cell spot, and in CuA_2 – $2A$ subtriangular with the basal margin concave and the distal margin deeply invaginated in its posterior half (these last four yellowish). Light yellow stripes at the costal base, in the subbasal area between the costal margin and the discal cell, in the costal margin two overlapped above to the hyaline cell spot, the superior slightly longer, and an elongated or diffuse at the superior and distal angle in the discal cell; a yellow patch, slightly darker than all the formers, in CuA_2 – $2A$ from the basal area to the hyaline spot, and a similar and diffuse patch at the submarginal area from M_3 (weaker) to tornus; central area of $2A$ to the anal margin darker, almost rusty. Fringes dark brown.

Hindwing with crenulated external margin, expanded in Rs – M_1 , CuA_2 , and $2A$. Dorsally dark rusty-brown or blackish with dark yellow or rusty areas. Rounded hyaline spot at the distal end of the discal cell. Costa to $Sc+Rs$ with dark yellow rusty scales throughout; rusty-brown to blackish on the 2/3 of the basal area between $Sc+R_1$ – Rs , the basal half of the discal cell, and central spots between Rs and $2A$, in Rs – M_1 elongated and transverse, in M_1 – M_3 subsquared and contiguous to the discocellular vein, in CuA_1 – CuA_2 elongated and in CuA_2 – $2A$ trapezoidal and in the narrow marginal area; light yellow at the base of Rs – M_1 and $2A$ to anal margin. Fringes whitish with the tip of the veins dark brown.

Forewing ventrally rusty-brown to black, lighter at the marginal area and with yellow spots. Basal area yellow with rusty-brown or black spots at the base and distally between Sc – CuA_1 ; two yellow stripes above the hyaline cellular spot, the anterior longer; discocellular veins weakly yellow; distally to the hyaline spots a diffuse and light yellow band between R_4 – CuA_2 and a larger and subsquared yellow spot between CuA_2 – $2A$; internal margin greyish-yellow almost to tornus. Fringes dark grey.

Hindwing ventrally with the basal area yellow and marginal area rusty-brown to blackish. Basal area with a dark brown band from the humeral angle to $2A$, a second central band between costal margin and $2A$, reaching the hyaline cellular spot proximally, and a yellow band of sparse scales from Rs to the upper half of $2A$ – $3A$ in the marginal area. Fringes whitish with the tips of the veins brownish black.

Genitalia: Typical of *Agara*. Valva with elongated and rectangular harpe, surpassing the ampulla level of about 1/4 of its length; aedeagus with subterminal and ventral spine; cornutus 1/3 of the aedeagus length.

Female: similar to the male. Forewing length 23 mm. Dorsally darker yellow and the costal spots reduced, the basal only present at the costal margin, the above of the discal cell of the same size, and no spot at the superior and distal angle in the discal cell. Fringes not differentiated.

Hindwing with crenulated margin and projected in M_3 – CuA_2 . Dorsally with discal blackish-brown spots wider and reaching the central area of $2A$ – $3A$, expanding up to the anal angle. Fringes as in the forewing.

Forewing ventrally with the yellow spots above the hyaline discal

spot of the same size and no yellow spot at the superior and distal angle in the discal cell. Fringes as dorsally.

Hindwing ventrally as in the male. Fringes as dorsally.

TAXONOMY – Species easily distinguished from all other species of *Agara* by the large hyaline spots on the forewing. The male genitalia are distinguished by the unique shape of the harpe, rectangular and elongated, and by the aedeagus. The latter is similar to that found in *A. belti* and *A. pardalina* by the presence of a subterminal and ventral spine. The DNA barcode corroborates the allocation of this species in *Agara*.

ETYMOLOGY – The specific name refers to our friend Jean-Yves Gallard, who we are grateful for the discovery of such beautiful species.

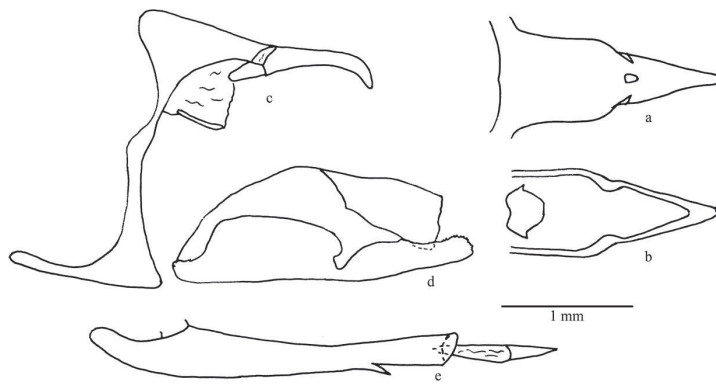
REMARKS – Known from five specimens.

TYPES:

Holotype male with the following labels: / Holotypus/ 425/ [French Guiana, Saint Laurent-du-Maroni] Saül Point de Vue [de Belvédère] 27.XII.2005, C. Faynel leg/ DNA sample ID: NVG-18087G01 c/o Nick V. Grishin/ Hermier n° 22567/ Holotypus *Agara gallardi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/. MSFR.

Allotype female with the following labels: / Allotypus/ .161. Montagne Boeuf Mort [Saül, Saint-Laurent-du-Maroni, French Guiana] 3° 38'30" N 53° 13'30" W 2.XI.2000 J.-Y. Gallard [leg.]/ Hermier n° 18666/ Allotypus *Agara gallardi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/. MSFR.

Paratypes: 1 ♂, Montagne Boeuf Mort, Saül, Saint-Laurent-du-Maroni, French Guiana, 3° 38'30" N 53° 13'30" W, 24-XI-2001, Hermier n° 19.966/ (BH); 1 ♂, Point de Vue du Belvédère, Saül, Saint-Laurent-du-Maroni, French Guiana, 16-VIII-2012, Fernandez leg., n° 283 (BH); 1 ♂, Montagne des Singes, Kourou, Cayenne, French Guiana, 24-XI-2001, Gallard leg., 5°4'30" N, 52°43'30" W, n° 19.966 (BH).



Text-Fig. 4. *Agara gallardi* sp. n. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen (part), uncus and gnathos, ventral view; c) tegumen, saccus and uncus, lateral left view; d) right valva, internal view; e) aedeagus, left view (Paratype n° 19.966).

Agara michaeli (Nicolay, 1975)

Pl. 1, Figs. 9 (♂), 10 (♀) (d, v); Pls. 4–5, Figs. 4 (♂) (d, v)

DISTRIBUTION – Southern Mexico to central Colombia.

Mexico: Chiapas (*Boca de Chajul, Bonampak*); Oaxaca (*Río Lacartún, Río Sarabia*). **Guatemala:** Izabal (*Cayuga*). **Costa Rica:** Alajuela (*Dos Ríos*); Guanacaste (*Área de Conservación Guanacaste, Carillo*); Puntarenas (*Zona protectora – Las Tablas*). **Panama:** Chiriquí; Colón (*Gamboá, Gatun, Piña, Santa Rita Ridge*); Panamá (*Bayano, El Llano, Madden Forest, Parque Nacional Soberanía*). **Colombia:** Boyacá (*Muzo*).

TYPE:

Myrcelus assaricus michaeli Nicolay, 1975. Holotype male with the following labels: / *Myrcelus assaricus* ssp. *michaeli* S. S. Nicolay Holotype ♂/ Madden Forest Panama, C. Z. [Panama] 15 July 69 G. B. Small/. AMNH.

***Agara perissodora* (Dyar, 1914)**

Pl. 1, Figs. 11 (♀) (d, v); Pls. 2–3, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Central-southern Mexico to western Ecuador and northwestern Venezuela.

Mexico: Chiapas; Guerrero (*Acahuizotla*); Oaxaca; Puebla (*Kolijke, Zihuatéutla*); Veracruz (*Catemaco, El Bastonal, Los Tuxtlas, Misantla, Xalapa*). **El Salvador** (?). **Nicaragua.** **Costa Rica:** Alajuela (*San Ramón*); Cartago (*Turrialba*); Guanacaste (*Área de Conservación Guanacaste*); Heredia (*Puerto Viejo*); Puntarenas (*San Vito*); San José (*Parque Braulio Carillo*). **Panama:** Bocas del Toro (*Chiriquí Grande*); Chiriquí (*Santa Clara*); Coclé (*El Cope*); Panama (*Panamá*); Panamá Oeste (*Cerro Campana*). **Colombia:** Boyacá (*Otanche*); Cauca (*Popayán*); Chocó (*Quebrada San Antonio*); Córdoba (*Montezuma*); Magdalena (*Sierra Nevada de Santa Marta*); Santander (*Contratación*); Tolima (*Mato Viejo*); Valle del Cauca (*Buga, Cali, Calima, Buenaventura – Juntas, Matagang, Montezuma, Río Anchicaya*). **Ecuador:** Esmeraldas (*Bocana, Chuchuvi*); Pichincha (*Pacto*). **Venezuela:** Táchira (*Chorro del Índio, Río Frío, San Cristóbal*).

REMARKS – About taxonomic issues with this species and *A. pegasus*, look under *A. pegasus*.

TYPE:

Myscelus perissodora Dyar, 1914. The original description mentions an unspecified number of specimens from Misantla, [Veracruz], Mexico, June 1912, R. Müller leg, nr. 15454 in USNM. To ensure the correct identification of *Myscelus perissodora*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type No. 15454 U.S.N.M./ Misantla [Veracruz] Mexico/ June 1912/ RMuller Collector/ 3596/ *Myscelus perissodora* Dyar Type/ Genitalia NO. X-49.24/ J.M.Burns 2000/; two other labels will be added: / Lectotypus/ Lectotypus *Myscelus perissodora* Dyar, 1914, O. Mielke, Brockmann & C. Mielke det. 2019/. USNM.

***Agara belti* (Godman & Salvin, 1879)**

Two subspecies.

DISTRIBUTION – Eastern Mexico to western Ecuador and northern Venezuela.

***Agara belti belti* (Godman & Salvin, 1879)**

Pls. 2–3, Figs. 1 (♂), 2–4 (♀) (d, v)

DISTRIBUTION – Eastern Mexico to western Ecuador and northern Venezuela.

Mexico: Chiapas (*Pichucalco*); Oaxaca (*Río Sarabia*); Veracruz (*Catemaco, El Bastonal, Los Tuxtlas, Montepío, Motzorongo, Presidio, Santa Rosa*). **Guatemala:** Alta Verapaz (*Poloichic Valley*); El Petén (*Parque Nacional Tikal*); Escuintla (*Palin*); Guatemala (*Pantaleón*); Izabal (*Morales – Cayuga*). **Honduras:** Atlántida (*La Ceiba*). **El Salvador:** San Salvador (*Cerro El Boquerón*). **Nicaragua:** Chontales; Jinotega. **Costa Rica:** Alajuela (*San Mateo*); Guanacaste (*Área de Conservación Guanacaste*); Heredia; Limón (*Guapiles*); Puntarenas (*Parque Nacional Corcovado*); San José (*Colón*); Cartago (*Cachí*). **Panama:** “Canal Zone”; Chiriquí; Panamá (*Madden Forest*). **Colombia:** Amazonas (*Letícia*); Boyacá (*Muzo, Otanche*); Meta (*Villavicencio*). **Ecuador:** Bolívar (*Balzapamba*); Esmeraldas (*Durango*). **Venezuela:** Miranda (*Guatopo*); Táchira (*Río Frío*); Trujillo (*Campo Elías*). **Peru:** Amazonas (*Rodríguez de Mendoza*).

REMARKS – The female specimen illustrated on Pls. 2–3, Figs. 4 may be aberrant, or a new subspecies of *A. belti*. There is no hyaline spot on the disc of the hindwing. Only this female specimen from Rodríguez de Mendoza, Amazonas, Peru (EB) was examined. The DNA barcode minimum divergence from *A. belti* (n= 9) is 1%.

TYPE:

Myscelus belti Godman & Salvin, 1879. The original description mentions an unspecified number of males from Chontales, Nicaragua, and Poloichic Valley, Guatemala in the Godman & Salvin collection. To ensure the correct identification of *Myscelus belti*, one syntype male is **herein designated as the lectotype**; this specimen with the following

labels: / Type – H.88/ Chontales, Nicaragua, Janson./ Sp. figured/ not found in Hws. Coll. 1878/ 303/ B.C.A.Lep.Rhop. *Myscelus belti*, G.&S./ Godman-Salvin Coll. 1912.-23/ NHMUK(E)# 1054254; two other labels will be added: / Lectotypus/ Lectotypus *Myscelus belti* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke leg./ NHMUK.

***Agara belti caucaensis* (Constantino, Salazar & Vargas, 2013)**

Pl. 1, Figs. 12 (holotype ♂) (d, v)

DISTRIBUTION – Western Colombia.

Colombia: Caldas (*Chinchiná – Reserva Florestal Protectora de Planalto*).

REMARKS – Only known from the holotype.

TYPE:

Myscelus belti caucaensis Constantino, Salazar & Vargas, 2013. Holotype male from Colombia, Caldas, Chinchiná, Reserva Florestal Protectora de Planalto, Cenicafé. CFC.

***Agara pegasus* (Mabille, 1903)**

Pls. 2–3, Figs. 7 (♂), 8 (♀) (d, v)

DISTRIBUTION – Western, northern and eastern Colombia, eastern Ecuador to northern Venezuela and French Guiana.

Colombia: Meta (*Upper Río Negro*); Santander (*Ocana*); Valle del Cauca (*Cali*). **Ecuador:** Zamora-Chinipe (*Zamora*). **Venezuela:** Azoátegui (*Valle de Guanape*); Aragua (*Choroní, Parque Nacional Henry Pittier – Rancho Grande*); Barinas (*Barinitas, La Chiménca*); Mérida (*Chorreras de las González, Via La Chorrera*); Táchira (*Chorro del Índio, Delicias, San Cristóbal*); Zulia (*Sierra de Perijá*). **French Guiana:** Cayenne (*Cayenne*).

TAXONOMY – The taxonomic status of this and *Agara perissodora* is questionable. The only known syntype of *Myscelus perissodora* is from Mexico: Veracruz (*Misantla*) and has the outer margin of the ventral hindwing very similar to that of *A. pegasus*: the discal band is not separated from the dark margin. Specimens of *A. perissodora* from Costa Rica have the outer margin of the ventral hindwing with the discal brown band separated from the outer dark margin. The genitalia of both taxa are similar. We have only examined a few specimens in collections and few localities are known for either species (misidentifications are possible).

TYPE:

Myscelus pegasus Mabille, 1903. The original description mentions an unspecified number of specimens from Cayenne, [French] Guiana. To ensure the correct identification of *Myscelus pegasus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *Myscelus pegasus* Mab. g. ins. Guyane [Française]/ *M. pegasus* Mab./ Ex musaeo P. Mabille 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Myscelus pegasus* Mabille, 1903, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Agara epimachia* (Herrich-Schäffer, 1869)**

Five subspecies.

DISTRIBUTION – Northern and western Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, northwestern Venezuela, French Guiana, northern, western and southern Brazil, Paraguay and northeastern Argentina.

***Agara epimachia epimachia* (Herrich-Schäffer, 1869)**

Pls. 2–3, Figs. 11 (♂), 12 (neotype ♂ of *Myscelus epimachia*) (d, v); Pls. 74–75, Figs. 5 (♀) (d, v)

DISTRIBUTION – Western Colombia to western Ecuador,

eastern Colombia to central-eastern Bolivia, northwestern Venezuela and northern Brazil.

Colombia: Caquetá (*Florencia*); Central Cordillera; Cundinamarca (*Bogotá*); Meta (*Villavicencio*); Occidental Cordillera; Valle del Cauca. **Ecuador:** Cotopaxi (*Río Mulato*); Guayas (*Palmar*); Morona-Santiago (*San Isidro, Santiago de Méndez*); Napo (*Puerto Napo, Tena*); Pastaza (*Mera*); Tungurahua (*Topo*). **Peru:** Amazonas (*Chachapoyas, Quebrada Chingaza*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo, La Merced, Perené, Río Colorado, San Ramón, Satipo, Shimá*); Loreto (*Iquitos, Upper Río Napo*); Madre de Dios (*Parque Nacional del Manu, Shintuya – Cerro Pontiacollo, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Huancabamba, Pozuzo*); Puno (*Yahuar Mayo*); San Martín (*Juanjuí, Moyobamba, Rioja, Roque, Tarapoto, Tocache*). **Bolivia:** Beni (*Reyes*); Cochabamba (*Chapare*); La Paz (*Apolobamba, Caranavi, Chulumani, Farinas, Río Zongo*); Santa Cruz (*Amboro, Andrés Baez – Potrerillos de Guendá, Sara*). **Venezuela:** Aragua (*Choroní, El Limón, Parque Nacional Henry Pittier – Rancho Grande*); Azoátegui (*Las Quiguas – San Esteban Valley*); Barinas (*Barinitas*); Bolívar (*Santa Helena*); Carabobo (*Puerto Cabello*); Táchira (*Veja de Aza, San Joaquín de Navay, San Félix, San Cristóbal, Río Frío*). **Brazil:** Amazonas; Acre (*Santa Rosa do Purus*); Rondônia (*Cacaulândia*).

TYPE:

Myscelus epimachia Herrich-Schäffer, 1869. The original description mentions an unspecified number of specimens from Tropical America. No syntypes were found in MfN or ANSP, where the types of Herrich-Schäffer are deposited, or in any museum or collection mentioned above (see “Abbreviations of the collections”). To ensure the correct identification of *Myscelus epimachia*, mentioned as *Myscelus epimachia epimachia* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Rondos, Tingo Maria, [Huánuco] Peru, 850 m, 27-X-1971, Villegas leg./ Neotypus *Myscelus epimachia* Herrich-Schäffer, 1869 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.215/. DZUP.

Agara epimachia nevada O. Mielke & Casagrande, *ssp. n.*

Pls. 2–3, Figs. 9 (holotype ♂) (d, v)

DISTRIBUTION – Northern Colombia

Colombia: Magdalena (*Sierra Nevada de Santa Marta*).

DESCRIPTION – In the Sierra Nevada, Colombia, there is a different population. When compared to the nominal subspecies, the male has the hyaline spots on the forewing in spaces R_5-M_1 and M_1-M_2 larger, connecting the apical spots to the subapical spots; dark and narrower border on the dorsal hindwing; colour of the dorsal face lighter; ventrally the discal band may enter very slightly into space $Sc+R_1-R_5$, as is usual in *Agara epimachia edix*; male genitalia with the valva similar to that of *Agara epimachia verina* (see Evans 1951).

ETYMOLOGY – The specific name refers to the type locality: Sierra Nevada de Santa Marta.

REMARKS – Only two male specimens are known from the type locality.

TYPES.

Holotype male with the following labels: / Holotypus/ 1971 Sierra Nevada de Santa Marta, [Magdalena] Colombia/ Holotypus *Agara epimachia nevada* O. Mielke & Casagrande. O. Mielke & Casagrande det. 2017/ OM 61.779/. OM-DZUP.

Paratype: 1 ♂, IX-1982, Sierra Nevada de Santa Marta, [Magdalena], Colombia, 400 m, OM 18.198 (OM).

Agara epimachia verina (Evans, 1951)

Pls. 2–3, Figs. 10 (♂), 13 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador.

Ecuador: Pastaza (*Mera, Puyo NW, Sarayacu*); Tungurahua (*Ambato, Río Topo*); Zamora-Chinchipe (*Zamora*).

TYPE:

Myscelus epimachia verina Evans, 1951. Holotype male with the following labels: / Type – *verina* Evans/ Zamora, [Zamora-Chinchipe] Ecuador, 3–4000ft. (O. T. Baron)./ Rothschild Bequest B.M.1939-1./ NHMUK.

Agara epimachia edix (Evans, 1951)

Pls. 2–3, Figs. 14–16, 18, 20–21 (♂), 17, 19, 22 (♀) (d, v)

DISTRIBUTION – Western to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Mato Grosso (*Diamantino, Pantanal, Pontes e Lacerda*); Mato Grosso do Sul (*Dourados*); Goiás (*Chapada dos Veadeiros, Goiás Velho, Leopoldo Bulhões*); Minas Gerais (*Marliéria*); Espírito Santo (*Linhares*); Rio de Janeiro (*Guapimirim, Magé – Vila Inhomirim, Petrópolis – Independência, Rio de Janeiro, Teresópolis*); São Paulo (*Casabranca, Indiana*); Paraná (*Diamante do Norte, Fênix, Foz do Iguaçu, Londrina, Querência do Norte, Rolândia, São Pedro do Ivaí, Terra Boa*); Santa Catarina; Rio Grande do Sul. **Paraguay:** Alto Paraná (*Itakyr*); Canindeyú; Paraguari (*Sapucai*). **Argentina:** Misiones (*General Belgrano, Puerto Iguazú*).

TYPE:

Myscelus epimachia edix Evans, 1951. Holotype male with the following labels: / Type – *edix* Evans/ Sapucay, [Paraguari] Paraguay, Foster 7-1804/ J.J.Joicey Coll. B.M.1925-451/. NHMUK.

Agara epimachia hermieri O. Mielke, Brockmann & C. Mielke, *ssp. n.*

Pls. 74–75, Figs. 6 (holotype ♂), 7 (♀ allotype) (d, v)

DISTRIBUTION – French Guiana.

French Guiana: Cayenne (*Roura*); Saint-Laurent-du-Maroni (*Arouary River*).

Arouary [*sic*], the locality of the allotype female, was not located, but Barbut *et al.* (2012: 42) mention that this locality is the Arouary River, a tributary of the Mana River in the Saint-Laurent-du-Maroni arrondissement.

DESCRIPTION – In French Guiana there is a different population. Male: forewing length 22–24 mm ($n=3$), dorsally very similar to the darker phenotype of *A. epimachia epimachia* from Peru, including the darker surroundings of the median and subapical semi-hyaline spots on the forewing, but the hindwing differs ventrally by the narrower yellow band between the median and the submarginal dark bands, and by this band being connected to the median dark band at the costal margin by the presence of a dark spot in R_5-M_1 ; in *A. epimachia epimachia* the yellow band is wider than the submarginal dark band and this is never connected to the median dark band at the costal margin by a dark spot in R_5-M_1 . Differs from *A. epimachia edix* by the darker ochreous colour on the dorsal face of the wings and by the smaller semi-hyaline spots on the forewing, mainly the cell spot, and at the ventral hindwing by the presence of the dark spot in R_5-M_1 , connecting the submarginal dark band to the median dark band at the costal margin; this spot may be present in *A. epimachia edix* (Pl. 5, Fig. 18), but always narrower than in *A. epimachia hermieri ssp. n.*

Female: forewing length 25–27 mm ($n=2$). Similar to the male, but with more rounded wings and hindwing ventrally with the yellow band between the median dark band and the submarginal dark band narrower than this band between M_3-CuA_2 .

ETYMOLOGY – The specific name refers to our friend Bernard Hermier, who we are grateful for the discovery of such beautiful subspecies.

REMARKS – Only known from five specimens.

TYPES:

Holotype male with the following labels: / Holotypus/ 370/ D6 pk 16 4°37'30" 52°17'W [nearly 16 km south of Roura and near Fourgassier] 9.I.2005 J.-Y. Gallard [leg.] ex coll. Bernard Hermier, Montjoly, Fr. Guiana IX-2017/ Hermier n° 21851/ DNA sample ID: NVG-18087G03 c/o Nick V. Grishin/ Holotypus *Agara epimachia hermieri* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2020/. MSFR, ex-EB.

Allotype female with the following labels: / Allotypus/ Arouary

[sic], Cayena, Fr[ench]. Guiana/ Allotypus *Agara epimachia hermierei* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2020/ [OM] 11.967/. OM-DZUP.

Paratypes: 1 ♂, 13-I-2001, D6 pk7,5–pk8, 4°40'30" N, 52°18'30" W, nearly 7,5–8 km south of Roura, Cayenne, French Guiana, P. Ducaud leg., Hermier n°19200 (BH); 1 ♂, 27-II-2005, D6 Km 16,6, 4°37' N, 52°17' W, nearly 16,6 km south of Roura, Cayenne, French Guiana, J.-Y. Gallard leg., Hermier n° 21722 (BH); 1 ♀, 20-XII-2013, Route de Kaw, Roura, Cayenne, French Guiana, Fernandez n° 315, Fernandez leg. (BH).

Agara draudti (Riley, 1926)

Pls. 2–3, Figs. 23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Eastern Peru to central-eastern Bolivia.

Peru: Amazonas (*Chachapoyas – Yambrasbamba, Pedro Ruiz Gallo*); Cusco (*Callanga, Cosñipata Valley – Quebrada Santa Isabel 1,200 m to Rocotal 2,000 m, Machu Picchu, Marcapata*); Huánuco (*Carpish, Puente Durand, Tingo Maria*); Junín (*Chanchamayo, El Porvenir, La Merced, San Ramón*); Pasco (*Huancabamba, Oxapampa*); Puno (*Río Inambari*); San Martín (*Juanjuí, Rioja – El Afluente 1,200 m, Santa Cruz*). **Bolivia:** Cochabamba (*Chapare, Bajo Palmar*); La Paz (*Caranavi, Farinas, Río Chairó, San Antonio*).

TYPE:

Myscelus draudti Riley, 1926. Holotype (one male is mentioned) with the following labels: / Type H T/ Chairó, [La Paz] Bolivia. C. Buckley. Druce Coll./ Godman & Salvin Coll. 1912.-23. *Myscelus epimachia* H.-S. [sic]/. NHMUK.

Agara santhilarius (Latreille, [1824])

Pls. 4–5, Figs. 1 (♂), 2–3 (♀) (d, v)

DISTRIBUTION – Northeastern to southern Brazil.

Brazil: Bahia; Espírito Santo (*Linhares, Santa Leopoldina – Serra das Andorinhas, Santa Teresa, São Mateus*); Minas Gerais; Rio de Janeiro (*Casimiro de Abreu, Cachoeiras de Macacu, Duque de Caxias, Guapimirim, Rio de Janeiro, Teresópolis*); São Paulo (*Santos, Ubatuba*); Paraná (*Antonina, Paranaguá*); Santa Catarina (*Camboriú, Corupá, Itajaí, Joinville, Massaranduba, São Francisco do Sul*).

The citations **Peru:** San Martín (Bryk 1953), **Bolivia** (Ureta 1941), **French Guiana** (Mabille 1903, Mabille & Boullet 1908, Mabille 1912, Draudt 1921, Spitz 1932, Bryk 1953, Lewis 1973, 1975), and **Brazil:** Amazonas (Perty 1833, Draudt 1921, Spitz 1932, Bryk 1953, Lewis 1973, 1975) are probably incorrect.

TAXONOMY – The DNA barcode is very similar to that of *A. assaricus* (divergence of 0.8% to 1.1% between specimens of *A. santhilarius* (n= 1) and specimens of *A. assaricus mapirica* (n= 5); these two taxa are maintained as species by morphological characters – the male genitalia are constantly different (see Evans 1951).

REMARKS – Inhabitant of lowlands along the coast.

TYPE:

Hesperia santhilarius Latreille, [1824]. The original description mentions an unspecified number of specimens from Brazil, Langsdorff leg. in collection Latreille. To ensure the correct identification of *Hesperia santhilarius*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *H. Santhilarius* Lat. Bresil/ Ex musaeo P. Mabille 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Hesperia santhilarius* Latreille, [1824], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Agara assaricus (Cramer, 1779)

Two subspecies.

DISTRIBUTION – Southeastern Colombia to central-eastern Bolivia and northern Venezuela to French Guiana and northern and northwestern Brazil.

Agara assaricus assaricus (Cramer, 1779)

Pls. 4–5, Figs. 5, 8 (♂), 6–7 (♀) (d, v)

Syn.: *Papilio janthibar* Butler, [1870], **nom. nud.** (ICZN, Art. 11.6)

DISTRIBUTION – Eastern Ecuador to southeastern Peru, northeastern Venezuela to French Guiana and northern Brazil.

Ecuador: Pastaza (*Mera*). **Peru:** Madre de Dios. **Venezuela:** Delta Amacuro (*Río Acure*). **Suriname.** **French Guiana** (*Cayenne*). **Brazil:** Roraima (*Alto Alegre*); Amazonas (*Maués, Tefé*); Pará (*Capim, Itaituba to Óbidos, Santo Antonio do Tauá*); Rondônia (*Cacaulândia*); Mato Grosso (*Alta Floresta*).

TYPES:

Papilio assaricus Cramer, 1779. The original description mentions an unspecified number of specimens from Suriname in the collection W. van der Meulen. To ensure the correct identification of *Papilio assaricus*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Calkoen Surin. [Suriname]/ Museum Leiden/ ♀/ Museum Leiden *Myscelus* cf. *epimachia* H-S det V. R. Altena '41/; two other labels will be added: / Lectotypus/ Lectotypus *Papilio assaricus* Cramer, 1779, O. Mielke, Brockmann & C. Mielke det. 2019/. NBC. Another syntype is herein designated as paralectotype; this specimen with the following labels: / Coll. Calkoen/ Museum Leiden/ ♀/ Museum Leiden *Myscelus* cf. *epimachia* H-S det V. R. Altena '41/; two other labels will be added: / Paralectotypus/ Paralectotypus *Papilio assaricus* Cramer, 1779, O. Mielke, Brockmann & C. Mielke det. 2019/. NBC.

Papilio janthibar Butler, [1870]. The name was published as a synonym of *Myscelus assaricus*, a **nomen nudum**.

Agara assaricus mapirica Strand, 1921

Pls. 4–5, Figs. 9–10, 12–14 (♂), 11 (♀) (d, v)

DISTRIBUTION – Southeastern Colombia to central-eastern Bolivia and northern Venezuela to northwestern Brazil (Acre).

Colombia: Vaupés (*Taraira – Estación Caparú*). **Peru:** Huánuco (*Tingo Maria*); Junín (*La Merced, Río Colorado, Satipo, Shimá*); Loreto (*Iquitos, Nauta*); Madre de Dios (*Parque Nacional Bahuaia Sonene, Parque Nacional del Manu, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Puno (*Sandia*); San Martín (*Chazuta, Juanjuí, Tarapoto*). **Bolivia:** La Paz (*Caranavi, Mapiri, Río Zongo, San Ernesto*); Santa Cruz (*Andrés Ibañez, Buenavista*). **Venezuela:** Sucre (*Finca Vuelta Larga*). **Brazil:** Acre (*Bujari, Porto Acre, Senador Guiomard*).

TYPE:

Agara pardalina var. *mapirica* Strand, 1921. Holotype (one male is mentioned) with the following labels: / Typus/ ♂/ Boliv[ia]., [La Paz] S. Ernesto bis Mapiri III/ Coll. W. Schnuse/ *Agara pardalina* Feld. v. *mapirica* m., Strand det./ *pardalina* Feld. V. *mapirica* Strand/. DEI.

Agara pardalina (C. Felder & R. Felder, 1867)

Four subspecies.

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia and Guyana, northern and southern Brazil, Paraguay and northeastern Argentina.

Agara pardalina pardalina (C. Felder & R. Felder, 1867)

Pls. 4–5, Figs. 15, 19–20 (♂), 16 (♀) (d, v)

Syn.: *Agara aurora* Röber, 1925

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia and Guyana to northern Brazil (Pará).

Colombia: Casanare (*Aguazul*); Cundinamarca (*Bogotá*); Meta (*Acacias, Río Negro, Villavicencio*). **Ecuador:** Morona-Santiago (*Macas*);

Napo (*Misahualli, Tena*); Sucumbíos (*Cerro Lumbaqui Norte*). **Peru:** Amazonas (*Quebrada Chingaza*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to Quebrada Santa Isabel 1,200 m*); Junín (*Chanchamayo, El Porvenir, La Merced, San Ramón, Satipo, Shimá*); Madre de Dios (*Río Madre de Dios, Shintuya, Tambopata Reserve*); San Martín (*Tarapoto*). **Bolivia:** Beni (*Beni*); La Paz (*Caranavi, Iténez, Iturrealde, Río Zongo*); Santa Cruz (*Andrés Ibañez – Potrerillos del Guendá, Buenavista, Velasco*). **Guyana. Brazil:** Amazonas (*Tefé*); Acre (*Porto Acre*); Rondônia (*Cacaulândia*).

The citation **Brazil:** Pará (Beccaloni *et al.* 2008) is incorrect; it is *M. pardalina guarea*.

TYPES:

Tamyris pardalina C. Felder & R. Felder, 1867. The original description mentions an unspecified number of males from Bogotá, Nova Granada [Colombia], Lindig leg. in collection Felder. To ensure the correct identification of *Tamyris pardalina*, on syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Bogota [Cundinamarca, Colombia] Lindig type/ Felder Collⁿ. / *pardalina*. n./ Rothschild Bequest 1939-1./; two other labels will be added: / Lectotypus/ Lectotypus *Tamyris pardalina* C. Felder & R. Felder, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Agara aurora Röber, 1925. Holotype (one male [*recte* female] is mentioned) with the following labels: / Holotype female *Agara aurora* Röber, 1925/ Macas- [Morona-Santiago] Ecuador 1905-06/ Staatl. Museum für Tierkunde Dresden/ Stauding. & Bang-Haas Dresden. Ankauf 1961/. SMT.

***Agara pardalina kesselringi* (O. Mielke & Casagrande, 2011)**

Pls. 4–5, Figs. 17 (paratype ♂), 18 (holotype ♂) (d, v)

DISTRIBUTION – Central Peru.

Peru: Huánuco (*Tingo Maria*).

REMARKS – Only known from the type locality.

TYPE:

Myscelus pardalina kesselringi O. Mielke & Casagrande, 2011. Holotype male with the following labels: / Holotypus/ 26-III-1966 Tingo Maria, [Huánuco], Peru/ Holotypus *Myscelus pardalina kesselringi* [O.] Mielke & Casagrande, [O.] Mielke & Casagrande det. 2010/ [OM]10.822/. OM-DZUP.

***Agara pardalina guarea* (Evans, 1951)**

Pls. 4–5, Figs. 21 (holotype ♂), 22 (paratype ♀) (d, v)

DISTRIBUTION – Northern Brazil (northeastern Pará).

Brazil: Pará (*Belém*).

The citation **Colombia:** Amazonian area (Andrade-C. *et al.* 2015) is probably incorrect.

REMARKS – Only known from the type locality.

TYPE:

Myscelus pardalina guarea Evans, 1951. Holotype male with the following labels: / Type – *guarea* Evans/ Pará [Brazil] A. M. Moss *Myscelus* sp. nov “Guarea”/ Miles Moss Coll. B.M.1947-453/ NHMUK(E)#1054054/. NHMUK.

***Agara pardalina yacutinga* (O. Mielke & Casagrande, 2011)**

Pls. 4–5, Figs. 23 (holotype ♂), 24 (allotype ♀) (d, v)

DISTRIBUTION – Southern Brazil (western Paraná), Paraguay and northeastern Argentina.

Brazil: Paraná (*Diamante do Norte, Foz do Iguaçu, São Pedro do Ivaí*). **Paraguay:** Guairá (*Villarica – Mbuevo*). **Argentina:** Misiones (*General Belgrano*).

TYPE:

Myscelus pardalina yacutinga O. Mielke & Casagrande, 2011. Holotype male with the following labels: / Holotypus/ 2-5-III-2007. Reserva Yacutinga, Almirante Brown, Gal. Belgrano, Misiones,

Argentina, [O.] Mielke & Casagrande leg./ *Myscelus pardalina yacutinga* [O.] Mielke & Casagrande, [O.] Mielke & Casagrande det. 2010/ DZ 16.725/. DZUP, transferred to IML.

***Myscelus* Hübner, [1819]**

Five species.

DISTRIBUTION – Mexico to southern Brazil and northern Argentina.

***Myscelus nobilis* (Cramer, 1777)**

Pls. 6–7, Figs. 1 (♂), 2 (♀) (d, v); Text-Fig. 5 (♂ genitalia)

Syn.: *Papilio salus* Fabricius, 1781; **nom. n.**

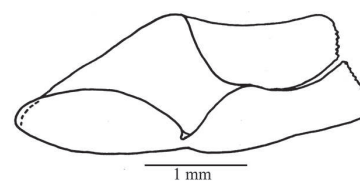
DISTRIBUTION – Eastern Ecuador to southeastern Peru, Venezuela to Suriname and northern Brazil.

Ecuador: Napo (*Archidona – Llaucana*). **Peru:** Huánuco (*Cucharas, Tingo Maria*); Junín (*Satipo, Shimá*); Loreto (*Contamana, Iquitos, Mishana, Nauta, Pebas, Pantoja, Puerto San Mateos, Tarapacá*); Madre de Dios (*Parque Nacional Bahuaja, Parque Nacional del Manu, Puerto Maldonado, Tambopata Reserve*); San Martín (*Juanjuí, Moyobamba, Tarapoto*). **Venezuela:** Aragua (*Choroní, La Victoria*); Bolívar (*El Dorado*). **Suriname. Brazil:** Amazonas (*Benjamin Constant, Lower Rio Purus, Manaus, Tefé*); Pará (*Óbidos*); Acre (*Porto Acre, Xapuri*); Rondônia (*Cacaulândia*); Mato Grosso (*Vera*).

TYPES:

Papilio nobilis Cramer, 1777. The original description mentions an unspecified number of males in the collection of Mr. L. Juliaans and collection J. C. Sylvius van Lennep. To ensure the correct identification of *Papilio nobilis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Surinam Cu. Leñep/ Felder Collⁿ. / Rothschild Bequest B.M.1939-1/ *Papilio nobilis* Cramer. Syntype. O. Mielke det. 1988/; two other labels will be added: / Lectotypus/ Lectotypus *Papilio nobilis* Cramer, 1777, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Papilio salus Fabricius, 1781 is a **nomen novum** for *Papilio nobilis* Cramer, 1777.



Text-Fig. 5. *Myscelus nobilis*. Male genitalia: right valva, internal view (Peru, Loreto, Río Amazonas, Iquitos, Picuroyacu, X-2009, J. Hernández leg., DZUP 21.067).

***Myscelus illustris* Mabille, 1903, stat. rev.**

Pls. 6–7, Figs. 3 (♂) (d, v); Text-Fig. 6 (♂ genitalia)

DISTRIBUTION – Peru to central-eastern Bolivia and central-western Brazil (Mato Grosso do Sul).

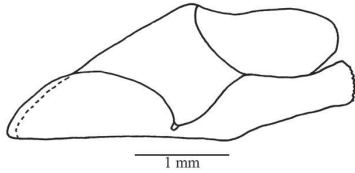
Peru. Bolivia: Beni (*Río Beni, Rurrenabaque, Trinidad*); Cochabamba (*Chapare, San Mateo*); La Paz (*Caranavi, Muchanes*); Santa Cruz (*Buenavista, Santa Cruz de la Sierra, Sara*). **Brazil:** Mato Grosso do Sul (*Nioaque*).

TAXONOMY – The species regarded as a synonym of *Myscelus nobilis* by Evans (1951) and by subsequent authors, is here revalidated to full species status.

The colour of the dorsal wings is brighter yellow, the outer part of the veins CuA₂, 2A and the line between on the forewing are very faintly darkened, and all other dark lines are narrower, including the marginal line; male genitalia with a rounder ampulla and a finer harpe, and both more separated than in *Myscelus nobilis*.

TYPE:

Myscelus illustris Mabilé, 1903. The original description mentions an unspecified number of specimens from Bolivia. To ensure the correct identification of *Myscelus illustris*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H.86/ Bolivia/ Ex musaeo P. Mabilé 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Myscelus illustris* Mabilé, 1903, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.



Text-Fig. 6. *Myscelus illustris* **stat. rev.** Male genitalia: right valva, internal view (Bolivia, Cochabamba, Chapare, 400 m, V-1948, DZUP 21.074).

Myscelus amystis (Hewitson, 1867)

Five subspecies.

DISTRIBUTION – Western and eastern Mexico to western Ecuador, northern, central and eastern Colombia to central-eastern Bolivia, Venezuela, Trinidad and Tobago and northwestern Brazil.

Myscelus amystis amystis (Hewitson, 1867)

Pls. 6–7, Figs. 11 (neotype ♂ of *Myscelus flavicollis*), 12 (♀), 13 (♂) (d, v)

Syn.: *Myscelus rogersi* Kaye, 1914

Myscelus flavicollis Röber, 1925

DISTRIBUTION – Northern, central and eastern Colombia, Venezuela and Trinidad and Tobago.

Colombia: Cesar (*Lake Sapatosa*); Cundinamarca (*Bogotá*); Cordillera Occidental; Magdalena (*Sierra Nevada de Santa Marta*); Meta (*Remolino, Upper Río Negro*). **Trinidad and Tobago:** Port of Spain; Penal-Debe (*Morne Diable, Rock Peñal Road*); Siparia. **Venezuela:** Aragua (*Choroní, Parque Nacional Henry Pittier, La Victoria*); Barinas (*Reserva Florestal Caparo*); Bolívar (*El Barroso, Río Caura, Suapure*); Distrito Capital (*Caracas, La Guaira*); Táchira (*Río Frío, San Cristóbal*); Zulia (*El Tucano, Kasmara, Lagunillas, Río Laja*).

TYPES:

Erycides amystis Hewitson, 1867. The original description mentions an unspecified number of specimens from New Grenada [Colombia] in collection Hewitson. To ensure the correct identification of *Erycides amystis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ N. Granada [Colombia] Hewitson Coll. 79.69. *Myscelus amystis*. 3./; two other labels will be added: / Lectotypus/ Lectotypus *Erycides amystis* Hewitson, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Myscelus rogersi Kaye, 1914. The original description mentions an unspecified number of specimens from Jan. 22, 1913, N. of Quenam Bay, Siparia, Trinidad, K. St. A. Rogers leg. To ensure the correct identification of *Myscelus rogersi*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Siparia 22/1/13/ ex drawer 6/ *Myscelus rogersi* Kaye Type/ 1913 1044/ *Myscelus* sp. t. W. J. Kaye 16-17.v.1913 in O. U. M./ Jan. 22/1913 N. of Quenam B., Trinidad, Siparia: c 200, S. Mtn. Range: prob. in young forest plant'n in high forest. St. A. Rogers/ Type. Kaye. *Myscelus rogersi*. Tr. Ent. Soc. L., 1913, p. 582./ Type Lep: 3478 *Myscelus rogersi* Kaye Hope Dept. Oxford/; two other labels two will be added: / Lectotypus/ Lectotypus *Myscelus rogersi* Kaye, 1914, O. Mielke, Brockmann & C. Mielke det. 2019/. OUMNH.

Myscelus flavicollis Röber, 1925. Holotype (one female is mentioned) without data. The holotype was not found in SMT or ZfB, where Röber's types are deposited, or in any museum or collection

mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Myscelus flavicollis*, mentioned as a synonym of *Myscelus amystis amystis* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ 27-VI-1966 Kasmara, Zulia, Venez[uela]. 250 m, Lichy leg./ Neotypus *Myscelus flavicollis* Röber, 1925 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 40.273/. OM-DZUP.

Myscelus amystis hages Godman & Salvin, 1893

Pls. 6–7, Figs. 4, 7–9 (♂), 5–6 (♀) (d, v)

DISTRIBUTION – Western and eastern Mexico to Panama.

Mexico: Campeche (*Calakmul*); Chiapas (*Boca de Chajul, Escuintla, Motozintla, Ocosingo – San Quentin, San Jerónimo, Yaxchilán*); Colima (*La Salada*); Guerrero (*Acahuizotla, Mezcala, Tierra Colorada*); Hidalgo (*Tehuacán*); Jalisco (*Boca de Tomatlán*); Michoacán (*Coahuayana*); Oaxaca (*Candelaria Loxicha, Chiltepec, Tuxtepec*); Quintana Roo (*Noh-bek, Nuevo Xcán*); Puebla (*Zihuatéutla*); San Luis Potosí (*El Naranjo, El Salto, Huasteca Potosina, Rascón, Tamazunchale, Valles*); Tamaulipas; Veracruz (*Catemaco, Dos Amates, El Tajín, Fortín, Los Tuxtlas, Misantla, Presidio, Rinconada, San Lorenzo, Xalapa*). **Guatemala:** Baja Verapaz (*San Jerónimo*); El Petén (*La Libertad, Parque Nacional Tikal*). **Honduras:** Cortés (*San Pedro Sula*). **El Salvador:** La Libertad (*Los Chorrros*). **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste*); San José (*Ciudad Colón*). **Panama:** Chiriquí (*David*); Colón (*Gatun, Lion Hill, Piña*); Panamá (*Bayano, Madden Forest, Paraíso*); Veraguas (*Ballena*).

TYPE:

Myscelus hages Godman & Salvin, 1893. The original description mentions an unspecified number of males and females from Tierra Colorada de Guerrero, S. Lorenzo near Córdoba e Rinconada, Mexico; San Geronimo [*sic*], Guatemala; David, Chiriquí, Panama, and Peru. To ensure the correct identification of *Myscelus hages*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ David Panama. Champion/ Sp. figured/ B.C.A.Lep. Rhop. *Myscelus hages* G. & S./ Godman & Salvin Coll. 1912. – 23./; two other labels two will be added: / Lectotypus/ Lectotypus *Myscelus hages* Godman & Salvin, 1893, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Myscelus amystis muttra Evans, 1951

Pls. 6–7, Figs. 10 (♂) (d, v)

DISTRIBUTION – Central to southwestern Colombia and western Ecuador.

Colombia: Boyacá (*Otanche*); Valle del Cauca (*Ciqueros, Buenaventura – Juntas and Río Dagua*); Nariño (*Caura*). **Ecuador:** El Oro (*Arenillas*); Esmeraldas (*Chuchuvi, Río Durango, San Lorenzo – Tunda Loma*); Pichincha (*Pacto*); Manabí (*Jama*).

TYPE:

Myscelus amystis muttra Evans, 1951. Holotype male with the following labels: / Type – *muttra* Evans/ N^{elle} Grenade [Colombia] Cauca [Valle del Cauca] Juntas M. De Mathan fin 1897 I-1898/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Myscelus amystis distinctus Röber, 1925

Pls. 6–7, Figs. 14 (neotype ♂) (d, v)

DISTRIBUTION – Eastern Ecuador.

Ecuador: Morona-Santiago (*Macas*); Napo (*Yanahurco*); Pastaza (*Mera*); Tungurahua (*Río Blanco*).

TYPE:

Myscelus nobilis distinctus Röber, 1925. Holotype (one specimen is mentioned) from Macas, [Morona-Santiago], Ecuador. The holotype was not found in SMT or ZfB where Röber's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Myscelus nobilis distinctus*, mentioned as *Myscelus amystis distinctus* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Yanahurco, Napo, Ecuador, 550 m 6-X-

1978 Lafebre leg./ Neotypus *Myscelus nobilis distinctus* Röber, 1925 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 34.826/. OM-DZUP.

***Myscelus amystis mysus* Evans, 1951**

Pls. 6–7, Figs. 15–16 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia and northwestern Brazil (Acre and Rondônia).

Ecuador: Napo; Pastaza (*Mera*); Tungurahua (*Río Blanco*). **Peru:** Huánuco; Junín (*Perené, Satipo, Shimá*); Loreto (*Ucayali*); Madre de Dios (*Iberia, Parque Nacional del Manu, Tambopata Reserve*); Puno (*Carabaya*); San Martín (*Juanjuí, Tarapoto*); Ucayali (*Pucallpa*). **Bolivia:** Beni (*Reyes, Río Beni, Rurrenabaque*); La Paz (*Caranavi, Muchanes*); Santa Cruz (*Buenavista, Santa Cruz de la Sierra, Sara*). **Brazil:** Acre (*Bujari, Porto Acre, Porto Walter, Santa Rosa do Purus*); Rondônia (*Cacaulândia*).

The citation **Bolivia** (Santiago de Estero, a city in Argentina) (Evans 1951, Palo Jr. 2017) is dubious.

TYPE:

Myscelus amystis mysus Evans, 1951. Holotype male with the following labels: / Type – *mysus* Evans/ Pérou [San Martín] Tarapoto Mai a Août 1886 M. de Mathan/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Myscelus epigona* Herrich-Schäffer, 1869, stat. rest.**

Pls. 6–7, Figs. 17 (♂), 18 (neotype ♂), 19–20 (♀) (d, v)

Syn.: *Myscelus orthrus* Hewitson, 1877

Pyrrhopyga [sic] *sothis* Mabilie, 1883

Pyrrhopyga [sic] *orbius* Mabilie, 1883

Myscelus nobilis var. (ssp.) *meridionalis* Röber, 1925

DISTRIBUTION – French Guiana, Brazil to northern Argentina.

French Guiana: Cayenne (*Montagne de Kaw – Camp Caïman*).

Brazil: Pará (*Óbidos*); Mato Grosso (*Pantanal*); Mato Grosso do Sul (*Jardim*); Goiás (*Vianópolis*); Minas Gerais (*Belo Horizonte, Carmo do Rio Claro, Conceição dos Ouros, Itamonte, Paraopeba, Passa Quatro, Poços de Caldas*); Espírito Santo (*Linhares, Santa Teresa*); Rio de Janeiro (*Cachoeiras de Macacu, Duque de Caxias, Itatiaia, Magé, Petrópolis, Rio de Janeiro*); São Paulo (*Anhangá, Apiaí, Araçatuba, Indiana, Jundiá, Rio Claro, Santos, São Paulo, Teodoro Sampaio – Parque Estadual do Morro do Diabo and Porto Cabral*); Paraná (*Antonina, Balsa Nova, Campina Grande do Sul, Campo Largo, Castro, Curitiba, Diamante do Norte, Foz do Iguaçu, Guaratuba, Londrina, Matinhos, Morretes 20 m, Planaltina do Paraná, Ponta Grossa, Quatro Barras, Rolândia, Terra Boa, Tijucas do Sul, Tunas do Paraná*); Santa Catarina (*Corupá, Dalbérgia, Ibirama, Joinville, Massaranduba, São Bento do Sul – Rio Vermelho and Rio Natal, São Francisco do Sul, Seara – Nova Teutônia, Taió, Timbó*); Rio Grande do Sul (*Encruzilhada do Sul, Ivoit, Feliz, Morro Reuter, Pelotas, Santa Rosa, São Luiz Gonzaga*). **Paraguay:** Alto Paraná (*Itakyry*); Canindeyú; Guairá (*Villarica*); Paraguari (*La Rosada, Sapucaí*). **Argentina:** Chaco; Formosa; Misiones (*Dos de Mayo, General Belgrano, Pozo Azul, Puerto Libertad, Puerto Iguazú, Salto Paredon*).

The citation **Venezuela** (Bell 1934b) is probably incorrect (Orellana [2010]).

TAXONOMY – This species regarded as a subspecies of *Myscelus amystis* by Evans (1951) and followers is here reinstated. This species differs from *Myscelus amystis* (including all subspecies) by the spot of the central band in CuA₁–CuA₂ shifted outwards in both sexes; male genitalia with the harpe distally serrate, not smooth as in *Myscelus amystis* (see Evans 1951, pl. 8). The DNA barcode reveals a minimum interspecific divergence of 3.7% (n= 6) from *Myscelus amystis hages* (n= 13) and *Myscelus amystis mysus* (n= 3).

TYPES:

Myscelus epigona Herrich-Schäffer, 1869. The original description mentions an unspecified number of specimens from Tropical America. None syntypes were found in MfN or ANSP, where Herrich-Schäffer's types are deposited, or in any museum or collection mentioned

above (see “Abbreviations of the collections”). To ensure the correct identification of *Myscelus epigona* Herrich-Schäffer, 1869, mentioned as *Myscelus amystis epigona* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Joinville [sic], Santa Catarina, Brasil, 10-200 m, 20-IV-1969 [O.] Mielke & Miers leg./ Neotypus *Myscelus epigona* Herrich-Schäffer, 1869 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.455/. DZUP.

Myscelus orthrus Hewitson, 1877. The original description mentions specimens in the collection Hewitson, without data [Brazil – on the label of the lectotype]. To ensure the correct identification of *Myscelus orthrus*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ *orthrus*/ Brazil. Hewitson Coll. 79-69 *Myscelus orthrus*. Hew./; two other labels will be added: / Lectotypus/ Lectotypus *Myscelus orthrus* Hewitson, 1877, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [sic] *sothis* Mabilie, 1883. The original description mentions specimens from Brazil in the collection Mabilie. To ensure the correct identification of *Pyrrhopyga sothis*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ *Pyrrhopyga sothis* Mab. am. Mer. [America meridional]/ Ex musao P. Mabilie 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga sothis* Mabilie, 1883, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [sic] *orbius* Mabilie, 1883. The original description mentions specimens from Brazil in the collection Mabilie. To ensure the correct identification of *Pyrrhopyga orbius*, one syntype male is **herein designated as the lectotype**; this specimen has the following labels: / Type H. T/ *Pyrrhopyga orbius* Mab. Bogota [Cundinamarca, Colombia]/ *Mysc. orbius* Mab./ Ex musao P. Mabilie 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga orbius* Mabilie, 1883. O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK. The locality Bogotá is probably incorrect; Evans (1951) mentions Brazil as the type locality.

Myscelus nobilis var. (ssp.) *meridionalis* Röber, 1925. Lectotype male (O. Mielke 1989a) with the following labels: / Type *meridionalis/ meridionalis*/ Stauding. u. Bang-Haas Dresden Ankauf 1961; Staatl. Museum für Tierkunde Dresden/ Lectotypus *Myscelus nobilis meridionalis* Röber, 1925. O. Mielke det. 1989/. SMT.

***Myscelus phoronis* (Hewitson, 1867)**

Two subspecies.

DISTRIBUTION – Guatemala, Panama, central and eastern Colombia to central-eastern Bolivia and northwestern Venezuela.

***Myscelus phoronis phoronis* (Hewitson, 1867)**

Pls. 6–7, Figs. 22–24 (♂) (d, v)

DISTRIBUTION – Guatemala, Panama to western and central Colombia, eastern Colombia to central-eastern Bolivia and northwestern Venezuela.

Guatemala: San Marcos (*San Cristóbal Cucho – Finca los Andes*).

Panama. Colombia: Amazonas (*Leticia*); Boyacá (*Muzo*); Casanare (*Guaicaramo*); Cundinamarca (*Bogotá*); Meta (*Upper Río Negro*); Quindío (*Río Roble x Río La Vieja*). **Ecuador:** Guayas (*Guayaquil*); Loja (*Loja*); Morona-Santiago (*Macas, Pablo Sexto*); Napo (*Archidona, Río Napo, Tena*); Pastaza (*Abitagua, Mera, Puyo, Río Pastaza, Sarayacu*); Tungurahua (*Ambato, Baños, El Topo, Hacienda La Mascota, Hacienda Santa Inés, Río Topo, Río Zunac, Yunguilla*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Rodríguez de Mendoza*); Cusco (*Callanga, Cosñipata Valley – Chontachaca 950 m to Yanamayo 2,350 m, Machu Picchu*); Huánuco (*Carpish, Cucharas, Tingo Maria*); Junín (*Andamarca, Chanchamayo, La Merced, Río Colorado, San Ramón, Satipo, Shimá*); Pasco (*Huancabamba, Oxapampa, Pozuzo, Río Pachitea*); Puno (*Carabaya*); San Martín (*Huayabamba, Juanjuí, Moyobamba, Rioja –*

San Agostin 1,050 m to Jorge Chávez 1,450 m, Tarapoto). **Bolivia**: Beni (*Bueyes*); Cochabamba (*Cochabamba*); La Paz (*Apolobamba, Caranavi, Chaco, Chulumani, Cusilluni, Farinas, Parque Nacional Madidi 1,500–1,800 m, Puente Villa, Río Chairó, Río Zongo, Yanacachi to San Felipe*); Santa Cruz (*Sara*). **Venezuela**: Táchira (*Río Frío, Río Negro*).

TAXONOMY – The specimens from Bolivia are normally lighter, although there are exceptions, in the dorsal hindwing than specimens from Colombia to Peru. Despite of the DNA barcode shows a minimum divergence of 2.1% to 2.4% (Ecuador and Peru specimens (n= 5), Bolivia specimens (n= 2)), none differences were found in the male genitalia and thus, we retain the Bolivian population within this taxon.

TYPE:

Erycides phoronis Hewitson, 1867. The original description mentions an unspecified number of specimens from New Grenada [Colombia] in collection Hewitson. To ensure the correct identification of *Erycides phoronis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ N. Granada [Colombia] Hewitson Coll. 79.-69. *Myseles phoronis*. 2/; two other labels will be added: / Lectotypus/ Lectotypus *Erycides phoronis* Hewitson, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Myseles phoronis caucanus* Staudinger, 1888**

Pls. 6–7, Figs. 21 (lectotype ♂ of *Myseles phoronis* var. *caucanus*) (d, v)

Syn.: *Pyrrhopyga* [*sic*] *persela* Mabilie, 1891

DISTRIBUTION – Northwestern and western Colombia.

Colombia: Antioquia (*Mesopotamia*); Caldas (*Manizales, Río Sucio, Supía – La Quebra, Villamaria – El Arroyo*); Cauca; Valle del Cauca; Department? (*Río Nabasco*).

The citation **Venezuela** (Godman & Salvin 1893) is probably incorrect (Orellana [2010]).

TYPES:

Myseles phoronis var. *caucanus* Staudinger, 1888. The original description mentions an unspecified number of specimens from Caucathal, [Colombia] in collection Staudinger. To ensure the correct identification of *Myseles phoronis* var. *caucanus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Lectotypus/ Cauca [Colombia] Patino/ *Phoronis* var. *Caucanus* Stgr./ *Persela* Mab. *Caucanus* Stgr./ Coll. Staudinger/; another label will be added: / Lectotypus *Myseles phoronis* var. *caucanus* Staudinger, 1888. O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published. There are two more syntypes, herein designated as paralectotypes; these specimens have the following labels: / Origin./ Paralectotypus/ Cauca [Colombia] Patino/ Coll. Staudinger/ Type *caucanus* Staudinger, 1888, O. Mielke det. 1979/; another label will be added to each specimen: / Paralectotypus *Myseles phoronis* var. *caucanus* Staudinger, 1888, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although paralectotypes labels were attached in 1979 by O. Mielke, the designation was never published.

Pyrrhopyga [*sic*] *persela* Mabilie, 1891. The original description mentions an unspecified number of males from Cauca, [Colombia]. To ensure the correct identification of *Pyrrhopyga persela*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin. *Persela* Mab./ Cauca [Colombia] Patino/ 16 : 10/ *Pyrr. persela* Mab./ Coll. Staudinger/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga persela* Mabilie, 1891, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

***Granila* Mabilie, 1903**

One species.

DISTRIBUTION – Eastern Brazil (eastern Bahia to Rio Grande do Sul), northern Argentina and Uruguay.

***Granila paseas* (Hewitson, 1857)**

Pls. 8–9, Figs. 1, 3 (♂), 2, 4 (♀) (d, v)

Syn.: *Granila paseas* var. *albimacula* Mabilie & Boulet, 1908

Granila paseas var. *pullata* J. Zikán, 1938

DISTRIBUTION – Eastern Brazil (eastern Bahia to Rio Grande do Sul), northern Argentina and Uruguay.

Brazil: Bahia (*Nova Venécia* – new name for *Leopoldina*); Minas Gerais (*Caxambu, Passa Quatro, Poços de Caldas*); Espírito Santo (*Linhares, Santa Leopoldina, Santa Teresa*); Rio de Janeiro (*Casimiro de Abreu, Cachoeiras de Macacu, Itatiaia, Niterói, Rio de Janeiro*); São Paulo (*Amparo, Avanhandava, Casabranca, Morro Agudo, Rio Claro, São Bernardo, São José do Rio Preto*); Paraná (*Balsa Nova, Curitiba, Foz do Iguaçu, Londrina, Piên, Ponta Grossa, Rolândia, Turvo*); Santa Catarina (*Blumenau, Dalbérgia, Joinville, Lages, Mafra, Massaranduba, Monte Castelo, Papanduva, São Bento do Sul – Rio Natal, Seara – Nova Teutônia*); Rio Grande do Sul (*Dois Irmãos, Rio Guarita, Ivoti, Morro Reuter, Novo Hamburgo – as Hamburgo Velho, Panambi, Pelotas*). **Paraguay**: Caazapá (*Tapyta*); Guairá (*Villarica*). **Uruguay. Argentina**: Chaco; Misiones (*Bonpland, Dos de Mayo, Puerto Iguazú, Salto Piedras Blancas, San Ignacio*).

The citations **Nicaragua** and **Colombia** (Kirby 1879) are incorrect.

TAXONOMY – There are two phenotypes, one with yellow and the other with white hyaline spots (var. *albimacula*), both sympatric and synchronic.

TYPES:

Pyrrhopyga [*sic*] *paseas* Hewitson, 1857. The original description mentions an unspecified number of males and females from Brazil. To ensure the correct identification of *Pyrrhopyga paseas*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Brazil Hewitson Coll. 79-69 *Pyrrhopyga paseas*. 4./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga paseas* Hewitson, 1857, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Granila paseas var. *albimacula* Mabilie & Boulet, 1908. Holotype (one female is mentioned) with the following labels: / Type/ Rio de Janeiro Brésil IV.1885 F. Woillard Coll. Boulet Museum Paris/. MNHN.

Granila paseas var. *pullata* J. Zikán, 1938. Holotype female with the following labels: / Typus/ 15.1.[19]-25 Maromba an Blüte [on flower] [Itatiaia, Rio de Janeiro, Brazil, Zikán leg., 1.200 m]/ ♀/ *Granila paseas* var. *pullata*, Zik[án]./ Coleção J. F. Zikan/ gen[itália]. prep[arada]., [O.] Mielke 1969/ *Granila paseas* (Hew., 1857) O. Mielke det. 1966/ I.O.C. Lepidoptera 33.016/. IOC.

***Passova* Evans, 1951**

Nine species.

DISTRIBUTION – Belize to southern Brazil, Paraguay and northeastern Argentina.

***Passova ganymedes* (Bell, 1931)**

Three subspecies.

DISTRIBUTION – Eastern Colombia to southeastern Peru and northern Brazil.

***Passova ganymedes ganymedes* (Bell, 1931)**

Pls. 8–9, Figs. 5–6 (♂), 7–8 (♀) (d, v)

DISTRIBUTION – Eastern Colombia to southeastern Peru and northern Brazil.

Colombia: Meta (*Río Negro*). **Ecuador**: Napo (*Río Shingupino*); Pastaza (*Mera*). **Peru**: Huánuco (*Tingo Maria*); Loreto (*Contamana, Muniches*); Madre de Dios (*Upper Río Madre de Dios – Amazonia Lodge*

500 m (near Atalaya, Cusco)); Pasco (Bosque de Protección San Matías – San Carlos). **Brazil:** Amazonas (São Paulo de Olivença); Rondônia (Cacaulândia); Mato Grosso (Diamantino).

TYPE:

Pyrrhopyge ganymedes Bell, 1931. Holotype male with the following labels: / Holotype *Pyrrhopyge ganymedes* Bell, 1931/ Colombia 1925/ Genitalia slide No. 1957/ Det. E.L. Bell/. ANSP, transferred to CMP.

***Passova ganymedes gulia* Evans, 1951**

Pls. 8–9, Figs. 9 (♂), 10 (holotype ♀) (d, v)

DISTRIBUTION – Eastern Ecuador to northern Peru.

Ecuador: Loja (*Loja*); Napo (*Misahualli*); Pastaza (*Mera – Shell*).

Peru: Amazonas (*Alto Nieva 1,930 m*).

TYPE:

Passova ganymedes gulia Evans, 1951. Holotype female with the following labels: / Type – *gulia* Evans/ Environs de Loja Equateur 1893/ J. J. Joicey Coll. B.M. 1925-451/ 32. 21. Ex. Coll. Dognin.1921/. NHMUK.

***Passova ganymedes gelina* Evans, 1951**

Pls. 8–9, Figs. 11–12 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to southeastern Peru.

Ecuador: Pastaza (*Upper Pastaza*). **Peru:** Cusco (*Quincemil*); Huánuco; Junín (*Chanchamayo, Perené*); Madre de Dios (*Tambopata Reserve*); Pasco (*Pozuzo*).

TYPE:

Passova ganymedes gelina Evans, 1951. Holotype male with the following labels: / Type – *gelina* Evans/ Pérou [Junín] Chanchamayo O. Schunke 1898/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Passova gellias* (Godman & Salvin, 1893)**

Pls. 8–9, Figs. 13 (holotype ♂), 14–15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Belize to central Colombia and northern Venezuela.

Belize: Toledo (*Gloria Camp*). **Honduras:** Atlántida (*La Ceiba*).

Costa Rica: Guanacaste (*Área de Conservación Guanacaste*). **Panama:** Chiriquí; Panamá (*El Llano, Madden Forest*); Panamá Oeste (*Cerro Campana*). **Colombia:** Boyacá (*Muzo*). **Venezuela:** Táchira (*Río Uracá*); Zulia (*Las Tres Bocas*).

The citation **Brazil:** Santa Catarina (*Águas Mornas – Teresópolis*) (Evans 1951) refers to *Passova polemon*.

TAXONOMY – There are two phenotypes, one with a little to completely red fore coxa and the other with a completely black fore coxa. There is no difference in the male genitalia.

TYPE:

Pyrrhopyge gellias Godman & Salvin, 1893. Holotype (one male is mentioned with the following labels: / Holotypus/ Costarica [??]/ ♂/ 776/ 1: 27./ *Pyrrhopyge gellias*. G.&S. ♂/ Sp. figured/ Coll. Staudinger/. MfN.

***Passova glacia* Evans, 1951**

Pls. 8–9, Figs. 18 (♂), 19–20 (♀) (d, v)

DISTRIBUTION – Suriname, French Guiana and northern Brazil (Amazonas and Pará).

Suriname. French Guiana. Brazil: Amazonas (*Manicoré, Maués*); Pará (*Óbidos*).

TYPE:

Passova glacia Evans, 1951. Holotype male with the following labels: / Type – *glacia* Evans/ Guyana française, Collect. C. Bar./ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Passova greta* Evans, 1951**

Pls. 8–9, Figs. 21–23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Northern Peru to central-eastern Bolivia.

Peru: Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to Quebrada Buenos Aires 2,400 m*); Junín (*Huancabamba, La Merced, San Ramón*); Pasco (*Huancabamba*).

Bolivia: Cochabamba (*Cristal Mayu*); La Paz (*Caranavi, Mapiri, Yungas*).

TYPE:

Passova greta Evans, 1951. Holotype male with the following labels: / Type – *greta* Evans/ Huancabamba [Pasco] N. E. Peru 3000-10,000ft./ NHMUK.

***Passova nigrocephala* (Bell, 1934)**

Pls. 10–11, Figs. 1 (♂) (d, v)

DISTRIBUTION – Western and central Colombia to southeastern Peru.

Colombia: Cundinamarca (*Bogotá*); Valle del Cauca (*Cali – Río Vitaco*). **Peru:** Cusco; San Martín (*Rioja – Naranjos*).

TYPE:

Pyrrhopyge nigrocephala Bell, 1934. Holotype male with the following labels: / Holotype/ Columbien/ 211/ G419/ *Pyrrhopyge nigrocephala* Bell Holotype ♂/. AMNH.

***Passova gazera* (Hewitson, 1866)**

Pls. 10–11, Figs. 2 (♂), 3 (♀) (d, v)

DISTRIBUTION – Colombia to southeastern Peru and northern Brazil.

Colombia. Ecuador: Napo (*Ahuano, Misahualli*). **Peru:** Junín (*Chanchamayo*); Loreto (*Achinamiza, Iquitos, Río Putumayo*); Madre de Dios (*Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*). **Brazil:** Amazonas (*Manaus, Rio Madeira, São Paulo de Olivença*); Pará (*Belém, Santarém*); Acre (*Bujari, Senador Guiomard*); Rondônia (*Alto Paraíso, Cacaulândia*).

TYPE:

Pyrrhopyga [sic] gazera Hewitson, 1866. The original description mentions an unspecified number of males from St. Paulo [de Olivença], Amazonas, [Brazil]. To ensure the correct identification of *Pyrrhopyga gazera*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Amazons. [Brazil] Hewitson Coll. 79.-69. *Pyrrhopyga gazera*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga [sic] gazera* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Passova vilna* Evans, 1951**

Two subspecies.

DISTRIBUTION – Southeastern Peru to Bolivia.

***Passova vilna vilna* Evans, 1951**

Pls. 10–11, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (*Callanga*). **Bolivia:** Beni (*Bueyes*); La Paz (*Parque Nacional Madidi 1,500–1,800 m*).

REMARKS – The specimen illustrated on Pls. 14–15, Figs. 12 is probably an aberration of this species. The genitalia are similar.

TYPE:

Passova vilna Evans, 1951. Holotype male with the following labels: / Type – *vilna* Evans/ Bueyes, [Beni] Bolivia. Garlepp./ Godman-Salvin Coll. 1912.-23. *Pyrrhopyge phylleia*, Hew./ NHMUK.

***Passova vilna vanta* Evans, 1951**

Pls. 10–11, Figs. 4 (holotype ♂) (d, v)

DISTRIBUTION – Bolivia.

Bolivia.

REMARKS – Only known from the holotype.

TYPE:

Passova vilna vanta Evans, 1951. Holotype male with the following labels: / Type – *vanta* Evans/ Bolivia Crowley Bequest. 1901-78./ 343/. NHMUK.

***Passova passova* (Hewitson, 1866)**

Six subspecies.

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia, northwestern Venezuela to French Guiana, northern to southern Brazil, Paraguay and northeastern Argentina

***Passova passova passova* (Hewitson, 1866)**

Pls. 10–11, Figs. 9 (♂), 10–11 (♀) (d, v)

DISTRIBUTION – Central Colombia, eastern Ecuador to central-eastern Bolivia, Suriname, French Guiana and northern Brazil (Amazonas and Rondônia).

Colombia: Boyacá (*Muzo*). **Ecuador:** Napo (*Jatun Sacha*). **Peru:** Junín (*Perené, San Ramón to Pueblo Pardo*); Madre de Dios; San Martín (*Roque*). **Bolivia:** La Paz (*Caranavi area*). **Suriname.** **French Guiana:** Cayenne (*Cayenne, Regina, Tonate*). **Brazil:** Amazonas (*Tefé*); Pará (*Itaituba*); Rondônia (*Cacaulândia*).

The citations **Bolivia** (Mabille & Boullet 1908, Mabille 1912, Draudt 1921) are probably incorrect.

TYPE:

Pyrrhopyga [**sic**] *passova* Hewitson, 1866. The original description mentions an unspecified number of males and females from Ega [Tefé, Amazonas, Brazil] and Cayenne, [French Guiana] in collection Hewitson. Evans (1951) mentioned the syntype female from French Guiana as the “type”. To ensure the correct identification of *Pyrrhopyga passova*, this “type” [syntype] female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Cayenne [French Guiana] Hewitson Coll. 79.-69. *Pyrrhopyga passova*. 3./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga passova* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Passova passova rudex* Evans, 1951**

Pls. 10–11, Figs. 7 (♂), 8 (♀) (d, v)

DISTRIBUTION – Eastern Colombia to southeastern Peru, northwestern Venezuela and northern Brazil.

Colombia: Amazonas (*Letícia, Loretoyacu*); Arauca; Meta (*La Macarena, Río Guejas, San Luis de Cubarral*). **Ecuador:** Morona-Santiago (*Santiago de Méndez*); Napo (*Cotundo, Jondachi, Tena, Yanahurco*); Pastaza (*Sarayacu*). **Venezuela:** Aragua (*Maracay, Parque Nacional Henry Pittier*); Carabobo (*San Esteban*); Táchira (*Otauales – San Felix, San Juan de Colón*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo, Perené, Río Colorado, Satipo, Shimá*); Loreto (*Caballococha, Iquitos*); Madre de Dios (*Parque Nacional del Manu, Salvación, Upper Río Madre de Dios*); Pasco (*Pozuzo*); Puno (*Pilcopata*); San Martín (*Juanjuí, Rioja – Mina de Sal, Shanusi – Cumbase, Tarapoto*). **Brazil:** Amazonas (*Benjamin Constant, Coari, Manaus, Mariscal Cáceres, Maués, São Paulo de Olivença, Tefé*); Pará (*Cametá, Itaituba*); Acre (*Porto Acre, Senador Guimard*); Rondônia (*Cacaulândia*).

TYPE:

Passova passova rudex Evans, 1951. Holotype male with the following labels: / Type – *rudex* Evans/ Chanchamayo [Junín] Peru (Schuncke)./ Rothschild Bequest B.M.1939-1./ NHMUK.

***Passova passova styx* (Möschler, 1879)**

Pls. 10–11, Figs. 12 (♂), 13 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *anina* Plötz, 1879, **nom. nud.** (ICZN, Art. 11.6)

DISTRIBUTION – Eastern Colombia to northern Argentina, Venezuela and Guyana.

Colombia: Meta (*Villavicencio*). **Peru:** Junín (*San Ramón*); Madre de Dios (*Parque Nacional Bahuaia Sonene, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco), Parque Nacional del Manu, Tambopata Reserve*). **Bolivia:** Cochabamba (*Chapare*); La Paz (*Caranavi*); Santa Cruz (*André Ibañez, Buenavista, Santa Cruz de la Sierra*). **Venezuela:** Amazonas (*Ugueto, Puerto Ayacucho*); Aragua (*Choroní, Pozo del Diablo*); Carabobo (*San Esteban*); Guárico (*Jabillal*); Sucre (*Finca Vuelta Larga*); Táchira (*Río Frío*). **Guyana:** East Berbice-Corentyne (*Friendship, Mount Roraima*). **Argentina:** Santiago del Estero.

Evans (1951) mentioned Santiago del Estero as in **Bolivia**, but this city is only known to us in Argentina. Some references to **Venezuela** could be of *P. p. stacer*. Also, *P. p. styx* is mentioned from **Guyana** by Hall (1939, 1940), probably a misidentification; at that time *P. p. stacer* was not described.

TYPES:

Pyrrhopyga [**sic**] *styx* Möschler, 1879. The original description mentions two specimens from Colombia, one in collection Möschler and the other in collection Pogge. To ensure the correct identification of *Pyrrhopyga styx*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Am: merid. Columbia 76. Sch./ Type Verhdlg. Zool.-Bot. Gesellschaft 1878. p. 209/ *styx* Möschl/ Coll. Möschl./ Coll. Staudinger/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga styx* Möschler, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyga [**sic**] *anina* Plötz, 1879. The name was published as a synonym of *Pyrrhopyga styx*, a **nomen nudum** (ICZN, Art. 11.6).

***Passova passova stacer* Evans, 1951**

Pls. 10–11, Figs. 14 (♂), 15 (♀) (d, v)

DISTRIBUTION – Northwestern Venezuela to Suriname and northern Brazil (Roraima).

Venezuela: Barinas (*Barinitas*); Carabobo (*Puerto Cabello, Yuma*). **Guyana:** Mazaruni-Lower Berbice (*Mont Roraima*); East Berbice-Corentyne (*Berbice, Friendship*). **Suriname.** **Brazil:** Roraima (*Amajari – Tepequén*).

TYPE:

Passova passova stacer Evans, 1951. Holotype male with the following labels: / Type – *stacer* Evans/ B[ritish]. Guiana.1916./ J.J.Joicey Coll. B.M.1925-451./ NHMUK.

***Passova passova gortyna* (Hewitson, 1866)**

Pls. 10–11, Figs. 16 (lectotype ♂ of *Pyrrhopyga gortyna*), 17–18 (♀) (d, v)

DISTRIBUTION – Northern Brazil (Amazonas and Pará).

Brazil: Amazonas (*Tefé*); Pará (*Belém, Gurupá*).

TYPE:

Pyrrhopyga [**sic**] *gortyna* Hewitson, 1866. The original description mentions an unspecified number of specimens from Ega [Tefé, Amazonas, Brazil] in collection Hewitson. To ensure the correct identification of *Pyrrhopyga gortyna*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Ega [Amazonas, Brazil] Hewitson Coll. 79.69. *Pyrrhopyga gortyna*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga gortyna* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Passova passova diamantino* O. Mielke, C. Mielke
& Casagrande, ssp. n.**

Pls. 10–11, Figs. 19 (holotype ♂), 20 (allotype ♀) (d, v)

DISTRIBUTION – Central-northern Brazil (Mato Grosso and Tocantins).

Brazil: Mato Grosso (*Barra do Bugres, Brasnorte, Cáceres, Diamantino*); Tocantins (*Ilha do Bananal*).

DESCRIPTION – Wing size similar to that of *Passova passova practa*, the smallest subspecies: male and female forewing length 19–22 mm and 23–25 mm, respectively. Red tornal lobe of the hindwing not reaching CuA₂ as it does in *P. p. practa*. The black ground colour lighter than in the other subspecies of *P. passova*. Ventral hindwing with a diffuse light blue basal band between the humeral angle and 2A and faintly from the latter to the anal margin (two males and two females), or this band more diffuse and reduced, between the humeral angle and 2A (two males and one female).

ETYMOLOGY – The specific name refers to Diamantino, one of the Mato Grosso municipalities where this subspecies occurs.

TYPES:

Holotype male with the following labels: / Holotypus/ 23-VII-1987 Brasnorte, MT. [Mato Grosso, Brasil], Miers leg./ Holotypus *Passova passova diamantino* O. Mielke, C. Mielke & Casagrande. O. Mielke, C. Mielke & Casagrande det. 2017/ OM 37.022/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Cáceres, MT [Mato Grosso, Brasil], 9-XI-1984, Buzzi, [O.] Mielke, Elias, Casagrande leg. Proj.[eto] Polonoroeste/ Allotypus *Passova passova diamantino* O. Mielke, C. Mielke & Casagrande. O. Mielke, C. Mielke & Casagrande det. 2017/ DZ 16.524/. DZUP.

Paratypes: 1 ♂, 28-V-1978, 1 male, 16-VI-1978, Alto Rio Arinos, Fazenda São João, Diamantino, Mato Grosso, Brazil, 300–400 m, E. Furtado leg., OM 37.214, OM 34.766 (OM); 1 ♂, 30-VIII-1979, Alto Rio Arinos, Fazenda São João, Diamantino, Mato Grosso, Brasil, 300–400 m, E. Furtado leg., DZ 22.583 (DZUP); 1 ♀, 8-IX-1974, Alto Rio Paraguay, Barra do Bugres, Mato Grosso, Brazil, 150 m, E. Furtado leg., DZ 2.084 (DZUP); 1 ♀, 23-VI-1979, Ilha do Bananal, Tocantins, Brazil, Gifford leg., DZ 22.599 (DZUP).

***Passova passova practa* Evans, 1951**

Pls. 10–11, Figs. 21 (♂), 22 (♀) (d, v)

DISTRIBUTION – Central to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Mato Grosso (*Chapada dos Guimarães – Buriti, Pantanal, São Vicente, Vale dos Sonhos*); Goiás (*Leopoldo Bulhões*); Minas Gerais (*Conceição dos Ouros*); São Paulo (*Campinas, Pereira Barreto*); Paraná (*Castro, Diamante do Norte, Foz do Iguaçu, Loanda, Mauá da Serra, Paranavaí, Planaltina do Paraná, Santo Antônio do Caiuá*); Rio Grande do Sul (*Tenente Portela*). **Paraguay:** Alto Paraná (*Hernandarias*); Canindeyú; Central (*San Bernardino*); Paraguari (*Sapucai*). **Argentina:** Misiones (*Loreto, Puerto Iguazú, Salto Piedras Blancas*).

TYPE:

Passova passova practa Evans, 1951. The holotype male with the following labels: / Type – *practa* Evans/ Castro, Parana [Brazil]./ E.D.Jones Coll. Brit. Mus. 1919-295./. NHMUK.

***Passova polemon* (Hopffer, 1874)**

Pls. 10–11, Figs. 23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Central to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Mato Grosso (*Pantanal*); Goiás (*Taguatinga*); Bahia; Espírito Santo (*Santa Teresa*); Minas Gerais (*Ibirité, Mar de Espanha, Poços de Caldas*); Rio de Janeiro (*Cachoeiras de Macacu, Itatiaia, Petrópolis*); São Paulo (*Campos do Jordão, Cananéia, Ilha do Cardoso, Intervales, Jundiá, Serra da Bocaina, Teodoro Sampaio – Parque Estadual do Morro do Diabo*); Paraná (*Antonina, Campina Grande do Sul, Curitiba, Foz do Iguaçu, Guarapuava, Guaraqueçaba, Guaratuba, Londrina, Matinhos, Morretes 20 m, Paranaguá, Pontal do Paraná,*

Quatro Barras, Tuneiras do Oeste, Ventania); Santa Catarina (*Agrolândia, Águas Mornas – Teresópolis, Joinville, Rio dos Cedros, São Bento do Sul, Taió*); Rio Grande do Sul (*Dois Irmãos, Pareci Novo, Tenente Portela*).

Paraguay: Guairá (*Villarica*). **Argentina:** Misiones (*General Belgrano*).

The citations **Brazil:** Amazonas (Mabille & Boulet 1908), **Peru:** Pasco (*Oxapampa*) (Martin 1941), **Venezuela:** Distrito Capital (*Caracas*) (Lichy 1943, Beccaloni *et al.* 2008) are incorrect. The citation **Venezuela** (Bell 1946) is also incorrect and refers to *Pyrrhopyge caribe caribe* (see Orellana [2010]).

TYPE:

Pyrrhopyge polemon Hopffer, 1874. The original description mentions an unspecified number of females from Brazil [Bahia – on the label of the lectotype]. To ensure the correct identification of *Pyrrhopyge polemon*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Lectotypus/ 4885/ Bahia [Brazil] Sello/ *Polemon* Hpfr*/ 1: 13./; another label will be added: / Lectotypus *Pyrrhopyge polemon* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published.

***Aspitha* Evans, 1951**

Five species.

DISTRIBUTION – Panama to central-eastern Bolivia, Trinidad and Tobago to Guyana and northern Brazil.

***Aspitha teffa* Evans, 1951, stat. rev.**

Pls. 12–13, Figs. 1 (♂), 2 (♀) (d, v); Text-Fig. 7 (♂ genitalia)

DISTRIBUTION – Northern Brazil (Amazonas and Acre).

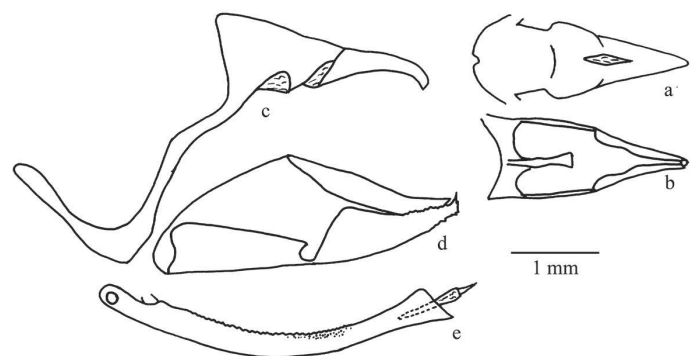
Brazil: Amazonas (*Tefé*); Acre (*Xapuri*).

TAXONOMY – This species was known for a long time only from two females (one illustrated here and the holotype). The discovery of the male from Xapuri (Acre, Brazil) shows that it is a valid species and not just the female of *A. aspitha aspitha*, as mentioned by O. Mielke & Casagrande (2002). So, this taxon returns to full species status.

The unique known male is similar to the female, but smaller (forewing length 21 mm), the fringes of the anal lobe are dorsally orange, and ventrally also an adjacent smooth area with the same colour. There are three white transverse stripes at the base of the abdomen ventrally. The male genitalia are similar to *Aspitha agenoria* and *A. aspitha*.

TYPE:

Aspitha teffa Evans, 1951. Holotype female with the following labels: / Type – *teffa* Evans/ Tefé (Ega) Amazonas [Brazil] M. de Mathan 7^{brc} 8^{brc} 1879/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.



Text-Fig. 7. *Aspitha teffa* stat. rev. Male genitalia: a) tegumen and uncus, dorsal view; b) gnathos and uncus, ventral view; c) tegumen, saccus and uncus, lateral left view; d) right valva, internal view; e) aedeagus, lateral left view (Brazil, Acre, Xapuri, Seringal Esperança, 6-7-IX-2004, Mielke & Casagrande leg., DZ 22.653).

Aspitha aspitha (Hewitson, 1866)

Three subspecies.

DISTRIBUTION – Panama, central and eastern Colombia to central-eastern Peru, Trinidad and Tobago to French Guiana and northern Brazil.

Aspitha aspitha aspitha (Hewitson, 1866)

Pls. 12–13, Figs. 3 (♂) (d, v); Pls. 74–75, Figs. 23 (♀) (d, v)

DISTRIBUTION – Central and eastern Colombia to central-eastern Peru, Trinidad and Tobago to Guyana and northern and northwestern Brazil.

Colombia: Amazonian area; Huila (*Palestina* – *Jericó*); Meta (*Villavicencio*). **Ecuador:** Napo (*Archidona*). **Peru:** Junín (*Chanchamayo* – *San Luis Shuaro*); Loreto (*Iquitos*). **Trinidad and Tobago:** Siparia (*Parry Lands*). **Guyana:** Upper Takutu – Upper Essequibo (*Kanuku Mountains, Nappi Creek*). **Brazil:** Amazonas (*Manaus, Marabitanas, São Gabriel da Cachoeira, São Paulo de Olivença, Rio Tacana, Tefé, Tonantins*); Pará (*Belém*).

TYPE:

Pyrrhopyga [*sic*] *aspitha* Hewitson, 1866. The original description mentions an unspecified number of males and females from Pará and St. Paulo [de Olivença], Amazonas, [Brazil]. To ensure the correct identification of *Pyrrhopyga aspitha*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ St. Paulo [de Olivença, Amazonas, Brazil]. Hewitson Coll. 79.69. *Pyrrhopyga aspitha* 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga aspitha* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK. Another syntype female is herein designated as paralectotype; this specimen with the following labels: / Type/ Para [Brazil] Hewitson Coll. 79.-69 *Pyrrhopyga aspitha*. 2./ 1039. 66/; two other labels will be added: / Paralectotypus/ Paralectotypus *Pyrrhopyga aspitha* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Aspitha aspitha rufescens (Riley, 1919)

Pls. 12–13, Figs. 4–5 (♂) (d, v)

DISTRIBUTION – Northwestern Brazil (Amazonas and Rondônia).

Brazil: Amazonas (*Manicoré, Maués*); Acre (*Bujari, Mâncio Lima* – *Parque Nacional da Serra do Divisor*); Rondônia (*Abunã, Cacaupallândia, Candeias do Jamarí*).

The citation **Trinidad and Tobago:** Penal-Debe (*Morne Diable*) (Kaye 1940) is probably incorrect.

TYPE:

Yanguna rufescens Riley, 1919. Holotype (one male is mentioned) with the following labels: / Type H T/ Abuna Upper Rio Madeira [Rondônia, Brazil] Oct. 1913 R. T. Wickham/ E.H.W. Wickham 1914-94/ B.M. Type No. Rh. 102/ *Yanguna* ♂ *rufescens* Riley./ NHMUK.

Aspitha aspitha parima (Plötz, 1886)

Pls. 12–13, Figs. 6 (♂), 7 (neotype ♂ of *Pyrrhopyga parima*) (d, v)

DISTRIBUTION – Panama to central Colombia, Suriname, French Guiana and northern Brazil (Amazonas and Pará).

Panama: Chiriquí. **Colombia:** Boyacá (*Muzo*). **Suriname:** French Guiana: Cayenne (*Montagne de Kaw: Roura, Camp Patawa*). **Brazil:** Amazonas; Pará (*Óbidos*).

The citations **Bolivia** (Mabille & Boullet 1908, Mabille 1912, Draudt 1921) are probably incorrect.

TYPE:

Pyrrhopyga [*sic*] *parima* Plötz, 1886. The original description mentions an unspecified number of specimens from Suriname. No syntype was found in MfN or ZSM, where nearly all Plötz's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Pyrrhopyga parima*, mentioned as *Aspitha aspitha parima* by Evans

(1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Surinam Cardua 24/ 20./; two other labels will be added: / Neotypus/ Neotypus *Pyrrhopyga parima* Plötz, 1886 O. Mielke, Brockmann & C. Mielke det. 2015/. MSFR.

Aspitha leander (Boullet, 1912)

Pls. 12–13, Figs. 8, 11 (♂), 9–10 (♀) (d, v); Pls. 14–15, Figs. 8 (♀) (d, v)

DISTRIBUTION – Southern Mexico to western and central Colombia and central-eastern Peru.

Mexico: Chiapas; Oaxaca (*Istmo* – *Caracol*). **Costa Rica:** Puntarenas (*Uvita*); San José (*Parque Braulio Carillo*). **Panama:** Colón (*Gatun, Piña*); Panamá (*El Llano* – *Cordillera de San Blas, Río Partí*). **Colombia:** Antioquia (*La Ratонера*); Boyacá (*Muzo*); Chocó (*San José del Palmar*); Cundinamarca (*Bogotá, Cananche, La Vega*). **Peru:** Pasco (*Pozuzo*).

TYPE:

Yanguna leander Boullet, 1912. Holotype (one male is mentioned) with the following labels: / Type/ Muzo [Boyacá] Colombia 800 m 1911 A. H. Fassl Coll. Boullet Museum Paris/ *Yanguna leander* Boullet Bull. Soc. Ent. Fr., 1912, p. 92/ *Y. leander* E. Boull. Bⁱⁿ S. E. F. p. (1911)/. MNHN.

Aspitha bassleri (Bell, 1940)

Pls. 12–13, Figs. 12 (♂), 13 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to southeastern Peru and northwestern Brazil (Rondônia).

Ecuador: Napo (*Archidona*). **Peru:** Amazonas (*Río Santiago*); Huánuco (*Río Huallaga*); Loreto (*Río Putumayo*); Madre de Dios (*Iberia, Tambopata Reserve*); Pasco (*Pozuzo*); San Martín (*Achinamiza*). **Brazil:** Rondônia (*Cacaupallândia*).

TYPE:

Pyrrhopyge bassleri Bell, 1940. Holotype male with the following labels: / Holotype, Achinamiza, [San Martín] Peru 11 IX '19]27 F 6001/ H. Bassler Collection Acc. 33591/ G 1408/ *Pyrrhopyge bassleri* Bell. Holotype ♂/. AMNH.

Aspitha agenoria (Hewitson, 1876)

Two subspecies.

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia

Aspitha agenoria agenoria (Hewitson, 1876)

Pls. 12–13, Figs. 14–15 (♂) (d, v)

Syn.: *Pyrrhopyge sanies* H. H. Druce, 1908, **syn. n.**

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia. Likely Suriname and northern Brazil (Pará?).

Colombia: Cundinamarca (*San Martín*); Meta (*Llanos, Villavicencio*). **Peru:** Cusco (*Cosñipata Valley* – *Quebrada Quitacalzón 1,100 m* – both phenotypes, *Illapani Viejo* – phenotype *sanies*); Huánuco (*Tingo Maria* – both phenotypes); Junín (*Chanchamayo* – both phenotypes, *La Merced, Perené* – phenotype *sanies*, *Río Colorado, Satipo* – both phenotypes, *Shimá* – both phenotypes); Madre de Dios (*Parque Nacional del Manu* – phenotype *sanies*, *Salvación* – phenotype *sanies*, *Shintuya* – phenotype *sanies*, *Upper Río Madre de Dios* – *Amazonia Lodge 500 m (near Atalaya, Cusco)* – phenotype *sanies*, *Upper Río Madre de Dios* – *Mascoitania* – phenotype *sanies*, *Tambopata Reserve* – phenotype *sanies*); San Martín (*Juanjuí* – phenotype *sanies*, *Mariscal Cáceres* – phenotype *sanies*, *Tarapoto*). **Bolivia:** Beni (*Río Beni* – phenotype *sanies*); La Paz (*Caranavi, Coroico, Farinas* – phenotype *sanies*, *Muchanes* – phenotype *sanies*, *Río Zongo, San Ernesto, Yungas*);

Cochabamba (*Chapare*); Potosí (*Tarna*); Santa Cruz (*Parque Nacional Amboró, Sara*). **Suriname** (?). **Brazil**: Pará (?).

TAXONOMY – *Aspitha agenoria agenoria* and *A. agenoria sanies* (H. H. Druce, 1908) present two phenotypes, the first without and the second with a central hyaline white band on the forewing. Both are sympatric at some localities and DNA barcode shows no difference, reason to consider *Aspitha agenoria sanies* a junior synonym.

TYPES:

Pyrrhopyga [*sic*] *agenoria* Hewitson, 1876. The original description mentions an unspecified number of specimens from Chanchamayo, Peru, Thamm leg. in collection Staudinger. To ensure the correct identification of *Pyrrhopyga agenoria*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Typus/ Chanchamayo [Junín, Peru] Thamm/ *agenorial Agenoria* Hew./ Coll. Staudinger/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga agenoria* Hewitson, 1876, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyge sanies H. H. Druce, 1908. The original description mentions an unspecified number of males from Farinas, La Paz, Bolivia, 1500 m in collection Druce. To ensure the correct identification of *Pyrrhopyge sanies*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Farinas La Paz Bolivia 1500 m/ *Pyrrhopyge sanies* H. H. Druce Type T.E.S. 1908./ ex coll. Hamilton Druce, 1919./ J.J. Joicey Coll. B.M.1925-451/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge sanies* H. H. Druce, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK. **Syn. n.**

***Aspitha agenoria cruor* (H. H. Druce, 1908)**

Pls. 12–13, Figs. 16 (lectotype ♂ of *Pyrrhopyge cruor*) (d, v)

DISTRIBUTION – Central-eastern Peru.

Peru: Pasco (*Pozuzo*).

TYPE:

Pyrrhopyge cruor H. H. Druce, 1908. The original description mentions an unspecified number of males from Pozuzo, Peru, 800 m, I. Egg leg. in collection Druce. To ensure the correct identification of *Pyrrhopyge cruor*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Pozuzo [Pasco] Peru I. Egg. 800.m/ *Pyrrhopyge cruor* H. H. Druce Type T.E.S. 1908./ ex coll. Hamilton Druce, 1919/ J.J. Joicey Coll. B.M.1925-451/; two other will be added: / Lectotypus/ Lectotypus *Pyrrhopyge cruor* H. H. Druce, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

ZONIINI O. Mielke, 2001

One genus.

TAXONOMY – The tribe is characterized by the genome (Zhang *et al.* 2019) and by some morphological characters: forewing discocellular cell larger than the inner margin plus the median and lower discocellular veins; forewing with veins R_5 and M_1 approximate at the basal fourth portion, then abruptly divergent for a short distance, and then more or less parallel until the margin; forewing with the lower discocellular vein more or less in line with m–cu; forewing of the male projected; metathoracic leg with short femurs, same size as the basal part of the tibia (base until first pair of spurs); gnathos U-shaped, with the union directed distally; vesica with three large cornuti, each of which composed by many small spines; futura inferior V-shaped and separated from the aedeagus; lamella postvaginalis transversally large, with a blunt projection on the central anterior portion, and distally with a little central concavity; bursa copulatrix with signa.

***Zonia* Evans, 1951**

One species.

DISTRIBUTION – Belize to southwestern Brazil.

***Zonia zonia* Evans, 1951**

Four subspecies.

DISTRIBUTION – Belize to northern Colombia, Eastern Peru, Guyana and northern to southern Brazil.

***Zonia zonia zonia* Evans, 1951**

Pls. 12–13, Figs. 19 (♂), 20 (♀) (d, v)

DISTRIBUTION – Eastern Peru, Guyana and northern Brazil.

Peru: Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m*); Loreto (*Iquitos*); Madre de Dios (*Pampas del Heath 200 m, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Oxapampa, Pozuzo*). **Guyana.** **Brazil:** Amazonas (*Tefé*); Pará (*Juruti, Santarém*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*); Mato Grosso (*Diamantino*).

TYPE:

Zonia zonia Evans, 1951. Holotype male with the following labels: / Type – *zonia* Evans/ Holotype/ Teffè (Ega) Amazonas [Brazil] M. de Mathan 1^{er} Trimestre 1879/ R. Oberthür Coll. Brit. Mus. 1931-136/ B.M.(N.H.) Rhopalocera slide 11,348/ *Zonia zonia* Evans det. R.I. Vane - Wright, 1973 Holotype ♂/ NHMUK(E)#982984/. NHMUK.

***Zonia zonia panamensis* Nicolay, 1975**

Pls. 12–13, Figs. 17 (holotype ♂), 18 (♀) (d, v)

DISTRIBUTION – Belize to northern Colombia.

Belize: Stann Creek (*Cockscomb*). **Honduras.** **Panama:** Panamá (*Madden Forest*). **Colombia:** Cesar (*Pueblo Bello*).

TYPE:

Zonia zonia panamensis Nicolay, 1975. Holotype male with the following labels: / Madden Forest Panama C. Z. [Panama] 6. Feb. '68 S. S. Nicolay/ ♂ genitalia slide/ vial # H. 379 Prep. S. S. Nicolay/ Photo/ Holotype ♂ *Zonia zonia panamensis* S. S., Nicolay/. AMNH.

***Zonia zonia diabo* O. Mielke & Casagrande, 1998**

Pls. 12–13, Figs. 21 (holotype ♂), 22 (♂) (d, v)

DISTRIBUTION – Central-southern to southwestern Brazil.

Brazil: Goiás (*Pirenópolis*); São Paulo (*Teodoro Sampaio – Parque Estadual do Morro do Diabo*); Paraná (*Foz do Iguaçu, Terra Rica*).

REMARKS – Only known from few males, all hilltopping.

TYPE:

Zonia zonia diabo O. Mielke & Casagrande, 1998. Holotype male with the following labels: / Holotypus/ 22-25-X-1987, Morro do Diabo, Teodoro Sampaio, SP [São Paulo, Brazil], 300-600 m, [O.] Mielke & Casagrande [leg.]/ Holótipo *Zonia zonia diabo* [O.] Mielke & Casagrande, O. Mielke det. 1997 / [OM] 17.893/. OM-DZUP.

***Zonia zonia eberti* O. Mielke, C. Mielke & Casagrande, ssp. n.**

Pls. 12–13, Figs. 23 (holotype ♂), 24 (allotype ♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil (Rio de Janeiro and Santa Catarina).

Brazil: Rio de Janeiro (*Casimiro de Abreu*); Santa Catarina.

DESCRIPTION – Male: forewing length 27 mm. Very similar to *Zonia zonia zonia* with respect to the semi-hyaline spots on the fore and hindwings. Differs, however, in the yellow colour of the two dorsal and longitudinal stripes on the thorax (the innermost over the tegulae), of the lateral sides of the abdominal tergum, of the basal and sub-basal stripes on the dorsal forewing, and of the two basal stripes, parallel to CuA, on the dorsal surface of the hindwing, which are blue in *Zonia zonia zonia*.

Female: forewing length 52 mm. The general appearance of the wings is more rounded than in the male. Differs from *Z. zonia zonia* by the semi-hyaline spot of the discal cell on the forewing being the same width of similar spots on the costal margin; by the semi-hyaline spot on M_3 –CuA₁, as well as the apical semi-hyaline spots, mostly between

M₁–M₂ and M₂–M₃, being a little wider (narrower in *Z. zonia zonia*). The blue colour of the submarginal band, on the dorsal face of the forewing, is quite spread out internally in CuA₁–CuA₂; the ventral face with no traces of blue between Sc–R₁, R₁–R₂ and the costal half of discal cell, after the median band. The hindwing on the ventral face, with the blue spots, next to the semi-hyaline cellular spot, very small (larger in *Z. zonia zonia*).

TAXONOMY – *Zonia zonia eberti* **ssp. n.** is also distinguished from the other two subspecies by the yellow colour on the dorsal part of the thorax, on the abdomen and by the basal and sub-basal bands on the dorsal surface of the forewing, and of the two basal stripes, parallel to CuA, on the dorsal surface of the hindwing; and in addition from *Zonia zonia diabo* on the forewing by the median semi-hyaline band and by the apical semi-hyaline spots, being both narrower. Also is distinguished from *Zonia zonia panamensis* on the forewing by the median semi-hyaline band, by the spot in M₂–M₃ and by the apical spots being clearly larger. In the male genitalia, there is a slight variation in the distal tip of the harpe, apparently a little wider in *Zonia zonia panamensis*, judging by the drawings of the original description.

It is one of the most poorly represented species in collections. *Zonia zonia zonia* is known from the holotype male from Tefé, Amazonas, Brazil, one male from Quebrada Quitacalzón Cosñipata Valley, Cusco, Peru, one male from Guyana, one male and one female from Juruti and one female from Santarém, both in Pará, Brazil; of *Z. zonia panamensis*, there are only the male holotype, plus four males paratypes from Madden Forest, Panamá, Panama and one female from Honduras; and of *Z. zonia diabo* two males from Pirenópolis, Goiás, 16 males (including the holotype) from the Parque Estadual do Morro do Diabo, Teodoro Sampaio, São Paulo, four males from the Parque Municipal Três Morrinhos, Terra Rica, Paraná, and one male from Foz do Iguaçu, Paraná, Brazil are known, while the female is unknown. In all localities *Zonia zonia diabo* specimens were collected hilltopping, resting occasionally between 11:00 and 13:00 hours, with exception of the male from Foz do Iguaçu encountered dead. It is probably a typical canopy species.

ETYMOLOGY – The specific name is in honour of our friend Heinz Ebert, a geologist of the University of São Paulo, Rio Claro, São Paulo, Brazil, who dedicated much of his time to butterflies.

REMARKS – Only known from three specimens.

TYPES:

Holotype male with the following labels: / Holotypus/ Casimiro de Abreu, [Estado do] R.[io] Jan.[eiro], [Brazil], 28-II-[19]75, [H.] Ebert [leg.]/ gen. prep. [O.] Mielke 1980/ DZ 1828/ 2487/ Holotypus *Zonia zonia eberti* O. Mielke, C. Mielke & Casagrande. O. Mielke, C. Mielke & Casagrande det. 2017/. DZUP. The month of capture is illegible.

Allotype female with the following labels: / Allotypus/ 3-X-1989 Barra de S.[ão] João, [Estado do] Rio de Janeiro, [Brazil], K. Brown leg., OM 37.316/ Allotypus *Zonia zonia eberti* O. Mielke, C. Mielke & Casagrande det. 2017/. OM-DZUP.

Paratype: 1 ♂, Santa Catarina, Brazil, OM 13.143 (OM).

OXYNETRINI O. Mielke, 2001

Three genera.

TAXONOMY – The tribe is characterized by the genome (Zhang *et al.* 2019) and by some morphological characters: forewing with the discocellular veins directed to tornus, not parallel to the outer margin; Sc absent or reduced in the forewing; anterior projection of saccus larger than long; gnathos ventrally rounded; distal third of aedeagus depressed; vesica without cornutus; fultura inferior conical, with open base directed distally; lamella postvaginalis more or less rectangular, with the posterior edges rounded and the anterior edges concave.

Olafia Nemésio, 2005

Syn.: *Cyclopyge* O. Mielke, 2002, *praeocc.* (Hafle & Corda, 1847) (Trilobita)

One species.

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Olafia roscius (Hopffer, 1874)

Three subspecies.

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Olafia roscius roscius (Hopffer, 1874)

Pls. 14–15, Figs. 1 (lectotype ♂ of *Pyrrhopyge roscius*), 2, 5–6 (♂), 3, 7 (♀) (d, v)

Syn.: *Oxynetra rufocincta* Hayward, 1932

Oxynetra flavomaculata Bell, 1937, **syn. n.**

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Brazil: São Paulo (*São José do Barreiro* – Parque Nacional da Serra da Bocaina); Paraná (*Campina Grande do Sul, Candói, Castro, Curitiba, Guarapuava, Inácio Martins, Lapa, Londrina, Morretes* – Alto da Serra 800 m, *Palmas, Palmeira, Piên, Ponta Grossa, Quatro Barras, São José dos Pinhais, Tibagi, Tijucas do Sul, Turvo, Tuneiras do Oeste, Turvo, Ventania*); Santa Catarina (*Dalbergia, Fraiburgo, Joaçaba, Joinville, Lages, between Massaranduba – Blumenau, Otacílio Costa, São Bento do Sul – Rio Vermelho, São Joaquim, Seara – Nova Teutônia, Urubici*); Rio Grande do Sul (*Bom Jesus, Cambará do Sul, Canela, Central Region, Pelotas, Vacaria*). **Paraguay:** Paraguari (*Sapucaí*). **Argentina:** Misiones (*Dos de Mayo, General Belgrano, Guaraní, Oberá, Puerto Iguazú, Puerto Libertad*).

The citation **Colombia** (Hayward 1947) is incorrect.

TAXONOMY – The authors have collected in the same place, *Olafia roscius roscius* and *Olafia roscius flavomaculata*, previously considered as distinct subspecies by Evans (1951) (as *Oxynetra*) and O. Mielke (2004, 2005) (as *Cyclopyge*), in Santa Catarina (*São Bento do Sul – Rio Vermelho 850 m, Urubici*) and Paraná (*São José dos Pinhais, Tibagi, Ventania*), and Rio Grande do Sul (*Cambará do Sul*). Their sympatry suggests synonymy of *Oxynetra flavomaculata* Bell, 1937, with *Pyrrhopyge roscius* Hopffer, 1874.

TYPES:

Pyrrhopyge roscius Hopffer, 1874. The original description mentions an unspecified number of males. To ensure the correct identification of *Pyrrhopyge roscius*, one syntype female (not male) is **herein designated as the lectotype**; this specimen with the following labels: / 4896/ Brasil. Sello/ *Roscus* Hpfr*/ Grosse Analogie mit “*Sarbia*” *erythrosoma* Mab. 11:8/ 1:19/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge roscius* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke det. 2019 / . MfN.

Oxynetra rufocincta Hayward, 1932. Holotype male with the following labels: / Iguazu, [Misiones], Argentina, A. Breyer X-1928/ *Oxynetra rufocincta* nov. Holotipo, det Hayward/ Tipus, 2559/ N° 31/. MLP.

Oxynetra flavomaculata Bell, 1937. Holotype male with the following labels: / Neu Bremen [= Dalbérgia] Sta [Santa] Cath. [Catarina] Mch [March] [19]27 Brazil/ G 1036/ Type/ *Oxynetra flavomaculata* Bell Holotype ♂/. AMNH. **Syn. n.**

Olafia roscius koikei O. Mielke, Brockmann & C. Mielke, **ssp. n.**

Pls. 14–15, Figs. 4 (holotype ♂) (d, v)

DISTRIBUTION – Southeastern Brazil (Rio de Janeiro).

Brazil: Rio de Janeiro (*Nova Friburgo – Pico Caledônia 2,200 m*).

DESCRIPTION – Male: forewing length 23 mm. Similar to *Olafia roscius roscius* but lacking any spot on the forewing. Differs from this subspecies by the light yellow scales at the extreme base on the hindwing ventrally, as in *Olafia roscius iphimedia*. The genitalia don't

differ from those of *Olafia roscius roscius*.

Female: unknown.

REMARK – On the dorsal hindwing the scales at the apex are lost (Pl. 14, Fig. 4).

ETYMOLOGY – The subspecific name is in honour of our friend Ricardo Koike, who collected the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ Brazil – Rio de Janeiro (RJ) Nova Friburgo, Pico Caledonia, 2,200 m, 18.I.1998. R. Koike leg. 22°21'12.16"S, 42°35'11.27"W/ gen[itália]. prep[arada]., [O.] Mielke 2018/ Holotypus *Olafia roscius koikei* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/ DZ 20.931/. DZUP.

Olafia roscius iphimedia (Plötz, 1886)

Pls. 14–15, Figs. 9 (lectotype ♂ of *Pyrrhopyga erythrosoma*), 10 (lectotype ♂ of *Pyrrhopyga iphimedia*), 11 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *erythrosoma* Mabilles, 1891

DISTRIBUTION – Southeastern Brazil.

Brazil: Minas Gerais (*Caldas, Conceição dos Ouros, Diamantina, Delfim Moreira, Poços de Caldas*); Rio de Janeiro (*Resende*); São Paulo (*Araras, Campos do Jordão, Jundiá, Piquete – Barreira de Piquete, Rio Claro, São Paulo*).

The citations **Brazil:** Amazonas (Mabilles 1903, Mabilles & Boulet 1908, Mabilles 1912, Draudt 1921, Lewis 1973, 1975) are incorrect.

TYPES:

Pyrrhopyga [**sic**] *iphimedia* Plötz, 1886. The original description mentions an unspecified number of specimens from Brazil. No syntype was found in MfN or ZSM, where nearly all Plötz's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Pyrrhopyga iphimedia*, mentioned as *Oxynetra roscius iphimedia* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ 15-II-1984, Barreira de Piquete, [Piquete], S[ão] Paulo [Brazil] 1400-1600 m., [O.] Mielke & Casagrande leg./ Neotypus *Pyrrhopyga iphimedia* Plötz, 1886 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.445/. DZUP.

Pyrrhopyga [**sic**] *erythrosoma* Mabilles, 1891. The original description mentions an unspecified number of males from St. [São] Paulo, [Brazil]. To ensure the correct identification of *Pyrrhopyga erythrosoma*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ St. Paulo [Brazil] Littk./ Coll. Staudinger/ *P. erythrosoma* Mab. ♂/ *Erythrosoma* Mab./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga erythrosoma* Mabilles, 1891, O. Mielke, Brockmann & C. Mielke det. 2019 / . MfN.

Oxynetra C. Felder & R. Felder, 1862

Two species.

DISTRIBUTION – Panama to central-eastern Bolivia, northwestern Venezuela and northwestern Brazil.

Oxynetra confusa Staudinger, 1888

Two subspecies.

DISTRIBUTION – Panama, central and eastern Colombia to central-eastern Bolivia, northwestern Venezuela and northwestern Brazil.

Oxynetra confusa confusa Staudinger, 1888

Pls. 14–15, Figs. 13 (lectotype ♂), 14 (♂), 15 (♀) (d, v)

DISTRIBUTION – Central and eastern Colombia to central-

eastern Bolivia, northwestern Venezuela and northwestern Brazil (Acre and Rondônia).

Colombia: Cundinamarca (*Chirajara, Llanos de San Martín, Fomeque, Quetamo*); Meta (*Villavicencio*). **Ecuador:** Morona-Santiago (*Macas*); Napo (*Misahualli, Puerto Napo, Río Shingupino*); Pastaza (*Canelos, Mera*). **Peru:** Amazonas (*Pongo del Rentema, Rodríguez de Mendoza*); Cusco (*Illapani Viejo, Quincemil*); Huánuco (*Carpish, Río Monzón, Tingo María*); Junín (*Chanchamayo, La Merced, Perené, Río Colorado, Satipo, Shimá*); Loreto (*Pebas*); Madre de Dios (*Puerto Maldonado, Shintuya, Tambopata Reserve*); Pasco (*Pozuzo*); San Martín (*Juanjuí, Rioja, Tarapoto, Yumbatos*); Puno (*Yahuar Mayo*); Yumbatos. **Bolivia:** Cochabamba (*Chapare*); La Paz (*Caranavi, Río Zongo*); Santa Cruz (*Buenavista, Sara, Velasco*). **Venezuela:** Barinas (near *Barinitas*); Táchira (*Las Cuevas, Loma de Pio, Río Frío, San Cristóbal, Via Chorro del Indio*). **Brazil:** Acre (*Upper Río Juruá*); Rondônia (*Cacaulândia, Jarú*).

The citations **Argentina** (Santiago del Estero) and **French Guiana** (Evans 1951) are probably from mislabelled specimens.

TYPE:

Oxynetra confusa Staudinger 1888. The original description mentions an unspecified number of males from Chanchamayo, [Junín, Peru] in collection Staudinger. To ensure the correct identification of *Oxynetra confusa*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ *Confusa* Stgr./ Chanchamayo Thamm 85/ Coll. Staudinger/ *Confusa* Stgr. *Semihyalina* Feld. ♂/ 18: 4/; two other labels will be added: / Lectotypus/ Lectotypus *Oxynetra confusa* Staudinger, 1888, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Oxynetra confusa franciscana Salazar & Vargas, [2017]

Pls. 14–15, Figs. 16 (paratype ♂) (d, v)

DISTRIBUTION – Panama.

Panama: Darién (*Garachiné – Cerro Sapo and Pinoguilla*).

REMARKS – Only known from the type locality.

TYPE:

Oxynetra confusa franciscana Salazar & Vargas, [2017]. Holotype male from Panama, Darién, Cerro Sapo – Garachiné, 9-II-2010, 1,000 m, F. S. Delgado leg., 20574. CJS.

Oxynetra semihyalina C. Felder & R. Felder, 1862

Pls. 14–15, Figs. 17 (♂), 18 (♀) (d, v)

Syn.: *Oxynetra felderi* Hopffer, 1874

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia.

Colombia: Boyacá (*Muzo*); Cundinamarca (*Medina*); Meta (*Villavicencio*). **Ecuador:** Bolívar (*Balzapamba*); Napo; Pastaza (*Mera, Río Pastaza*); Morona-Santiago (*Macas*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Rodríguez de Mendoza, Alto Nieva 1,930 m*); Cajamarca (*Charapi*); Cusco (*Callanga, Cosñipata Valley – Quebrada Quitacalzón 1,050 m to San Pedro 1,400 m, Machu Picchu, Río Pampaconas*); Huánuco (*Carpish, Puente Cayumba, Tingo María*); Junín (*Andamarca, Chanchamayo, La Merced, Puhanki, Río Colorado, Río Perené, Satipo, Shimá*); Loreto (*Pebas*); Madre de Dios; Pasco (*Huancabamba, Pozuzo*); Puno (*Uruhasi*); San Martín (*Jorge Chávez, Juanjuí, Moyobamba, Rioja, Santa Cruz 1,200 m*); Ucayali (*road Previsto – Aguaytia*). **Bolivia:** Cochabamba (*Chapare, San Jacinto*); La Paz (*Caranavi, Chulumani, Coroico, Río Zongo*).

The citations **Mexico** (Hopffer 1874, Kirby 1877, Hoffmann 1941, 1976), **Venezuela** (Hopffer 1874) and **Brazil:** Amazonas (*Rio Negro*) (C. Felder & R. Felder 1862, Kirby 1871, Hopffer 1874, Bell 1934b, 1946) and Acre (*Upper Juruá River*) (Bell 1934b) are probably incorrect. The citation **Venezuela** (Kirby 1877) probably refers to *Oxynetra confusa* (Orellana [2010]).

There is a mislabelled specimen from **Brazil:** Pará (*Óbidos*) in the collection of the American Museum of Natural History, New York,

USA.

TYPES:

Oxynetra semihyalina C. Felder & R. Felder, 1862. Holotype (one female [**recte** male] is mentioned) with the following labels: / Type H T/ 361/ Rio Negro type/ *Oxynetra semihyalina* Feld./ *semihyalina* n./ Rothschild Bequest B. M. 1939-1./ NHMUK. The type locality “fluminis Negro superioris in Brasilia septentrionalis” refers probably to the Río Negro, near Rioja, San Martín, Peru (see Lamas 1976).

Oxynetra felderi Hopffer, 1874. The original description mentions an unspecified number of males from Rio Negro, [Amazonas, Brazil] and Chanchamayo, [Junín, Peru]. To ensure the correct identification of *Oxynetra felderi*, one syntype male is **herein designated as lectotype**; this specimen with the following labels: / Type/ 17788/ *Felderi* Hpfr* ? *Semihyalina* Feld W. [??] 1862 9/ Chanchamayo [Junín, Peru] Tham/; two other labels will be added: / Lectotypus/ Lectotypus *Oxynetra felderi* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke 2014/. MfN.

Dis Mabilles, 1889

Three species.

DISTRIBUTION – Eastern Mexico to Panama.

***Dis stangelandi* (Grishin & Burns, 2013)**

Pls. 14–15, Figs. 20 (holotype ♂), 24 (paratype ♀) (d, v)

DISTRIBUTION – Costa Rica.

Costa Rica: Guanacaste (*Área de Conservación Guanacaste*); Puntarenas (*Monteverde*).

REMARKS – Only known from two localities.

TYPE:

Oxynetra stangelandi Grishin & Burns, 2013. Holotype male from Costa Rica, Alajuela, Área de Conservación Guanacaste, Sector Cacao, Sendero Derrumbe, 1,220m, lat 10.92918, long -85.46426, Harry Ramirez leg. USNM.

***Dis hopfferi* (Staudinger, 1888)**

Pls. 14–15, Figs. 19 (holotype ♂), 22 (♀), 23 (holotype ♀ of *Dis annulatus*) (d, v)

Syn.: *Dis annulatus* Mabilles, 1889

DISTRIBUTION – Costa Rica to Panama.

Costa Rica: Guanacaste (*Área de Conservación Guanacaste*); Puntarenas (*Coto Brus, Monteverde*); Heredia (*San José de la Montaña*).

Panama: Chiriquí (*Santa Clara*).

The citations **Brazil:** Amazonas (Mabilles 1903, Mabilles & Boullet 1908) are probably incorrect, and the citations **Mexico:** Veracruz (*Motzorongo*) (Hoffmann 1941), Veracruz (*Los Tuxtlas*) (Raguso & Llorente 1997) and Chiapas (Maza E., R. G. de la & Maza E., J. de la 1993, Austin *et al.* 1999) probably refer to *Oxynetra aureopecta*.

TYPES:

Oxynetra hopfferi Staudinger, 1888. Holotype (one male is mentioned) with the following labels: / Origin./ Chiriqui [Panama] Ribbel/ *O. hopfferi* felderi Hopf var. of *semihyalina* Feld/ 18: 3./ Coll. Staudinger/. MfN.

Dis annulatus Mabilles, 1889. Holotype (one female is mentioned) with the following labels: / Origin./ *Dis annulatus* Mab. ♀/ *Annulatus* Mab./ *D. annulatus*/ ist wohl ♀ zu 1, nicht zu 4/ Chiriqui [Panama] William Jr./ Coll. Staudinger/. MfN.

***Dis aureopecta* (Warren & Grishin, 2017)**

Pls. 14–15, Figs. 12 (holotype ♂), 21 (♀) (d, v)

DISTRIBUTION – Eastern Mexico and El Salvador.

Mexico: Chiapas (?); Hidalgo (*Puerto del Caballo*); Veracruz (*Los Tuxtlas, Motzorongo, Presidio*). **El Salvador:** Ahuachapán (*Tacuba*).

TYPE:

Oxynetra aureopecta Warren & Grishin, 2017. Holotype male with the following labels: / Puerto del Caballo, Hidalgo, Mexico Sept[ember] 8, [19]’87/ William W. Howe collector/ Holotype ♂ *Oxynetra aureopecta* A. Warren & Grishin/ DNA sample ID: NVG-14113A02/. LACM.

PYRRHOPYGINI Mabilles, 1877

Four subtribes.

TAXONOMY – The tribe is characterized by the genome (Zhang *et al.* 2019) and by some morphological characters: gnathos normally of either two separated pieces, or V-shaped with the union prolonged distally – only *Nosphistia zonara* (Hewitson, 1866); fultura inferior either large, more than five times the diameter of aedeagus, or forming a ring around the aedeagus; vesica without cornutus or, if present, smaller than the opening of the aedeagus (absent also in Oxynetrini); lamella post-vaginalis membranous, or represented as a weakly sclerotized area of the diaphragm, or as a thick membranous fold – only *Mysarbia sejanus* (Hopffer, 1874), or sclerotised and nearly rectangular, with the proximal and external corners expanded and with a concavity on the median posterior margin – only *Aerardaris aerata* (Godman & Salvin, 1879).

PYRRHOPYGINA (Mabilles, 1877) Warren, Ogawa & A. Brower, 2008

Three genera.

TAXONOMY – The subtribe is characterized by the genome; no morphological characters are known (Grishin *in* Zhang *et al.* 2019).

***Pyrrhopyge* Hübner, [1819]**

Syn.: *Tamyris* Swainson, 1821

Pachyrhopala Wallengren, 1858

Seventy-one species, the most speciose genus.

DISTRIBUTION – Mexico to southern Brazil, Paraguay, northeastern Argentina and Uruguay.

***Pyrrhopyge guianae* Bell, 1932**

Pls. 16–17, Figs. 1 (holotype ♂) (d, v)

DISTRIBUTION – Guyana and French Guiana.

Guyana: Essequibo Islands – West Demerara (*Parika*). **French Guiana.**

TYPE:

Pyrrhopyge guianae Bell, 1932. Holotype male with the following labels: / French Guiana/ Type *Pyrrhopyge guianae* E. L. Bell 7754/ *Pyrrhopyge guianae* Bell Holotype ♂/ Genitalia slide No. 1979/ Slide Phil. 33./ ANSP, transferred to CMP.

***Pyrrhopyge zenodorus* Godman & Salvin, 1893**

Pls. 16–17, Figs. 2, 4 (♂), 3 (♀) (d, v)

DISTRIBUTION – Central Mexico to Costa Rica.

Mexico: Chiapas (*Boca de Chajul, Chajul, Comitán de Domínguez* – *Santa Rosa, San Antonio*); Mexico (*Tabasco*); Oaxaca (*Candelaria Loxicha, Chimalapa, Comaltepec, Sierra de Juárez, Santa María Yacochi, Sierra de Mazateca, Usila*); Puebla (*Zihuatéutla*); Quintana Roo; Veracruz (*Atayac, Coatepec, Córdoba, Fortín, Los Tuxtlas, Mozorongo,*

Presidio, Rinconada, Tezonapa, Xalapa, Zongolica); Yucatán Peninsula. **Belize:** Belize (*San Pedro, Stann Creek*). **Guatemala:** Alta Verapaz (*Cahabón, Chejel, Cubilguitz, Polochic Valley*); Guatemala (*Guatemala*); Zacapa (*La Unión*). **Honduras.** **Nicaragua:** Managua; Matagalpa (*Selva Negra*). **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste*); Limón (*Guapiles*); Puntarenas (*Osa*); San José (*Cantón de Mora – El Rodeo, Colón, Palmichal, Puriscal, San Isidro de El General, Santa Ana*).

The citations **Venezuela** (Bell 1946, Schmid & Endicott 1968) are probably incorrect (Orellana [2010]). The citations **Argentina:** Misiones (Hayward 1938, 1941, 1948) are incorrect.

TYPE:

Pyrrhopyge zenodorus Godman & Salvin, 1893. The original description mentions an unspecified number of males and females from Mexico, Coatepec and Rinconada, Schaus leg., Cordova, Rümeli leg., Atoyac, H. H. Smith leg.; Guatemala, Cubilguita, Cahabón, Champion leg., Polochic Valley, O. Salvin & F. D. Godman leg.; Costa Rica, van Patten leg. and E. Peru. To ensure the correct identification of *Pyrrhopyge zenodorus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Polochic Valley, [Alta Verapaz, Guatemala], F. D. G. & O. S./ B.C.A. Lep. Rhop. *Pyrrhopyge zenodorus* G. & S./ Godman-Salvin Coll. 1912-23./ Sp. figured/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge zenodorus* Godman & Salvin, 1893, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge evansi Bell, 1947

Two subspecies.

DISTRIBUTION – Northwestern Mexico to western Ecuador and to central and northern Colombia, eastern Ecuador, northern and eastern Venezuela to French Guiana.

Pyrrhopyge evansi evansi Bell, 1947

Pls. 16–17, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Northwestern Mexico to western Ecuador and central and northern Colombia, eastern Ecuador and eastern Venezuela to French Guiana.

Mexico: Durango. **Costa Rica:** Puntarenas (*San Vito*). **Panama:** Chiriquí (*Lino*); Colón (*Barro Colorado Island, Gatun Lake*); Panamá (*Ancon, Corozal, Summit*); Panamá Oeste (*Cerro Campana*); Veraguas (*Isla Coiba*). **Colombia:** Arauca; Boyacá (*Muzo, Oranche*); Cundinamarca (*Cananche, Fusagasugá*); Santander (*Barbosa, Contratación*); Tolima (*La Aurora*); Valle del Cauca (*Calima*). **Ecuador:** Cañar (*La Troncal*); Carchi (*Río Baboso*); Esmeraldas (*Chuchuvi, Cotacachi, Durango, Lita, Pihuambi, San Francisco*); Imbabura (*Lita, Paramba*); Pastaza (*Puyo*); Pichincha (*Nanegalito, Pacto, San Miguel de los Bancos*); Zamora-Chinchipe (*Zamora*). **Venezuela:** Bolívar (*El Dorado*); Delta Amacuro (*Río Grande*). **French Guiana:** Cayenne (*Camp Patawa, Mont Grand Matoury, Coralie*).

TYPE:

Pyrrhopyge evansi Bell, 1947. Holotype male with the following labels: / Holotype/ Muzo, [Boyacá] Col[ombia], IV-1945/ G1837/ *Pyrrhopyge evansi* Bell Holotype ♂/. AMNH.

Pyrrhopyge evansi borbura Orellana, [2010]

Pls. 16–17, Figs. 7 (paratype ♂), 8 (paratype ♀) (d, v)

DISTRIBUTION – Northern Venezuela.

Venezuela: Carabobo (*Río Borburata, San Esteban*).

REMARKS – Only three specimens are known.

TYPE:

Pyrrhopyge evansi borbura Orellana, [2010]. Holotype male from San Esteban, Carabobo, Venezuela, 12-IX-1953, F. Fernández-Y. leg. MIZA.

Pyrrhopyge phidias (Linnaeus, 1758)

Eight subspecies.

DISTRIBUTION – Northern, western and eastern Colombia to central-eastern Bolivia, Trinidad and Tobago, Venezuela to French Guiana and northern and northeastern to southeastern Brazil.

TAXONOMY – Three subspecies, *P. phidias phidias* (n= 4), *P. phidias leucoma* (n= 1) and *P. phidias rusca* (n= 2), with less than 1% difference in the DNA barcode, fly together in the Cosñipata Valley, Cusco, Peru, between 675 m and 1,400 m, *P. phidias phidias* and *P. phidias rusca* in Tingo Maria, Huánuco, Peru, and *P. phidias phidias* and *P. phidias leucoma* in the Yungas, La Paz, Bolivia; probably they are only different phenotypes.

REMARKS – Red fore coxae, so far only observed in subspecies *P. phidias bixae* (Pl. 17, Fig. 24).

Pyrrhopyge phidias phidias (Linnaeus, 1758)

Pls. 16–17, Figs. 9–11 (♂), 12 (♀), 13 (neotype ♂ of *Papilio phidias* and neotype ♂ of *Hesperia zealeucus*) (d, v)

Syn.: *Hesperia zealeucus* Fabricius, 1793

Pyrrhopyge williamsi Bell, 1931

DISTRIBUTION – Northern, western, central and eastern Colombia to central-eastern Bolivia, Trinidad and Tobago, Venezuela to Suriname and northern Brazil (Pará and Mato Grosso).

Colombia: Arauca; Boyacá (*Muzo*); Central Cordillera (*east side*); Cesar (*Chimichagua*); East Cordillera (*east side*); Meta (*Remolino, Villavicencio*). **Peru:** Cusco (*Cosñipata Valley – Chontachaca 950 m to San Pedro 1,400 m*); Huánuco (*Tingo Maria*); Madre de Dios (*Puerto Maldonado*); Puno (*Sandia*). **Bolivia:** La Paz (*Caranavi*); Santa Cruz (*Sara*). **Venezuela:** Amazonas (*Puerto Ayacucho, San Carlos de Río Negro, Yavita*); Aragua (*Choroní, Maracay, Parque Nacional Henry Pittier – Rancho Grande, Sabana Grande – vía a Belén, Tasajera, Via de Tiara – Loma de Hierro, Villa de Cura*); Barinas (*San Isidro*); Bolívar (*road El Dorado – Santa Elena km 17, Maripa*); Carabobo (*carretera Tacarigua – Los Naranjos km 14, Cerro de Paja, Güigüe*); Distrito Capital (*Caracas, El Junquito, Loma del Muerte, Los Venados*); Falcón (*Sierra de San Luis*); Guárico (*Altagracia de Orituco*); Lara (*Iribarren, Terepaima, Parque Nacional Yacambú*); Miranda (*Cortada del Guayabo, Parque Nacional Guatopo*); Monagas (*Sabana de Piedra, Jusepin, La Pica*); Táchira (*Cárdenas, Delicias, La Parada, Los Naranjos, Parque Nacional Chorro del Indio, Río Frio, Rubio*); Trujillo (*Río Negro – Los Pozuelos*); Yaracuy (*Cerro Zapatero, Nirgua*); Zulia (*El Rosado*). **Trinidad and Tobago:** Port of Spain (*St. Ann's Valley*). **Guyana:** Demerara-Mahaica (*Soesdyke*). **Suriname:** Brokopondo (*Brownsberg*); Para (*Zuid*); Paramaribo; Sipaliwini (*Suriname*). **Brazil:** Pará (*Juruti*); Mato Grosso (*Diamantino*).

The citations **Brazil:** Paraná and **Paraguay** (Evans 1951) are probably incorrect (as *P. phidias williamsi*).

TYPES:

Papilio phidias Linnaeus, 1758. The original description mentions an unspecified number of specimens from Asia in M.L.U. [Musei Ludovicae Ulrica]. A lectotype male was designated by Honey & Scoble (2001) in UZIU. As this lectotype designation is based on a specimen of the “*bixae* form” of this species (ventral hindwing with a white band) not included in the original description, and not of the “*phidias* form” (ventral hindwing without a white band) as described by Linnaeus, the designation must be considered as invalid (ICZN, Art. 73.2). See also Evans (1940) for the correct identification of *Papilio phidias*. There is no other specimen that agrees with the original description of *Papilio phidias* in the collection Linnaeus, currently in UZIU and LSL. To ensure the correct identification of *Papilio phidias*, mentioned as *Pyrrhopyge phidias phidias* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ 22-III-1981 30 km E de Villavicencio, Meta, Colombia, Callaghan leg./ gen[italia]. pre[arada]., [O.] Mielke 2014/ Neotypus *Papilio phidias* Linnaeus, 1758 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 37.170/. OM-DZUP.

Hesperia zealeucus Fabricius, 1793. The original description

mentions an unspecified number of specimens from Indiis in collection Drury. No syntypes were found in the above mentioned museums and collections (see “Abbreviations of the collections”). To ensure the correct identification of *Hesperia zealeucus*, mentioned as a synonym of *Pyrrhopyge phidias phidias* by Evans (1951), a male is **herein designated as the neotype**; the specimen is the same as the neotype of *Papilio phidias* Linnaeus, 1758 and the following other label will be added: / Neotypus *Hesperia zealeucus* Fabricius, 1793 O. Mielke, Brockmann & C. Mielke det. 2015/. OM-DZUP.

Pyrrhopyge williamsi Bell, 1931. Holotype male with the following labels: / Peru/ Type/ G423/ 5/ Holotype ♂ *Pyrrhopyge williamsi* Bell/. AMNH.

***Pyrrhopyge phidias garata* Hewitson, 1866**

Pls. 16–17, Figs. 14 (♂), 15 (♀) (d, v)

DISTRIBUTION – Guyana and Suriname.

Guyana: Demerara-Mahaica (*Demerara River, Georgetown*); Pomeroon-Supenaam (*Mocha*); Upper Demerara-Berbice (*Linden, Omai*). **Suriname:**

The citation **Venezuela** (Evans 1951) is probably incorrect (Orellana [2010]).

TYPE:

Pyrrhopyga [**sic**] *garata* Hewitson, 1866. The original description mentions an unspecified number of specimens from Suriname in collection Hewitson. To ensure the correct identification of *Pyrrhopyga garata*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Type H T/ Surinam. Hewitson Coll. 79-69. *Pyrrhopyga garata*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga garata* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge phidias latifasciata* Butler, 1873**

Pls. 16–17, Figs. 16–17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Western, central and eastern Colombia.

Colombia: Caldas (*Manizales*); Chocó (*San Jose del Palmar*); Cundinamarca (*Bogotá, Medina*); Meta (*Upper Río Negro, Villavicencio*); Putumayo (*Mocoa, Puerto Leguizamo*); Valle del Cauca (*Buenaventura*).

TYPE:

Pyrrhopyga [**sic**] *latifasciata* Butler, 1873. The original description mentions an unspecified number of specimens from Bogotá, [Cundinamarca, Colombia] in NHMUK. To ensure the correct identification of *Pyrrhopyga latifasciata*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *latifasciata* Butl. Type/ Bogota [Cundinamarca, Colombia] Stevens 56-142./ Genitalia Slide No. [?]/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga latifasciata* Butler, 1873, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge phidias rusca* Evans, 1951**

Pls. 16–17, Figs. 19 (♂), 20 (♀), 21 (holotype ♂) (d, v)

DISTRIBUTION – Eastern Ecuador to southeastern Peru.

Ecuador: Morona-Santiago (*Macas, San Isidro, Santiago de Méndez*); Napo (*Cotundo, Misahualli*); Pastaza (*Mera*); Zamora-Chinchipe (*Zamora*); Tungurahua (*Palora*). **Peru:** Amazonas (*Huambo, Santa Maria de Nieva*); Cajamarca (*Charapi, Manchara*); Cusco (*Cosñipata Valley – Pilcopata 500 m to San Pedro 1,400 m*); Huánuco (*Tingo Maria*); Madre de Dios (*Shintuya – 3.3 km N, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco), Salvación*); Pasco (*Pichis Road*); Puno (*Carabaya, Río Inambari*); San Martín (*Moyobamba, Rioja – San Agostin 1,000 m to Jorge Chávez 1,450 m, Santa Cruz, Tarapoto, Tocache*).

TYPE:

Pyrrhopyge phidias rusca Evans, 1951. Holotype male with the following labels: / Type – *rusca* Evans/ Perou [Amazonas] Huambo M. de Mathan IV^e. Trim. 1889/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Pyrrhopyge phidias bixae* (Linnaeus, 1758)**

Pls. 16–17, Figs. 22, 24 (♂), 23 (♀) (d, v); Pls. 18–19, Figs. 1, 10 (♂), 2 (♀) (d, v)

Syn.: *Papilio maenas* Fabricius, 1787

Telegonus meriani Kirby, 1871; **nom. n.**

DISTRIBUTION – Southeastern Colombia to southeastern Peru, French Guiana and northern Brazil.

Colombia: Amazonas (*La Pedrera, Leticia*). **Peru:** Loreto (*Contamana, Iquitos, Pebas*); Madre de Dios (*Tambopata Reserve*). **Suriname:** Para (*Colacreek, Zanderj*). **French Guiana:** Cayenne (*Camp Patawa, Crique Mortio, Montagne Favard, Montagne de Kaw*); Saint-Laurent-du-Maroni. **Brazil:** Amapá (*Serra do Navio*); Amazonas (*Benjamin Constant, Fonte Boa, Itacoatiara, Manaus, Manicoré, Maraã, Maués, Nhamundá, Novo Airão, Parintins, Rio Negro, Rio Preto da Eva, São Paulo de Olivença, Tonantins*); Pará (*Ilha de Marajó, Juruti, Óbidos, Rio Arapiuns, Santarém – Fazenda Taperinha*); Acre (*Cruzeiro do Sul, Senador Guiomard, Xapuri*); Rondônia (*Ariquemes, Cacaulândia, Candeias do Jamari, Rio Crespo*); Mato Grosso; Maranhão.

TAXONOMY – The lectotype of *Papilio maenas* was not examined. So, the identification and the synonymy are dubious.

TYPES:

Papilio bixae Linnaeus, 1758. Lectotype male (Hemming 1964 and O. Mielke 1989b) from Suriname, without labels in collection Maria Sibylla Merian. SMW.

Papilio maenas Fabricius, 1787. The original description mentions an unspecified number of specimens from America in collection Dr. Hunter. Two females were found in HMUG, one with and another without a head. To ensure the correct identification of *Papilio maenas*, the syntype female without head is **herein designated as the lectotype** (no labels were examined); two other labels will be added: / Lectotypus/ Lectotypus *Papilio maenas* Fabricius, 1787, O. Mielke, Brockmann & C. Mielke det. 2019/. HMUG.

Telegonus meriani Kirby, 1871 is a **nomen novum** for *Papilio bixae* Linnaeus, 1758.

***Pyrrhopyge phidias hyperici* Hübner, 1823**

Pls. 18–19, Figs. 3 (lectotype ♂ of *Pyrrhopyga hyleus*), 4, 7, 9 (♀), 5 (neotype ♂), 6 (♂) (d, v); Text-Fig. 8 (♂ genitalia)

Syn.: *Pyrrhopyge hyperici* Hübner, [1819], **nom. nud.** (ICZN, Art. 12.1)

Pyrrhopyga [**sic**] *hyleus* Mabille, 1891, **syn. n.**

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (*Acará, Belém, Benevides, Cametá, Itaituba, Marituba, Murutucum, Novo Progresso, Santo Antônio de Tauá*).

TAXONOMY – *Pyrrhopyga* [**sic**] *hyleus* was mentioned as a synonym of *Pyrrhopyge proculus cardus* Mabille, 1891 (Evans 1951) and as a subspecies of *Pyrrhopyge proculus* (O. Mielke 2004, 2005); but the dissection of the male genitalia of the lectotype of *Pyrrhopyga* [**sic**] *hyleus* (Fig. 9) reveals that it is synonymous with *Pyrrhopyge phidias hyperici*.

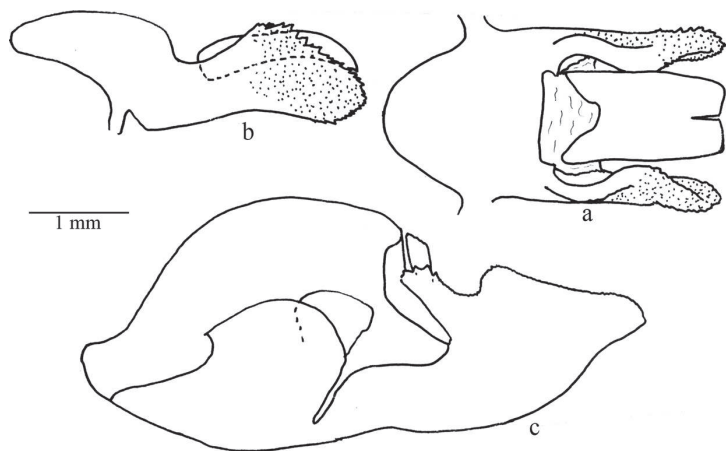
TYPES:

Pyrrhopyge hyperici Hübner, [1819]. There is no description; a **nomen nudum** (ICZN, Art. 11.1)

Pyrrhopyge hyperici Hübner, 1823. The original description mentions an unspecified number of specimens from Brazil. No syntypes were found in MfN, where a few Hübner's types deposited, or in any museum or collection mentioned above (see “Abbreviations of the collections”). To ensure the correct identification of *Pyrrhopyge hyperici*, mentioned as *Pyrrhopyge phidias hyperici* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ V-1965 Belém Pará [Brazil]/ gen[italia]. prep[arada]., [O.] Mielke 2014/ Neotypus *Pyrrhopyge hyperici* Hübner, 1823 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 10.843/. OM-DZUP.

Pyrrhopyga [**sic**] *hyleus* Mabille, 1891. The original description mentions an unspecified number of males from Itaituba, [Pará], Brazil.

To ensure the correct identification of *Pyrrhopyga hyleus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Itaituba Tapajos, [Pará, Brazil] 86 Hhl./ Coll. Staudinger/ *P. Hyleus* Mab./ *Hylaeus* Mab./ gen. prep. [O.] Mielke 1979/ Genital-Unters. Nr. 4723, Zool. Mus. Berlin/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga* [**sic**] *hyleus* Mabilie, 1891. O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. **Syn. n.**



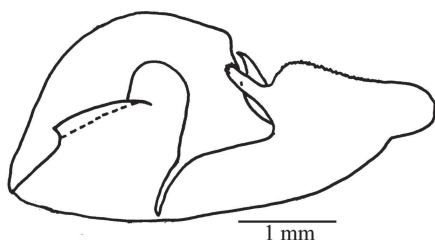
Text-Fig. 8. *Pyrrhopyge phidias hyperici*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen and uncus, lateral left view; c) right valva, internal view (lectotype of *Pyrrhopyge hyleus*).

Pyrrhopyge phidias aldo* O. Mielke, Brockmann & C. Mielke, *ssp. n.

Pls. 18–19, Figs. 11 (holotype ♂), 12 (allotype ♀), 13 (paratype ♂), 14 (paratype ♀) (d, v); Text-Fig. 9 (♂ genitalia)

DISTRIBUTION – Northeastern to southeastern Brazil.

Brazil: Maranhão (*Mirador – Parque Estadual Mirador*); Paraíba (*João Pessoa, Mamanguape*); Pernambuco (*Camaragibe, Goiana, Paulista, Recife, São Lourenço da Mata*); Alagoas (*Campo Largo, Maceió*); Bahia (*Itanagra, Santo Antônio de Jesus*); Espírito Santo (*Serra – Jacaraípe*).



Text-Fig. 9. *Pyrrhopyge phidias aldo* **ssp. n.** Male genitalia: right valva, internal view (Brazil, Pernambuco, São Lourenço da Mata, Tiuma, 9-VIII-1958, H. Ebert leg., paratype, OM 7.644).

DESCRIPTION – Male: forewing length 20–22 mm. The subspecies shows two phenotypes: “*phidias* and *bixae* forms”, similar to *P. phidias phidias* and *P. phidias bixae*, respectively. However, the forewing length is typically less than all the other subspecies (24–25 mm), where smaller specimens are seldom observed. In the “*bixae* form” the bluish-white band at the base of the ventral side of the hindwing very narrow, not beyond the half of the discal cell. In the male genitalia, harpe dilated distally, rarely observed in the other subspecies and the dorso-basal expansion (“nose”, Evans 1951: 8) always very reduced which was not observed in the other subspecies.

Female: forewing length 20–25 mm, always smaller than in the two above mentioned subspecies (26–29 mm). Genitalia similar to the other *P. phidias* subspecies.

It represents the populations from the northeastern Brazil, penetrating to the northern portion of the Southeast region, from Maranhão to Espírito Santo. The populations from Maranhão (Imperatriz and Mirador) and from Paraíba (João Pessoa and Mamanguape) are of the “*bixae* form”; those from Pernambuco are of

both the “*phidias* form” (Garanhuns) and “*bixae* form” (Camaragibe, Goiana, Paulista, Recife and São Lourenço da Mata), as are those from Alagoas (“*bixae* form”) and Chão de Pilar, Maceió and Rio Largo (“*phidias* form”). Specimens from Bahia (Itanagra and Santo Antônio de Jesus) and Espírito Santo (Serra) are only of the “*phidias* form”.

ETYMOLOGY – The new subspecies is named after the collector of many of the paratypes, Aldo de Sá Cardoso who was a dermatologist in Maceió, Alagoas, Brazil and good friend of O. H. H. Mielke.

TYPES:

Holotype male with the following labels: / Holotypus/ XI-1982 São Lourenço da Mata – PE[rambuco, Brasil] Kesselring leg./ Holotypus *Pyrrhopyge phidias aldo* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 37.354/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ 18-VII-1978 Guajú, Mamanguape, PB [Paraíba, Brasil] 170 m, O.-C. Mielke leg./ *Pyrrhopyge phidias aldo* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 36.295/. OM-DZUP.

Paratypes: 1 ♀, 30-VII-2006–VIII-2011, Base da Geraldina, Parque Estadual Mirador, Mirador, Maranhão, Brazil, 06°37'25" S/ 45°52'08"W, F. Limeira-de-Oliveira, D. W. A. Marques & M. S. Oliveira leg. DZ 47.880 (DZUP); 1 ♀, 27-VIII-1974, Imperatriz, excursão Departamento de Zoologia – UFPR, DZ 33.419 (DZUP); 1 ♂, 14-VIII-1954, João Pessoa, Paraíba, Brasil, H. Ebert leg., ex coleção H. Ebert, DZ 33.458 (DZUP); 2 ♀, 3-X-1965, 1 ♀, 27-XI-1973, João Pessoa, Paraíba, Brazil, Kesselring leg., OM 7.645, OM 7.646, OM 12.478 (OM); 1 ♂ 1 ♀, 24-XI-1957, 1 ♂ 1 ♀, 28-I-1961, 1 ♀, 5-I-1962, Camaragibe, Pernambuco, Brazil, H. Ebert leg., ex coleção H. Ebert, DZ 33.498, DZ 33.508, DZ 33.518, DZ 33.528, DZ 33.329; 1 ♀, 8-III-1967, Garanhuns, Pernambuco, Brazil, 800-900 m, H. Ebert leg., ex coleção H. Ebert, DZ 33.339; 1 ♂, 2-XII-1954, Goiana, Pernambuco, Brazil, H. Ebert leg., OM 12.479 (OM); 1 ♂, 2-III-1968, Paulista, Pernambuco, Brazil, H. Ebert leg., ex coleção H. Ebert., DZ 2.220 (DZUP); 1 ♀, IV-1969, Paulista, Pernambuco, Brazil, A. F. Costa leg., OM 36.149 (OM); 1 ♂, IX-1969, 1 ♂, 13-III-1976, Paulista, Pernambuco, Brazil, Kesselring leg., OM 36.998, OM 37.126 (OM); 1 ♂, I-1971, 1 ♀, 4-VII-1972, 1 ♂, X-1975, 1 ♂, 28-VII-1979, 2 ♂, 9-X-1982, 1 ♀, XI-1982, 2 ♂, XI-1991, São Lourenço da Mata, Pernambuco, Brazil, OM 36.188, OM 36.098, OM 1.756, OM 37.050, OM 12.273, OM 37.119, OM 37.239, OM 37.107, 36.322 (OM); 1 ♂, 26-VII-1958, 1 ♀, 15-VII-1973, Tiuma, São Lourenço da Mata, Pernambuco, Brazil, Kesselring leg., OM 9.952, OM 36.134 (OM); 1 ♂, 9-VIII-1958 Tiuma, São Lourenço da Mata, Pernambuco, Brazil, H. Ebert leg., OM 7.644 (OM); 1 ♂, 26-VII-1985, Tiuma, São Lourenço da Mata, Pernambuco, Brazil, H. Ebert leg., ex coleção H. Ebert, DZ 33.468 (DZUP); 1 ♀, 10-XII-1961, Recife, Pernambuco, Brazil, H. Ebert leg., OM 10.194 (OM); 1 ♂, 25-XI-1961, Várzea, Recife, Pernambuco, Brazil, H. Ebert leg., ex coleção H. Ebert, DZ 33.478 (DZUP); 1 ♂, 28-I-2015, Mata do Brennard, Recife, Pernambuco, Brazil, P. Grossi leg., DZ 33.488 (DZUP); 1 ♀, 7-I-1945, 1 ♂, 21-X-1945, Chão de Pilar, Alagoas, Brazil, A. Cardoso leg., ex coleção A. Cardoso, DZ 33.349, DZ 33.359 (DZUP); 1 ♂, 22-VIII-1950, 1 ♀, V-1959, 1 ♀, 15-II-1973, 1 ♂, 6-VI-1977, 1 ♀, 2-II-1978, 1 ♀, 15-IX-1986, 1 ♀, 18-VII-1986, 1 ♀, 15-IX-1986, Maceió, Alagoas, Brazil, A. Cardoso leg., ex coleção A. Cardoso, DZ 33.369, DZ 33.379, DZ 33.489, DZ 33.459, DZ 33.399, DZ 33.429, DZ 33.389, DZ 33.469 (DZUP); 1 ♀, 14-III-1950, 1 ♂, 20-VII-1950, 2 ♂, 12-VIII-1961, 1 ♂ 1 ♀, VI-1965, 1 ♂, 1-XII-1973, 1 ♂, 7-IV-1974, Maceió, Alagoas, Brazil, A. Cardoso leg., OM 10.870, OM 37.066, OM 10.905, OM 10.906, OM 10.967, OM 10.968, OM 10.872, OM 37.152 (OM); 1 ♂, Maceió, Alagoas, Brazil, A. Cardoso leg., ex coleção A. Cardoso, DZ 33.439 (DZUP); 3 ♂ 3 ♀, Maceió, Alagoas, Brazil, A. Cardoso leg., OM 10.871, OM 10.900, 10.909, OM 10.873, OM 10.895, OM 10.907 (OM); 1 ♂ 2 ♀, 28-X-1973, 2 ♂, 9-XII-1973, 1 ♀, 2-XI-1976, Fazenda Riachão, Rio Largo, Alagoas, Brazil, A. Cardoso leg., OM 37.122, OM 37.041, OM 37.105, OM 37.116, OM 37.225, OM 37.062 (OM); 1 ♀, 22-VII-2012, Lontra, Itanagra, Bahia, Brazil, Linhares & Paluch leg., DZ

25.046 (DZUP); 1 ♂, 19-II-1987, Santo Antonio de Jesus, Bahia, Brazil, J. Becker leg., OM 21.914 (OM); 1 ♂, 23-XI-2013, Arraial d'Ajuda, Porto Seguro, Bahia, Brazil, 40 m, 16°27' S, 39° 03' W, V. O. Becker leg., OM 82.898 (OM); 1 ♂ 1 ♀, 11-18-II-1967, Jacaraípe, Serra, Espírito Santo, Brazil, C. Elias leg., DZ 33.449, DZ 22.414 (DZUP).

***Pyrrhopyge phidias leucoloma* Erschoff, 1875**

Pls. 18–19, Figs. 15 (neotype ♂), 16–17 (♂) (d, v)

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia.

Peru: Ayacucho (*Choimacota Valley, Monterrico, Río San Miguel*); Cusco (*Cosñipata Valley – Patria 675 m to Mirador 1,700 m, Illapani Viejo, Quillabamba*); Junín (*Chanchamayo, La Merced, Perené, Río Colorado, San Ramón, Satipo, Shimá*); Pasco (*Huancabamba, Pozuzo*). **Bolivia:** La Paz (*Yungas*).

TYPE:

Pyrrhopyge leucoloma Erschoff, 1875. The original description mentions an unspecified number of males from Monterico [*sic*], río San Miguel, Vale Choimacota, [Ayacucho], Peru, 25-X-??? in collection Mus. Varsovia. No syntypes were found in this museum, or in any museum or collection mentioned above (see “Abbreviations of the collections”). To ensure the correct identification of *Pyrrhopyge leucoloma*, mentioned as *Pyrrhopyge phidias leucoloma* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Peru, Cusco, Cosñipata Valley, Patria, 675 m, 25-IV-2015 O. Mielke leg./ gen[itália] pre[parada], [O.] Mielke 2015/ Neotypus *Pyrrhopyge leucoloma* Erschoff, 1875 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 77.323/. OM-DZUP.

***Pyrrhopyge aziza* Hewitson, 1866**

Three subspecies.

DISTRIBUTION – Northwestern, western and central-northern Colombia to central-eastern Bolivia.

***Pyrrhopyge aziza aziza* Hewitson, 1866**

Pls. 18–19, Figs. 21 (lectotype ♂) (d, v); Pls. 20–21, Figs. 1 (♀) (d, v)

DISTRIBUTION – Northwestern, western and central-northern Colombia.

Colombia: Antioquia (*Mesopotamia*); Santander (*Río Frío*); Valle del Cauca (*Calima Valley, Santa Rita Valley*).

The citation **Brazil:** Pará (*Río Tocantins*) (Weymer & Maassen 1890) is incorrect.

TYPE:

Pyrrhopyga [*sic*] *aziza* Hewitson, 1866. The original description mentions an unspecified number of specimens from New Granada [Colombia] in collection Hewitson. To ensure the correct identification of *Pyrrhopyga aziza*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ N. Granada [Colombia]. Hewitson Coll. 79-69. *Pyrrhopyga aziza*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga aziza* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge aziza araethyrea* Hewitson, 1870**

Pls. 18–19, Figs. 18 (lectotype ♂), 19–20 (♂) (d, v); Pl. 76, Figs. 1 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Peru.

Ecuador: Bolívar (*Balzapamba*); Loja (*San Francisco*); Morona-Santiago (*Huamboya*); Pastaza (*Mera, Río Pastaza*); Tungurahua (*Baños, El Topo, Hacienda La Mascota, Hacienda Santa Inés, Machay, Mapoto, Río Topo, Yunguilla*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas

(*Aramango – Numparque*); Pasco (*Oxapampa*); San Martín (*Rioja, Rioja – Naranjos 1,000 m to El Afluente 1,300 m*).

TAXONOMY – There is some variation in the colour of the upper hindwing. All our specimens from Ecuador (n= 2) and **Peru:** San Martín (*Rioja*) (n= 5) have a blue margin, while all our specimens from **Peru:** San Martín (without other indication) (n= 3) have this area almost white. The DNA barcode minimum divergence between *P. aziza araethyrea* (n= 2) and *P. attis attis* **stat. rest.** (n= 1) is 3.4%, between *P. aziza araethyrea* and *P. attis lexis* **comb. n.** (n= 5) is 3.0%, and between *P. aziza araethyrea* and *P. rafael* **sp. n.** (n= 4), a species with similar phenotype, is 2.4%.

TYPE:

Pyrrhopyga [*sic*] *araethyrea* Hewitson, 1870. The original description mentions an unspecified number of specimens from Mapoto, [Tungurahua], Ecuador, Buckley leg. To ensure the correct identification of *Pyrrhopyga araethyrea*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Ecuador Hewitson Coll. 79-69. *Pyrrhopyga araethyrea*. 1./ *araethyreal*; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga araethyrea* Hewitson, 1870, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge aziza troja* Evans, 1951**

Pls. 18–19, Figs. 22–23 (♂), 24 (holotype ♂) (d, v)

DISTRIBUTION – Western Colombia and central-eastern Peru to central-eastern Bolivia.

Colombia: Valle del Cauca. **Peru:** Cusco (*Callanga, Cosñipata Valley – Quebrada Quitacalzón 1,050 m to San Pedro 1,400 m*); Huánuco (*Puente Cayumba, Río Monzón, Tingo Maria*); Junín (*Chanchamayo, El Porvenir, Río Colorado, San Ramón*); Pasco (*Huancabamba, Oxapampa, Pozuzo*). **Bolivia:** La Paz.

TYPE:

Pyrrhopyge aziza troja Evans, 1951. Holotype male with the following labels: / Type – *troja* Evans/ San Ramon, [Junín], Peru, 3000 ft. X. '03 Watkins & Tomlinson/ Adams Bequest. B.M. 1912-399/. NHMUK.

Pyrrhopyge attis* Bell, 1931, *stat. rest.

Six subspecies.

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia, eastern Venezuela to French Guiana, northern, northeastern, southeastern and southern Brazil, Paraguay, northeastern Argentina and Uruguay.

TAXONOMY – The DNA barcode minimum divergence between *P. attis* **stat. rest.** (*P. attis attis* **stat. rest.**, *P. attis lexis* **comb. n.**) (n= 5) and *P. aziza* (*P. aziza araethyrea*) (n= 2) is 3.0%, consequently we believe these two taxa should be considered as distinct species. So, *Pyrrhopyge attis* **stat. rest.** has its status reinstated; the taxon was previously considered a subspecies of *Pyrrhopyge aziza*. It was impossible to detect differential characters in the male genitalia, which is quite variable in the appearance of the harpe of the valva. In contrast, the wing shape and colorations are quite characteristic for both species; while in *P. aziza* both fore and hindwings are more rounded and the only known phenotypes are of the “*hyperici* form” (*P. aziza aziza*) and “*sergius* form” (*P. aziza araethyrea* and *P. aziza troja*), while in *P. attis* **stat. rest.** the forewings are more pointed and the only known phenotypes are of the “*hyperici* form” (*P. attis cardus* **comb. n.**), “*bixae* form” (*P. attis lexis* **comb. n.**) and “*phidias* form” (*P. attis attis* **stat. rest.** and *P. attis arbor* **comb. n.**). The taxonomy of the subspecies is quite confusing, and certainly more specimens from areas not so far surveyed could contribute to a better understanding. DNA studies will certainly contribute to a more satisfactory resolution.

Pyrrhopyge attis lexis **comb. n.** and *P. aziza troja* are sympatric in Peru, Huánuco, Tingo Maria, corroborating the proposition that they are two species.

Pyrrhopyge attis attis* Bell, 1931, *stat. rest.

Pls. 20–21, Figs. 2–4 (♂) (d, v); Pl. 76, Figs. 2 (allotype ♀) (d, v)

Syn. *Pyrrhopyge denieri* Hayward, 1938

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia, northern, northeastern and southeastern Brazil.

Peru: Cusco (*Quincemil*); Madre de Dios (*Parque Nacional Bahuaja Sonene, Puerto Maldonado, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge (near Atalaya, Cusco)*). **Bolivia:** La Paz (*Caranavi, Río Zongo*); Santa Cruz. **Brazil:** Pará (*Juruti, Óbidos, Rio Arapiuns*), Mato Grosso (*Chapada dos Guimarães – Buriti*); Bahia (*Uruçuca*); Espírito Santo.

The citations **Colombia:** Cundinamarca (*Viotá*) (Bell 1931, Hayward 1947) are probably of *Pyrrhopyge attis arbor comb. n.*

REMARKS – Disjunct distribution in Brazil.

TYPE:

Pyrrhopyge attis Bell, 1931. Holotype male with the following labels: / Type/ Santa Cruz Bolivia/ G451/ *Pyrrhopyge attis* Bell Holotype ♂ / AMNH.

Pyrrhopyge denieri Hayward, 1938. Holotype male with the following labels: / Tipus/ Caranavi, [La Paz] Bolivia/ H. 87/ *Pyrrhopyge denieri* Hayw. Holótipo ♂ / Prep. N° 122/ IML.

Pyrrhopyge attis arbor* Evans, 1951, *comb. n.

Pls. 20–21, Figs. 5 (holotype ♂) (d, v); Pl. 76, Figs. 3 (♀) (d, v)

DISTRIBUTION – Central Colombia to eastern Venezuela.

Colombia: Arauca; Cundinamarca (*Viotá*); Tolima (*Cunday*). **Venezuela:** Amazonas (*San Juan de Manapiare*); Aragua (*Parque Nacional Henry Pittier – Rancho Grande*); Carabobo (*San Esteban*); Lara (*Sabana Alta*); Mérida (*Chorrera de las González, Estanques, Mérida*).

TYPE:

Pyrrhopyge aziza arbor Evans, 1951. Holotype male with the following labels: / Type – *arbor* Evans/ Venezuela/ Venezuela Dyson. 47-9/ I. NHMUK.

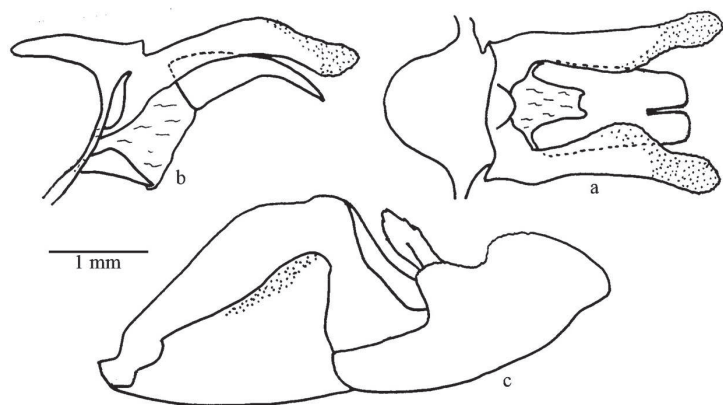
Pyrrhopyge attis cardus* Mabille, 1891, *comb. n.

Pls. 20–21, Figs. 6 (lectotype ♂) (d, v); Pl. 76, Figs. 4 (♀) (d, v); Text-Fig. 10 (♂ genitalia)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (*Itaituba, Óbidos, Santarém*).

TAXONOMY – Examination of the male genitalia of the lectotype confirms this *comb. n.* *Pyrrhopyga [sic] cardus* was considered a subspecies of *Pyrrhopyge proculus* Hopffer by Evans (1951).



Text-Fig. 10. *Pyrrhopyge attis cardus comb. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, uncus and gnathos, lateral left view; c) right valva, internal view (lectotype).

TYPE: *Pyrrhopyga [sic] cardus* Mabille, 1891. The original description mentions an unspecified number of males from Brazil [and Pará, Tapajos, Itaituba – on the label of the lectotype]. To ensure the correct identification of *Pyrrhopyga cardus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Itaituba, Tapajos, [Pará, Brazil] 86 Hhl./ *cardus* Mab./ *Pyrr.*

cardus Mab./ Coll. Staudinger/ gen. prep. [O.] Mielke 1979/ Genital-Unters. Nr. 4725, Zool. Mus. Berlin/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga cardus* Mabille, 1891. O. Mielke, Brockmann & C. Mielke det. 2019/ MfN.

Pyrrhopyge attis lexis* Evans, 1951, *comb. n.

Pls. 20–21, Figs. 7 (♂), 8 (♀) (d, v)

DISTRIBUTION – Central and eastern Colombia to southeastern Peru, Guyana, Suriname, French Guiana and northern Brazil.

Colombia: Cundinamarca (*Bogotá*); Meta (*Villavicencio*). **Ecuador:** Napo (*Jatun Sacha*); Pastaza (*Mera*); Zamora-Chinchipe (*Zamora*). **Peru:** Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Santa Isabel 1,200 m, Quincemil*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo*); Loreto (*Iquitos, Pebas, Upper Putumayo*); Madre de Dios (*Cerro Pantiacolla, Puerto Maldonado, Shintuya, Upper Río Madre de Dios – Amazonia Lodge (near Atalaya, Cusco)*); San Martín (*Juanjui, Sacanche, Satipo, Tocache*). **Guyana:** Barima-Waini (*Mabaruma*); Iwokrama Mountains. **Suriname:** Brokopondo (*Brownsberg*). **French Guiana:** Saint-Laurent-du-Maroni (*Saül*). **Brazil:** Pará (*Santarém*); Acre (*Bujari, Mâncio Lima – Parque Nacional da Serra do Divisor, Porto Acre, Senador Guimard, Marechal Thaumaturgo*); Rondônia (*Ariquemes, Rio Jamari*); Mato Grosso (*Brasnorte, Diamantino*).

TAXONOMY – The subspecies is associated to *P. attis stat. rest.* by the DNA barcode. The DNA barcode minimum divergence between *P. attis attis stat. rest.* (n= 1) and *P. attis lexis comb. n.* (n= 5) is 2.7%, suggesting they comprise two distinct species, but more studies are necessary, and between *P. attis lexis comb. n.* and *P. aziza araethyrea* (n= 2) is 3.0%, and between *P. attis attis stat. rest.* and *P. aziza araethyrea* is 3.4%.

TYPE:

Pyrrhopyge aziza lexis Evans, 1951. Holotype male with the following labels: / Type – *lexis* Evans/ Mabaruma [Barima-Waini] British Guiana Dec. 1929-Jan. 1930. A. Hall/ I. NHMUK.

Pyrrhopyge attis miersi* O. Mielke, Brockmann & C. Mielke, *ssp. n.

Pls. 20–21, Figs. 9 (holotype ♂), 10 (allotype ♀) (d, v)

DISTRIBUTION – Southeastern and southern Brazil (Rio de Janeiro and Santa Catarina).

Brazil: Rio de Janeiro (*Angra dos Reis*); Santa Catarina (*Joinville, São Francisco do Sul*).

DESCRIPTION – Male: forewing length 24–26 mm. Head, palpi and anal tuft red. Wings as the others subspecies, black with blue shades, white fringes on both wings, on the forewing reaching the apex; external margin of the forewing slightly concave between CuA₂–2A and of the hindwing between M₁–2A.

Female: forewing length 30–32 mm. As in the male, but external margins of the wings more rounded.

The subspecies is distinguished from the others “*phidias* form” subspecies, *P. attis attis stat. rest.*, *P. attis arbor comb. n.* and *P. attis subnubilus comb. n.*, by the longer forewing length and the white fringes present from the apex to tornus of the forewing, while in the others the forewing length, at least in the males, is shorter and the white fringes not reaching the apex. In addition, the geographic distribution is disjunct.

ETYMOLOGY – The subspecific name honours Herbert Miers, a good friend and collaborator in many field trips, in addition to being one of the best experts of Lepidoptera in Joinville and its surroundings, Santa Catarina, Brazil, where he collected throughout his life only one specimen of this new subspecies – the holotype.

REMARKS – Inhabitant of lowlands along the coast.

TYPES:

Holotype male with the following labels: / Holotypus/ 7-I-1968, São Francisco do Sul, S[anta]. Catarina [Brazil] Miers leg./ Holotypus *Pyrrhopyge attis miersi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke, det. 2017/ OM 36.167/ I. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Joinville

[sic] S[anta]. Catarina [Brazil]/; Allotypus *Pyrrhopyge attis miersi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 36.192/. OM-DZUP.

Paratypes: 1 ♂ 1 ♀, Joinville, Santa Catarina, Brazil, OM 36.102, OM 36.301 (OM); 1 ♂ without data, probably Joinville, Santa Catarina, Brazil, ex coleção Defesa Sanitária Vegetal, Rio de Janeiro, Brazil, where the Schmidt collection from Joinville is deposited, DZ 9.032 (DZUP); 1 ♂ 1 ♀, I-1933, Angra dos Reis, Estado do Rio de Janeiro, Brazil, L. Travassos Filho leg., OM 8.556, OM 8.557 (OM); 1 ♀, 23-II-1936, Jussara, Angra dos Reis, Estado do Rio de Janeiro, Brazil, d'Almeida leg., OM 66.054 (OM); 1 ♂, II-1937, Japuíba, Angra dos Reis, Estado do Rio de Janeiro, Brazil, L. Travassos Filho leg., OM 66.047 (OM).

***Pyrrhopyge attis subnubilus* Hayward, 1935, comb.**

n.

Pls. 20–21, Figs. 11 (♂), 12 (♀) (d, v)

DISTRIBUTION – Central-eastern Peru to central-southern Brazil, Paraguay, northeastern Argentina and Uruguay.

Peru: Pasco (*Pozuzo*). **Brazil:** Amazonas (*Eirunepé*); Goiás (*Ilha do Bananal*); Minas Gerais (*Conceição dos Ouros, Pouso Alegre*); São Paulo (*Campinas, Itapetininga, Ribeirão Preto, Rio Claro, Teodoro Sampaio – Parque Estadual do Morro do Diabo*); Paraná (*Foz do Iguaçu*); Rio Grande do Sul (*Barra do Quaraí*). **Paraguay:** Guairá (*Villarica*). **Argentina:** Misiones (*General Belgrano, Puerto Iguazú, Puerto Libertad (= Bemberg)*). **Uruguay:** Artigas (*San Gregorio*).

TAXONOMY – The subspecies is associated to *P. attis* **stat. rest.** by the male and female genitalia similarity and by the similar forewing shape.

TYPE:

Pyrrhopyge subnubilus Hayward, 1935. Holotype female with the following labels: / Typus/ Bemberg, Misiones, [Argentina] ... 1933 Hayward/ H. 86/ N° 4 K. J. Hayward det./ *Pyrrhopyge subnubilus* Holotipo ♀/ 2512/. MLP.

***Pyrrhopyge pusca* Evans, 1951**

Pls. 20–21, Figs. 13–14 (♂), 15 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia and northwestern Brazil (Acre).

Ecuador: Morona-Santiago (*Limón Indanza – General Leonidas Plaza Gutierrez*). **Peru:** Amazonas (*Cordillera del Condor – Falso Pakisha*); Cusco (*Cosñipata Valley – Quebrada Santa Isabel*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo, La Merced, Perené, Puerto Unión, Satipo, Quebrada Agua Meliza, Río Colorado, Shimá*); Madre de Dios (*Parque Nacional del Manu, Tambopata Reserve*); Pasco (*Puente Paucartambo*). **Bolivia:** Beni (*Río Beni*); La Paz (*Parque Nacional Madidi 1,500–1,800 m, Río Mapiri*). **Brazil:** Acre (*Bujari*).

TYPE:

Pyrrhopyge pusca Evans, 1951. Holotype male with the following labels: / Type –pusca Evans/ Pérou [Junín] Chanchamayo O. Schuncke 1892/ P 20 R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Pyrrhopyge tingo* O. Mielke, Brockmann & C.**

Mielke, sp. n.

Pls. 20–21, Figs. 16 (holotype ♂) (d, v); Text-Fig. 11 (♂ genitalia)

DISTRIBUTION – Central-eastern Peru.

Peru: Huánuco (*Tingo Maria*).

DESCRIPTION – Male: “*bixae* form”. Forewing length 25 mm. Ground colour black with some bluish iridescence. Head, palpi, collar mesally and anal tuft red. Fringes white, on the forewing from R_5 to tornus. Forewing with outer margin not excavated in CuA_2 –2A. Hindwing ventrally with the basal area light bluish-white from costal margin until inner margin not reaching the end of the cell, interrupted

by the dark veins (larger at 2A) and outwardly straight from costal margin to nearly 2A. Genitalia very characteristic; apophyses of tegumen nearly rectangular, short and spiny; uncus bifid with a very short slit; valva short, with the harpe ending in a dark and spiny fold overlapping nearly all the costa, and sacculus with a dorsal expansion, also spiny. Only one male is known.

Female: unknown.

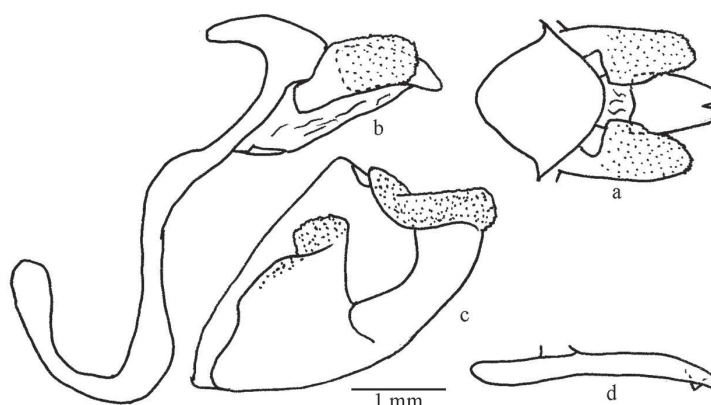
TAXONOMY – A species of the “*bixae* form” only possible to identify by an examination of the very typical male genitalia.

ETYMOLOGY – The specific name refers to Tingo Maria, the type locality of the species.

REMARKS – Only known from the type locality.

TYPE:

Holotype male with the following labels: / Holotypus/ Tingo Maria, Huánuco, Peru, 670 m 18-II-1993 Büche leg./ gen[italia] prep[arada].. [O.] Mielke 2014/ Holotypus *Pyrrhopyge tingo* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 36.042/. OM-DZUP.



Text-Fig. 11. *Pyrrhopyge tingo* **sp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

***Pyrrhopyge infantilis* H. H. Druce, 1908**

Two subspecies.

DISTRIBUTION – Northern Peru to central-eastern Bolivia, southern Venezuela to French Guiana and northern Brazil.

***Pyrrhopyge infantilis infantilis* H. H. Druce, 1908**

Pls. 20–21, Figs. 17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Northern Peru to central-eastern Bolivia, southern Venezuela to French Guiana and northern Brazil.

Peru: Amazonas (*Abra Wawajin 4 km W*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to Quebrada Santa Isabel 1,200 m, Quincemil*); Junín (*Chanchamayo, Río Colorado, Satipo*); Loreto (*Caballococha, Iquitos, Pebas, Río Sucuari – Explornapo-Aceer, Upper Río Putumayo*); Puno (*Sandia*); San Martín (*Juanjui*); Ucayali (*Contamana*). **Bolivia:** Beni (*Reyes*); La Paz (*Parque Nacional Madidi 1,500–1,800 m, Yungas*); Santa Cruz (*Buenavista*). **Venezuela:** Amazonas (*Yavita*). **French Guiana:** Cayenne (*Montagne des Chevaux – Galion*); Saint-Laurent-du-Maroni (*Saül*). **Brazil:** Amazonas (*Benjamin Constant, Manicoré, Caiari – Rio Uaupés, São Paulo de Olivença*); Rondônia (*Ariquemes, Candeias do Jamari – Fazenda Urupá*); Mato Grosso (*Cáceres, Brasnorte*).

TYPE:

Pyrrhopyge infantilis H. H. Druce, 1908. The original description mentions an unspecified number of males from Peru in collection Druce. To ensure the correct identification of *Pyrrhopyge infantilis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Peru/ *Pyrrhopyge infantilis* H. H. Druce Type T.R.S. 1908./ ex coll. Hamilton Druce, 1919/ J.J.Joicey Coll. B.M. 1925-451./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge infantilis* H. H. Druce, 1908, O. Mielke,

***Pyrrhopyge infantilis agala* Evans, 1951**

Pls. 20–21, Figs. 19 (♂), 20 (♀) (d, v)

DISTRIBUTION – Central-eastern Bolivia and northern Brazil (Pará).

Bolivia: Cochabamba (*San Jacintho*); La Paz (*Caranavi, Río Zongo*). **Brazil:** Pará (*Juruti*).

TYPE:

Pyrrhopyge infantilis agala Evans, 1951. Holotype male with the following labels: / Type – *agala*/ San Jacinto, [Cochabamba] Bolivia 6-8000ft. Garlepp/ Godman-Salvin Coll. 1912-23./. NHMUK.

***Pyrrhopyge rafael* O. Mielke, Brockmann & C. Mielke, sp. n.**

Pls. 20–21, Figs. 21 (holotype ♂), 22 (allotype ♀) (d, v)

DISTRIBUTION – Central-eastern Peru.

Peru: Huánuco (*Río Monzón, Tingo Maria*).

DESCRIPTION – Species of the “*sergius* form”. Male: forewing length 22–25 mm; female 26 mm. Head, palpi, collar and anal tuft red. Ground colour dark greyish-black, not bluish as in all *P. aziza* subspecies. Forewing fringe white from M_2 – M_3 to tornus; ventrally sometimes with white marginal suffusion between M_2 and M_3 . Hindwing fringe white between R_5 and anal angle; dorsal submarginal area, between apex and anal angle, white, cut by the dark veins (absent in M_2) and larger between M_1 and 2A, reaching 1/3 of the distance between the margin and the discal cell; ventrally outer half between the apex and the anal angle and until the discal cell or even entering a short distance in the cell, light blue and white near the margin, cut by the dark veins (absent in M_2).

This taxon differs from *Pyrrhopyge aziza araethyrea*, which it resembles somewhat due to the colour pattern of the hindwings, by the more pointed wings, the outer margin of the hindwing always being white (sometimes bluish in *P. a. araethyrea*) and narrower, and by a DNA barcode minimum divergence of 2.4% (*P. rafael* sp. n. n= 4, *P. a. araethyrea* n= 2). *Pyrrhopyge rafael* sp. n. also differs by its greyish black general wing colouration without the bluish reflection as in other taxa and by the DNA barcode. The DNA barcode minimum divergence between *P. attis attis* stat. rest. (n= 1), *P. attis lexos* comb. n. (n= 5), a close related species, and *P. rafael* sp. n. (n= 4) is 1.7%. No male genitalia characters that may separate the three taxa were found.

ETYMOLOGY – The specific name is in honour of Rafael Mielke Schwartzburd, grandson of Olaf H. H. Mielke and nephew of Carlos G. C. Mielke.

TYPES:

Holotype male with the following labels: / Holotypus/ Tingo Maria, Huánuco, Peru, 670 m, 15-III-1992, Büche leg./ Holotypus *Pyrrhopyge rafael* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 35.894/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Tingo Maria, Huánuco, Peru, 670 m, 30-VII-1991, Büche leg./ Allotypus *Pyrrhopyge rafael* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 37.153/. OM-DZUP.

Paratypes: 1 ♂, V-1964, 1 ♂, XI-1964, Tingo Maria, Huánuco, Peru, OM 7.100, OM 7.101 (OM); 1 ♂, 6-III-1966, Río Monzón, Huánuco, Peru, OM 36.143 (OM); 1 ♂, 24-IV-1991, 1 ♂, 6-IX-1991, 1 ♂, 26-X-1991, 2 ♂, 5-I-1992, 18-IV-1992, Tingo Maria, Huánuco, Peru, 670 m, Büche leg., OM 35.593, OM 35.664, OM 35.396, OM 35.901, OM 35.949, OM 35.131 (OM); 1 ♂, 22-VII-1981, Las Palmas, Tingo Maria, Huánuco, Peru, 650 m, [O.] Mielke & Casagrande leg., OM 25.838 (OM); 2 ♂, Tingo Maria, Huánuco, Peru, OM 76.533, OM 76.543 (OM); 1 ♂, 27-X-1971, 1 ♂, XI-1971, Rondos, Tingo Maria, Huánuco, Peru, 850 m, Villegas leg., DZ 29.445, DZ 29.446 (DZUP); 11 ♂ 5 ♀, 19-III-1985, 7-XII-1985, 16-XI-1986, 4-I-1987, 17-I-1987, 7-IV-1987, 3-IX-1987, 22-IV-

1989, 16-VIII-90, 10-X-1990, III-1997, IV-2003, IX-X-2003, Tingo Maria, Huánuco, Peru, ex coll. M. Büche, HESP-EB 01 107, HESP-EB 01 108, HESP-EB 01 110, HESP-EB 01 113, HESP-EB 01 114, HESP-EB 01 156, HESP-EB 01 157, HESP-EB 01 158, HESP-EB 01 251, HESP-EB 01 252, HESP-EB 01 253, HESP-EB 01 254, HESP-EB 01 255 (EB); 1 ♂, 2-X-2014 (Genitalia-160098), 1 ♂, 2-X-2015, Tingo Maria, Huánuco, Peru, J. Uehara leg.; 2 ♀, IX-2015, Tingo Maria, Huánuco, Peru (CJU).

***Pyrrhopyge iquitos* O. Mielke, Brockmann & C. Mielke, sp. n.**

Pls. 20–21, Figs. 23 (holotype ♂) (d, v); Text-Fig. 12 (♂ genitalia)

DISTRIBUTION – Eastern Ecuador to south-eastern Peru.

Ecuador: Napo (*Llaucana*). **Peru:** Cusco (*Quincemil*); Loreto (*Iquitos*).

DESCRIPTION – Male: “*bixae* form”. Forewing length 24 mm. Ground colour black with some bluish iridescence. Head, palpi and anal tuft red. Fringes white, on forewing from M_3 to tornus. Forewing with outer margin excavated in CuA_2 –2A. Hindwing ventrally with the basal area light bluish-white from costal margin to the inner margin, just reaching the end of the cell, interrupted by the dark veins (larger at 2A) and outwardly curved. Genitalia very characteristic; apophyses of tegumen long, angled, spined dorsally and laterally and reaching the uncus tip; uncus bifid with a very short and narrow slit; valva with a dorso-distally expansion, strongly constricted at the base. Only one male is known.

Female: unknown.

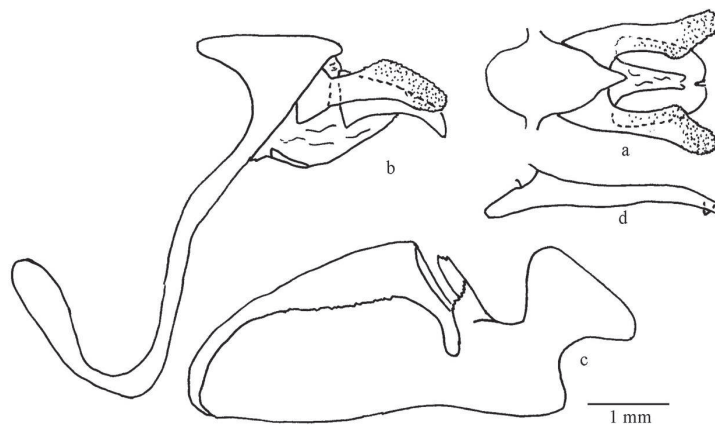
TAXONOMY – A species of the “*bixae* form”, only possible to identify by an examination of the genitalia. The bifid uncus with a very small slit and the typical valva are diagnostic for the species. This uncus aspect is also shared with *P. phidias*, *P. aziza*, *P. attis* stat. rest., *P. infantilis*, *P. pusca*, *P. draudti* stat. rest., and *P. proculus*, suggesting a close relationship.

ETYMOLOGY – The specific name refers to Iquitos, the type locality of the species.

TYPE:

Holotype male with the following labels: / Holotypus/ Iquitos, [Loreto], Peru, 100 m I-1993 Büche leg./ gen[italia]. prep[arada]., [O.] Mielke 2014/ Holotypus *Pyrrhopyge iquitos* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 35.987/. OM-DZUP.

Paratypes: 3 ♂, 6-8-X-2015 (Genitalia-160120, 160122, 160125), 2 ♂, 13-14-IV-2017 (Genitalia-180015, 180057), Quincemil 700 m, Cusco, Peru, J. Uehara leg. (CJU); 1 ♂, 14-IV-2017, Quincemil 700 m, Cusco, Peru, M. Solis leg., Genitalia-180018 (CJU); 1 ♂, 20-XII-2016, Llaucana, Napo, Ecuador, Genitalia-180076 (CJU).



Text-Fig. 12. *Pyrrhopyge iquitos* sp. n. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

***Pyrrhopyge sabrina* O. Mielke, Brockmann & C.**

Mielke, sp. n.

Pls. 20–21, Figs. 24 (holotype ♂) (d, v); Text-Fig. 13 (♂ genitalia)

DISTRIBUTION – Central-eastern Peru.

Peru: Junín (Andamarca – Villa El Salvador).

DESCRIPTION – Male: “*phidias* form”. Forewing length 22 mm, not produced, more rounded, and not excavated at outer margin. Ground colour black with some very dark bluish iridescence. Head black with some orange scales on the frons; palpi, genae, collar, lateral parts of patagia and base of tegulae similar. Anal tuft orange red. Fringes white, on forewing from M_3 – CuA_1 to tornus, and on the hindwing between Rs and 2A, ventrally a little larger and crenulated. Genitalia very characteristic: apophyses of tegumen broad, angled, spined dorsally and laterally, and shorter than uncus; uncus bifid with a very short and narrow slit, as in *P. phidias*, dorso-distally with a small projection on the left part of the uncus, on the right part only a small callosity; valva with a short dorso-distally expansion, not expanded distally, but with strong spines at the distal point. Only one male is known.

Female: unknown.

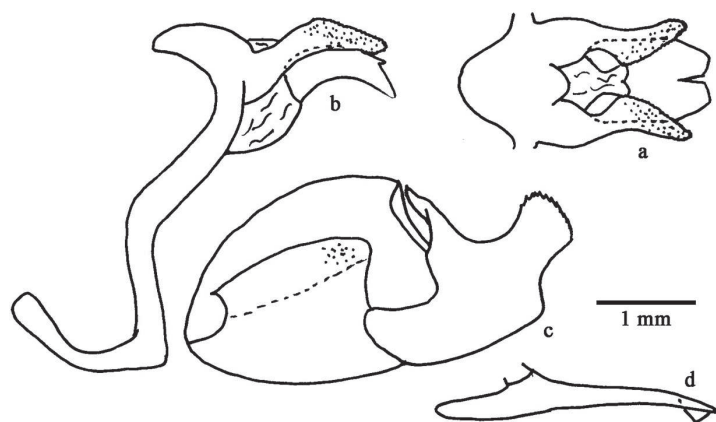
TAXONOMY – A species of the “*phidias* form” apparently very different from any other species of the “*phidias* group”. It is the only known species with very few orange, instead of red, scales on the head. The bifid uncus with a very small slit and the typical valva are diagnostic for the species. This uncus aspect is also shared with *P. phidias*, *P. aziza*, *P. attis* **stat. rest.**, *P. infantilis*, *P. pusca*, *P. draudti* **stat. rest.**, and *P. proculus*, suggesting a close relationship.

ETYMOLOGY – The specific name is in honour of Sabrina Brockmann, the daughter of Ernst Brockmann.

REMARKS – Only known from the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ 5-X-2007, El Salvador, Andamarca, Junín, Peru, 1600 m S 11°54'38", W 74°44'59"/ gen[italia]. prep[arada]., Mielke 2014/ Holotypus *Pyrrhopyge sabrina* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 68.847/. OM-DZUP.



Text-Fig. 13. *Pyrrhopyge sabrina* sp. n. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

Pyrrhopyge draudti* Bell, 1931, **stat. rest.*

Four subspecies.

DISTRIBUTION – Colombia to central-eastern Bolivia, Trinidad and Tobago, Venezuela to French Guiana and northern Brazil.

Pyrrhopyge draudti draudti* Bell, 1931, **stat. rest.*

Pls. 22–23, Figs. 5, 7 (♂), 6 (♀) (d, v)

DISTRIBUTION – Colombia to central-eastern Bolivia and northwestern Venezuela to French Guiana.

Colombia. Ecuador: Morona-Santiago (*Gualaquiza, Pablo Sexto*,

San Isidro, Palora – *San Vicente de Tarqui, Santiago de Méndez*); Napo (*Yanahurco, Tena*); Pastaza (*Puyo*); Tungurahua (*Baños*). **Peru:** Amazonas (*Santa Maria de Nieva*); Cusco (*Chontachaca 950 m, Quincemil*); Huánuco (*Puente Cayumba, Tingo Maria*); Junín (*Río Colorado, Río Perené, Satipo*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado, Tambopata Reserve*); Puno (*Sandia*); San Martín (*Juanjui, Tarapoto*). **Bolivia:** Beni; La Paz (*Caranavi, Coroico, Río Zongo*); Santa Cruz (*Buenavista*). **Venezuela:** Barinas (*Barinas*); Carabobo (*Puerto Cabello, Palmichal*); Mérida (*Zea*); Táchira (*La Morita, Los Naranjos, Río Frio, San Felix, San Joaquín de Navay*); Trujillo (*La Gira*). **French Guiana:** Saint-Laurent-du-Maroni (*Saint-Laurent-du-Maroni*).

TAXONOMY – Evans (1951) considered all the taxa here included in *Pyrrhopyge draudti* **stat. rest.** as subspecies of *Pyrrhopyge proculus* Hopffer, 1874, but this species is the senior synonym of *Pyrrhopyge thericles grinda*, consequently *Pyrrhopyge draudti* **stat. rest.**, as the oldest available name, is the valid name for this taxon.

TYPE:

Pyrrhopyge draudti Bell, 1931. Holotype male with the following labels: / Type/ Santa Cruz Bolivia/ *Pyrrhopyge draudti* Bell Holotype ♂/. AMNH.

***Pyrrhopyge draudti bruno* O. Mielke, Brockmann & C. Mielke, sp. n.**

Pls. 22–23, Figs. 1 (holotype ♂), 2 (allotype ♀) (d, v); Text-Fig. 14 (♂ genitalia)

DISTRIBUTION – Eastern Colombia, Trinidad and Tobago, northeastern Venezuela to Guyana and northern Brazil (Pará).

Colombia: Meta (*Remolino, Villavicencio*). **Venezuela:** Monagas (*Caripe, Caripito*); Sucre (*Finca Vuelta Larga* – 8 km S *Guaraunos*). **Trinidad and Tobago. Guyana. Brazil:** Pará.

TAXONOMY – This new subspecies refers to *Pyrrhopyge proculus* **sensu** Evans (1951). It represents the only “*phidias* form” of the species.

Male: forewing length 20–22 mm. Typical appearance of a *Pyrrhopyge* species of the “*Pyrrhopyge phidias* species group” (Evans 1951). Head, palpi and anal tuft red. Wings dorsally and ventrally black with white fringes, on the forewing between M_2 and tornus and slightly larger on the hindwings. Fore and hindwings excavated. In one of the specimens, the ventral face of the hindwing with a slight indication of a bluish-white band, also mentioned in a few specimens in the NHMUK by Evans (1951) for *Pyrrhopyge proculus proculus*, the species already suggested by the same author as possibly being *Pyrrhopyge thericles guianae* Bell, 1932, name later corrected to *Pyrrhopyge thericles grinda* Evans, 1953. Genitalia similar to that of *Pyrrhopyge draudti draudti* **stat. rest.**

Female: similar to the male. Forewing length 23 mm. Wings less excavated and ventral forewing with a few bluish-white scales at the base between C and R.

There are four females from **Guyana** and one from **Brazil**: Pará in NHMUK (Evans 1951).

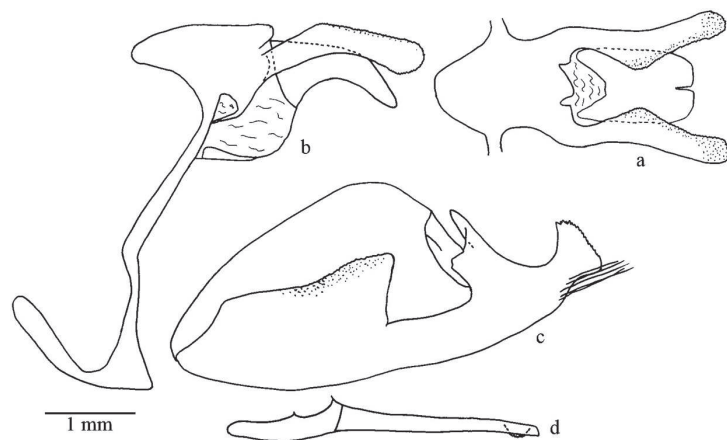
ETYMOLOGY – The specific name is in honour of Bruno Mielke Schwartzburd, grandson of Olaf H. H. Mielke and nephew of Carlos G. C. Mielke.

TYPES:

Holotype male with the following labels: / Holotypus/ 22-III-1982 30 km E. de Villavicencio, Meta, Colombia, Callaghan leg./ gen. prep. [O.] Mielke 2014/ Holotypus *Pyrrhopyge draudti bruno* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ BC-OM NG [barcoded by Nick Grishin]/ OM 37.102/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Colombia, Meta, Villavicencio, La Union, 4° 03' 43".3 N 73° 42' 04.6 W, 24-XI-2012, 500 m, G. Nielsen leg./ Allotypus *Pyrrhopyge draudti bruno* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ BC-OM [barcoded]/ OM 77.877/. OM-DZUP.

Paratypes: 1 ♂, 15-X-1992, Remolino, Meta, Colombia, 600 m, Le Crom leg., OM 35.711 (OM); 1 ♂, no data, OM 12.028 (OM). The last specimen with the light blue band on the ventral hindwing very reduced.



Text-Fig. 14. *Pyrrhopyge draudti bruno* **ssp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, left view (holotype).

Pyrrhopyge draudti lina* Bell, 1947, *comb. rest.

Pls. 22–23, Figs. 3 (♂), 4 (♀) (d, v)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (Belém, Capitão Poço, Jacareacanga, Ourém, Santarém – Fazenda Taperinha, Santo Antonio do Tauá).

TAXONOMY – Here the original combination with *Pyrrhopyge draudti* **stat. rest.** is reinstated.

TYPE:

Pyrrhopyge draudti lina Bell, 1947. Holotype male with the following labels: / Holotype/ Para, Brazil/ A. M. Moss/ G1812/ *Pyrrhopyge draudti* s. sp *lina* Bell Holotype ♂/. AMNH.

Pyrrhopyge draudti cintra* Evans, 1951, *comb. n.

Pls. 22–23, Figs. 8, 10 (♀), 9, 11–12 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to southeastern Peru, southern Venezuela to French Guiana and northern Brazil.

Ecuador: Napo (*Sacha*); Pastaza (*Puyo*). **Peru:** Loreto (*Florida, Iquitos, Pebas, Upper Putumayo*); Madre de Dios (*Tambopata Reserve*). **Venezuela:** Amazonas (*Yavita*). **Guyana.** **Suriname:** Marowijne (*Río Patamakka*); Paramaribo (*Zanderij*); Sipaliwini (*Lely Gebergte, Suriname*); Wanica (*Domburg*). **French Guiana:** Cayenne (*Camp Patawa, Tonate*); Saint-Laurent-du-Maroni (*Maroni*). **Brazil:** Amazonas (*Barcelos – Comunidade Acuquaia-Rio Padauari, Fonte Boa, Itacoatiara, Manaus, Manicoré, Marã, Maués, Parintins, São Gabriel da Cachoeira, São Paulo de Olivença, Tefé*); Pará (*Altamira, Itaituba, Juruti, Óbidos, Rio Arapiuns, Santarém*); Acre (*Porto Acre, Senador Guimard, Xapuri*); Rondônia (*Cacaulândia, Candeias do Jamari*); Mato Grosso (*Diamantino*).

TAXONOMY – See comments under *P. draudti draudti* **stat. rest.** for the new combination. In a few specimens from Maués, Amazonas, Brazil there is some blue suffusion on the dorsal hindwing, similar as in *Pyrrhopyge attis cardus* **comb. n.** – the “*hyperici* form”.

TYPE:

Pyrrhopyge draudti cintra Evans, 1951. Holotype male with the following labels: / Type – *cintra* Evans/ Villa Nova [=Parintins], Amazons [Brazil]. Bates/ ♂/ Godman-Salvin Coll. 1912–23. *Pyrrhopyge fluminis*, Butl./ NHMUK.

***Pyrrhopyge proculus* Hopffer, 1874**

Two subspecies.

DISTRIBUTION – Southern Venezuela to French Guiana and northern Brazil.

***Pyrrhopyge proculus proculus* Hopffer, 1874**

Pls. 22–23, Figs. 13 (lectotype ♂ of *Pyrrhopyge proculus*), 14 (♀) (d, v); Text-Fig. 15 (♂ genitalia)

Syn.: *Pyrrhopyge thericles grinda* Evans, 1953, *syn. n.*

DISTRIBUTION – Southern Venezuela to French Guiana.

Venezuela: Bolívar (*road Dorado – Santa Elena, km 125*). **Guyana:** Mazaruni-Potaro (*Mazaruni*); Demerara-Mahaica (*Demerara River*).

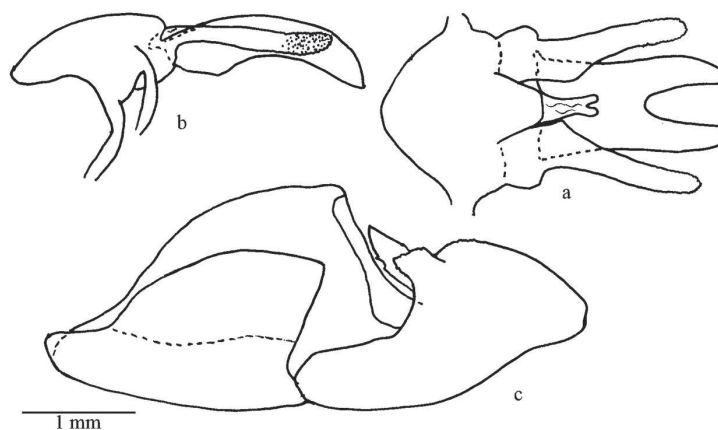
TAXONOMY – The male genitalia of the lectotype (Fig. 15) confirm that *P. proculus* is the senior synonym of *Pyrrhopyge thericles grinda* Evans, 1953 as suggested by Evans (1951), but here mentioned as *P. thericles guianae*, a correction published by Evans (1953).

The male genitalia of *Pyrrhopyge proculus ponicia* **comb. n.** are similar to that of *P. proculus proculus*, the reason to consider them subspecies of the same species. The remaining *P. thericles* subspecies considered by Evans (1951), *P. thericles thericles*, *P. thericles orientis*, *P. thericles pseudophidias*, *P. thericles ronda*, *P. thericles fola*, and *P. thericles rileyi* differ from the male genitalia of *P. proculus* on the length of the apophyses of the tegumen, not reaching the distal portion of the uncus in the latter and going much beyond the uncus in the former. Thus, both taxa are here considered as distinct species. In addition, *P. proculus ponicia* **comb. n.** is sympatric to *P. thericles thericles* in Brazil (Pará: Itaituba and Óbidos), which reinforces the separation of these two species.

TYPES:

Pyrrhopyge proculus Hopffer, 1874. The original description mentions an unspecified number of specimens of males from Guiana. To ensure the correct identification of *Pyrrhopyge proculus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Lectotypus/ Guiana Angl[aise]. Schomb./ 4883/ *Proculus* Hpf*. *zeleucus* Et./ 1: 18/ gen. prep. [O.] Mielke 1979/ Genital.-Unters. Nr. 4721, Zool. Mus. Berlin/; another label will be added: / Lectotypus *Pyrrhopyge proculus* Hopffer, 1874. O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published.

Pyrrhopyge thericles grinda Evans, 1953. Holotype male with the following labels: / Type – *grinda* Evans/ B[ritish]. Guiana. 1916./ J.J.Joicey Coll. B.M. 1925-451/. NHMUK. **Syn. n.**



Text-Fig. 15. *Pyrrhopyge proculus proculus*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen and uncus, lateral left view; c) right valva, internal view (lectotype).

Pyrrhopyge proculus ponicia* Evans, 1951, *comb. n.

Pls. 22–23, Figs. 15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Suriname, French Guiana and northern Brazil (Amazonas and Pará).

Suriname: Brokopondo (*Rama*); Para (*Zanderij*); Sipaliwini (*Suriname*). **French Guiana:** Cayenne (*Montagne de Kaw – Camp Caiman, Port Inini, Wacapou*); Saint-Laurent-du-Maroni (*Montagne Sable, Nouveaux Chantier*). **Brazil:** Amazonas (*Manicoré*); Pará (*Itaituba, Óbidos*).

TAXONOMY – See comments under *P. proculus proculus* for the new combination.

TYPE:

Pyrrhopyge thericles ponicia Evans, 1951. Holotype male with the following labels: / Type – *ponicia* Evans/ Cayenne [French Guiana]/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Pyrrhopyge thericles Mabilles, 1891

Seven subspecies.

DISTRIBUTION – Costa Rica to western Ecuador, central and eastern Colombia to southeastern Peru, Trinidad and Tobago, northwestern, northern and southern Venezuela to French Guiana and northern, central-northern, northeastern to southeastern Brazil.

REMARKS – Red fore coxae, so far only observed in subspecies *P. thericles orientis* (Pl. 19, Fig. 8).

Pyrrhopyge thericles thericles Mabilles, 1891

Pls. 22–23, Figs. 23 (♂), 24 (♀) (d, v); Pls. 24–25, Figs. 1, 3 (♂), 2 ♀ (d, v)

DISTRIBUTION – Northern Brazil.

Brazil: Amazonas (*Boa Vista do Ramos – Massauari, Careiro, Manaus, Maués, São Gabriel da Cachoeira, São Paulo de Olivença, Tefê*); Pará (*Itaituba, Juruti, Óbidos, Rio Arapiuns, Santarém*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor, Senador Guiomard, Xapuri*); Rondônia (*Alto Paraíso, Ariquemes, Cacaúlândia, Candeias do Jamari, Ouro Preto, Porto Velho, Presidente Médici*); Mato Grosso.

TAXONOMY – In a few specimens from Maués there is some blue suffusion on the dorsal hindwing, similar to *Pyrrhopyge attis cardus comb. n.*

TYPE:

Pyrrhopyga [*sic*] *thericles* Mabilles, 1891. The original description mentions an unspecified number of males from Masauary [Massauari, Boa Vista do Ramos, Amazonas, Brazil]. To ensure the correct identification of *Pyrrhopyga thericles*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Massauary Hhl./ 229. *Pyrr. thericles* Mab./ *Thericles* Mab./ Coll. Staudinger/ Genital-Unters. 4724. Zool. Mus. Berlin/ Gen. prep. [O.] Mielke 1979/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga thericles* Mabilles, 1891, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyge thericles pseudophidias Bell, 1931

Pls. 22–23, Figs. 17, 19, 21 (♂), 18, 20, 22 (♀) (d, v)

DISTRIBUTION – Costa Rica to western Ecuador, central and eastern Colombia to southeastern Peru, northwestern Venezuela to French Guiana and northern Brazil (Amazonas and Maranhão).

Costa Rica: Puntarenas (*Guaycará – La Gamba*); San José (*Cantón de Mora – El Rodeo*); San Ramón (*Bajo Rodríguez*). **Panamá:** Chiriquí; Colón (*Barro Colorado Island, Gamboa, Gatun*); Panamá (*Arraiján, Cocoli, Corozal – Cruces Trail, Farfan, Kobbe, Madden Forest, Pedro Miguel – Chiva Chiva Trail, Summit*); Panamá Oeste (*Cerro Campana*). **Colombia:** Amazonas (*La Pedrera*); Boyacá (*Muzo, Otanche, Turebia*); Caldas (*Victoria*); Cundinamarca (*Bogotá*); Meta (*Villavicencio*); Putumayo (*Río Mulato*). **Ecuador:** Esmeraldas (*Chuchuvi, Durango, Lita*); Guayas (*Guayaquil*); Imbabura (*Paramba*); Loja (*Loja*); Napo (*Tena*); Morona-Santiago (*Pablo Sexto, San Isidro, Santiago de Méndez, Sucúa*); Orellana (*Loreto*); Pastaza (*Puyo, Río Pastaza*); Pichincha (*Nanegalito, Pacto*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Rodríguez de Mendoza, Santa María de Nieva*); Cajamarca (*Charapi*); Cusco (*Cosñipata Valley – Chontachaca 900 m to San Pedro 1,400 m, Quincemil*); Huánuco (*Tingo Maria, Yuyapichis*); Junín (*Chanchamayo, La Merced, Perené, Satipo, Shimá*); Loreto (*Contamana, Iquitos, Nauta, Pebas, Upper Putumayo*); Pasco (*Pichis Road, Pozuzo*); Puno (*Sandia*); San Martín (*Juanjuí, Moyobamba, Rioja – Naranjos 1,000 m to El Afluente 1,300 m, Sacanche, Shapaja, Tarapoto, Tocache*); Ucayali (*Pucallpa*). **Venezuela:** Barinas (*Barinitas, La Soledad*); Carabobo (*Puerto Cabello*); Lara (*Guayamure, La Mejía, Parque Nacional Yacambú, Terepaima*); Táchira (*Angarabeca from Lobatera, between Capacho and La Cristalina, Chorro del Índio, El Junco, Parque Nacional El Tamá – between Río Frío and El Morretón, La Grita, La Morita, Michelena, Palmira – Quebrada La Flautera, Palo Gordo, Potreru, Río Frío, Río Negro, Rubio, Saladito, San Cristóbal – Paramillo, San Cristóbal – Río Chucurí, San Félix – Quebrada La*

Uruca, San Joaquín de Navay, Upper Río Doradas – Las Cuevas); Trujillo (*Campo Elías, La Gira de Betijoque, Mosquey, Rafael Rangel – La Gira*); Zulia (*El Tucuco, Perijá – San Marino*). **Suriname:** Para (*Zanderij Island*); Wanica (*Lelydorp*). **French Guiana:** Saint-Laurent-du-Maroni (*Saint-Laurent-du-Maroni*). **Brazil:** Amazonas (*Tonantins*); Maranhão (*Penalva*).

TYPE:

Pyrrhopyge pseudophidias Bell, 1931. Holotype male with the following labels: / Type/ Muzo, [Boyacá] Colombia/ R. C. Williams, Jr. Collection/ G433/ *Pyrrhopyge pseudophidias* Bell Holotype ♂/. AMNH.

Pyrrhopyge thericles ronda Evans, 1953

Pls. 24–25, Figs. 4 (♀) (d, v); Pl. 76, Figs. 5 (holotype ♂) (d, v)

DISTRIBUTION – Trinidad and Tobago.

Trinidad and Tobago: Port of Spain; Saint George.

TYPE:

Pyrrhopyge thericles ronda Evans, 1953. Holotype male with the following labels: / Type – *ronda* Evans/ Pt. [Port] of Spain [Trinidad and Tobago] I. 97. (Dr. Rendall)/ Rothschild Bequest B.M. 1939-1/. NHMUK.

Pyrrhopyge thericles fola Evans, 1951

Pls. 24–25, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Central and eastern Colombia to southern Venezuela.

Colombia: Cundinamarca (*Bogotá, Medina*); Meta (*Upper Río Negro, Villavicencio*). **Venezuela:** Amazonas (*Comunidad Mavaco – Río Autana*); Bolívar (*Las Claritas, Sierra de Lema, Uruyén*).

TYPE:

Pyrrhopyge thericles fola Evans, 1951. Holotype male with the following labels: / Type – *fola* Evans/ Bogota. [Cundinamarca, Colombia] 89-151/. NHMUK.

Pyrrhopyge thericles rileyi Bell, 1931

Pls. 24–25, Figs. 7, 9, 11 (♂), 8, 10 (♀) (d, v); Pl. 76, Figs. 6 (♂) (d, v)

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia and central-northern, northeastern to southeastern Brazil.

Colombia: Cundinamarca (*Medina*); Meta (*Villavicencio*). **Ecuador. Peru:** Cusco (*Cosñipata Valley – Chontachac 950 m, Quincemil*); Junín (*Satipo – 100 km SE*); Madre de Dios (*Tambopata Reserve*). Puno (*Chaquimayo, Sandia*). **Bolivia:** La Paz (*Caranavi, Chulumani, Ixianas, Puente Villa, Río Zongo, Tumapasa*); Santa Cruz (*Buenavista*). **Brazil:** Rondônia (*Vilhena*); Pernambuco (*Camaragibe*); Mato Grosso (*Diamantino*); Alagoas (*Maceió*); Bahia (*Amargosa, Camacan*); Minas Gerais (*Caratinga, São Roque de Minas*); Espírito Santo (*Alegre, Conceição da Barra, Guarapari, Linhares, Santa Leopoldina, Santa Teresa, São Mateus, Serra – Jacaraípe, Sooretama*); Rio de Janeiro (*Cachoeiras de Macacu, Casimiro de Abreu, Duque de Caxias, Magé, Nova Iguaçu, Poço das Antas, Rio de Janeiro, Silva Jardim – Reserva Biológica Poço das Antas, Teresópolis*).

TYPE:

Pyrrhopyge rileyi Bell, 1931. Holotype male with the following labels: / Type/ Santa Cruz Bolivia/ G453/ *Pyrrhopyge rileyi* Bell Holotype ♂/. AMNH.

Pyrrhopyge thericles raymondi Orellana, [2010]

Pls. 24–25, Figs. 12 (♀), 13 (holotype ♂), 14 (paratype ♀) (d, v)

DISTRIBUTION – Northern Venezuela.

Venezuela: Aragua (*Choroní, Parque Nacional Henry Pittier – Rancho Grande*); Guárico (*Parque Nacional Guatopo – La Colonia*); Portuguesa (*Agua Blanca*).

TYPE:

Pyrrhopyge thericles raymondi Orellana, [2010]. Holotype male from La Colonia, PN Guatopo, Guárico, Venezuela, 550 m, 16-IX-1985, F. Cerdá & A. Chacón leg. MIZA.

***Pyrrhopyge thericles orientis* Bell, 1947**

Pls. 18–19, Figs. 8 (♀) (d, v); Pls. 24–25, Figs. 15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (*Acará, Belém, Capitão Poço, Santo Antonio do Tauá*).

TYPE:

Pyrrhopyge rileyi orientis Bell, 1947. Holotype male with the following labels: / Holotype/ Para, Brazil/ A. M. Moss/ G1813/ *Pyrrhopyge rileyi* sub Spec. *orientis* Bell Holotype ♂/. AMNH.

***Pyrrhopyge amythaon* Bell, 1931**

Four subspecies.

DISTRIBUTION – Central-northern and eastern Colombia to central-eastern Bolivia, Venezuela and northern, central, northeastern to southeastern Brazil.

***Pyrrhopyge amythaon amythaon* Bell, 1931**

Pls. 24–25, Figs. 23–24 (holotype ♂) (d, v); Pl. 76, Figs. 7 (♀) (d, v)

DISTRIBUTION – Northern, central, northeastern to southeastern Brazil.

Brazil: Roraima (*Amajari – Tepequén*); Pernambuco (*Recife*); Mato Grosso (*Diamantino, Vera*); Distrito Federal (*Brasília*); Minas Gerais (*Diamantina, Santana do Riacho – Serra do Cipó*).

TAXONOMY – The male genitalia are very different from those of *Pyrrhopyge josephina* **stat. rest.** a close related species. The arms of the uncus are closer and the notch between the arms is half of the uncus length (Fig. 16 – *P. amythaon podina*); in *P. josephina* **stat. rest.** the arms of the uncus are wide apart and the notch is more than half of the uncus length (Fig. 17); in both species the valvae are variable. The species are sympatric, at least in Bolivia, La Paz, Caranavi. The DNA barcode minimum divergence between specimens from Bolivia, La Paz, Caranavi, of both phenotypes (“*bixae* and *phidias* forms”) (*Pyrrhopyge amythaon podina*) (n= 4) and of *P. josephina josephina* **stat. rest.** (n= 1) from Peru, Amazonas, Huambo, is 2.3%.

TYPE:

Pyrrhopyge amythaon Bell, 1931. Holotype male with the following labels: / Brazil Pernam[buco]/ G 1179/ *Pyrrhopyge amythaon* Bell ♂ Holotype/. AMNH.

***Pyrrhopyge amythaon peron* Bell, 1947**

Pls. 24–25, Figs. 17, 19 (♂), 18, 20 (♀) (d, v)

DISTRIBUTION – Central-northern and eastern Colombia to central-eastern Bolivia and southern Venezuela to northern Brazil.

Colombia: Santander (*Contratación*); Vaupés (*Puerto Matapi*).

Ecuador: Pastaza (*Mera*). **Peru:** Loreto (*Iquitos, Río Putumayo*); San Martín (*Yumbatos*). **Bolivia:** La Paz (*Río Chimate*). **Venezuela:** Bolívar (*between Santa Elena de Uairén and Ikabará*). **Brazil:** Amazonas (*São Paulo de Olivença*); Pará (*Óbidos*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*); Rondônia (*Candeias do Jamari, Porto Velho*).

TYPE:

Pyrrhopyge amythaon peron Bell, 1947. Holotype male with the following labels: / Holotype/ Iquitos [Loreto, Peru] Feb. 16 1932/ G687/ *Pyrrhopyge amythaon* ssp. *peron* Bell Holotype ♂/. AMNH.

***Pyrrhopyge amythaon gradens* Evans, 1951**

Pls. 24–25, Figs. 21 (♂), 22 (paratype ♀) (d, v)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (*Belém, Santo Antonio do Tauá*).

TYPE:

Pyrrhopyge amythaon gradens Evans, 1951. Holotype male with the following labels: / Type – *gradens* Evans/ ♂ Pará/ ♂/ Pará, L. Amazons. [Brazil] H. W. Bates/ 35/ Godman-Salvin Coll. 1912.–23. *Pyrrhopyge hyperici*, Hubn./ NHMUK.

***Pyrrhopyge amythaon podina* Evans, 1951**

Pls. 26–27, Figs. 1 (♀), 2, 4 (♂), 3 (holotype ♂) (d, v); Text-Fig. 16 (♂ genitalia)

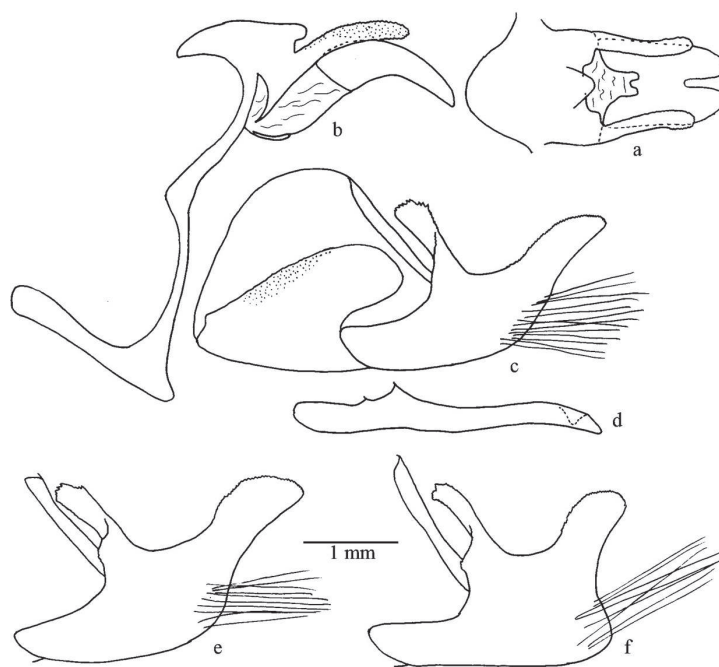
DISTRIBUTION – Northern Peru to central-eastern Bolivia and north-western Brazil.

Peru: Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to Quebrada Santa Isabel 1,200 m, Quillabamba*); Huánuco (*Tingo Maria*); Loreto (*Iquitos*); Puno (*Carabaya – Tinguiri, Oroya – Río Inambari*); San Martín (*Rioja – El Afluente 1,300 m*). **Bolivia:** Cochabamba; La Paz (*Caranavi, Río Zongo*); Santa Cruz. **Brazil:** Rondônia (*Vilhena*).

TAXONOMY – There are two sympatric phenotypes in Bolivia, La Paz, Caranavi, the “*phidias* form” as described by Evans (1951), and the “*bixae* form” with a very variable and light blue band at the base of the ventral hindwing; this band rarely reaches the discocellular veins at the hindwing as in *P. amythaon peron*. There is no difference in the DNA barcode between these two phenotypes. There are also intermediate phenotypes where the band is reduced to a very small spot at the humeral angle. The wings of this taxon are more rounded as in *P. amythaon peron*, where they are more or less excavated in CuA₂–2A, less than in *P. josephina polka* **comb. n.**

TYPE:

Pyrrhopyge amythaon podina Evans, 1951. Holotype male with the following labels: / Type – *podina* Evans/ Bolivie. ex Germain./ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.



Text-Fig. 16. *Pyrrhopyge amythaon podina*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus gnathos and uncus, lateral left view; c) right valva, internal view; d) aedeagus (Bolivia, La Paz, Caranavi, II-III-1989, Tello leg., OM 22.126; e) right valva, internal view of harpe (Brazil, Rondônia, Vilhena, 10-X-1986, Elias leg., DZ 29.137); f) right valva, internal view of harpe (Peru, Huánuco, Tingo Maria, Rondos, XI-1971, Villegas leg., DZ 29.431).

Pyrrhopyge josephina* Draudt, 1920, *stat. rest.

Four subspecies.

DISTRIBUTION – Eastern Colombia to central-eastern Bolivia, southern Venezuela to Suriname and northern Brazil.

***Pyrrhopyge josephina josephina* Draudt, 1920,**

stat. rest.

Pls. 26–27, Figs. 5 (holotype ♂ of *Pyrrhopyge amythaon perula*), 6, 8 (♀), 7 (♂), 9 (holotype ♂) (d, v); Text-Fig. 17 (♂ genitalia)

Syn.: *Pyrrhopyge amythaon perula* Evans, 1951, **syn. n.**

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia and northern Brazil (Amazonas and Rondônia).

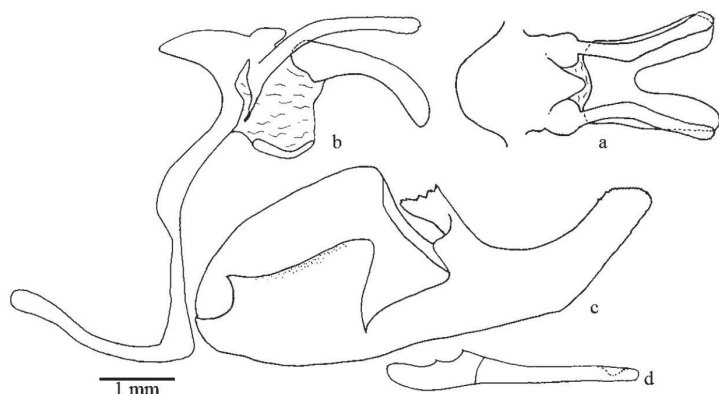
Ecuador: Pastaza (*Mera, Puyo*). **Peru:** Amazonas (*Huambo, Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Quitacalzón 1,100 m, Quincemil*); Huánuco (*Tingo Maria*); Junín (*La Merced, Río Colorado, Río Perené, Satipo, Shimá*); Loreto (*Río Ampiyacu*); Madre de Dios (*Parque Nacional Bahuaja Sonene, Puerto Maldonado, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Huancabamba, Pichis Road*); Puno (*Limbaní, Oroya*); San Martín (*Juanjuí, Moyobamba – Calzada, Juanjuí, Tarapoto, Tocache*). **Bolivia:** La Paz (*Caranavi, Río Chimate, Río Zongo*). **Brazil:** Amazonas (*São Paulo de Olivença*); Rondônia (*Ariquemes, Cacaulândia*).

TAXONOMY – *Pyrrhopyge josephina* **stat. rest.** is here separated from *Pyrrhopyge amythaon* as a species by morphological and DNA barcode characters. See above under *P. amythaon amythaon*. Both species are sympatric at least in Peru, Cusco, Cosñipata Valley, Quebrada Quitacalzón, 1,100 m (only the “*bixae* form”) and Bolivia, La Paz, Caranavi (“*bixae* and *phidias* forms”). The genitalia of the holotype of *Pyrrhopyge josephina* **stat. rest.** (Fig. 17) confirms the species reinstated status. The fringes of the fore and hindwings are light yellow in the holotype, similar to a few specimens of *Pyrrhopyge mina mina* **stat. rest.** In the only specimen we have of the “*phidias* form” from Bolivia, La Paz, Caranavi, the collar is red and the fringes are white. *Pyrrhopyge amythaon perula* Evans, 1951, the “*bixae* form” of this subspecies is sympatric to *P. josephina josephina* **stat. rest.**, the “*phidias* form”, at least in Bolivia, La Paz, Caranavi, reason for the new synonymy. There is considerable variation in the ventral hindwing: the light blue subbasal band, when present, normally does not reach the cell end, but sometimes it extends beyond it. Evans (1951) mentioned *P. josephina* **stat. rev.**, as a subspecies of *P. sergius*.

TYPE:

Pyrrhopyge josephina Draudt, 1920. The holotype (one male was mentioned) with the following labels: / Río Songo [*sic*] [La Paz] Bolivia 750 m Coll. Fassl; and the following two will be added: / Holotypus/ Holotypus *Pyrrhopyge josephina* Draudt, 1920, O. Mielke, Brockmann & C. Mielke det. 2019/. ZfB.

Pyrrhopyge amythaon perula Evans, 1951. Holotype male with the following labels: / Type – *perula* Evans/ La Merced, [Junín] Peru, 3000 ft. XI. 02-I. 03. H. Watkins/ Adams Bequest. B.M. 1912-399/. NHMUK. **Syn. n.**



Text-Fig. 17. *Pyrrhopyge josephina josephina* **stat. rest.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; f) aedeagus, lateral left view (holotype).

***Pyrrhopyge josephina polka* Evans, 1951, comb. n.**

Pls. 26–27, Figs. 10 (holotype ♂ of *Pyrrhopyge amythaon polka*), 11 (♀) (d, v)

DISTRIBUTION – Southern Venezuela to Suriname and northern Brazil (Roraima and Pará).

Venezuela: Amazonas (*Caño Matucha/ Caño Wiraki*). **Suriname:** Sipaliwini (*Kabalebo*). **French Guiana:** Cayenne (*La Comté*); Saint-Laurent-du-Maroni (*Saint-Jean-du-Maroni, Saint-Laurent-du-Maroni*). **Brazil:** Roraima (*Surucucu*); Pará (*Óbidos*).

TAXONOMY – The male genitalia justifies the new combination. **TYPE:**

Pyrrhopyge amythaon polka Evans, 1951. Holotype male with the following labels: / Type – *polka* Evans/ 64. 21. St. Jean de Maroni, French Guiana. Jan.-Mar/ J.Joycey Coll. B.M. 1925-451/. NHMUK.

***Pyrrhopyge josephina orino* Evans, 1951, comb. n.**

Pls. 26–27, Figs. 12 (holotype ♂ of *Pyrrhopyge amythaon orino*), 13 (♀) (d, v)

DISTRIBUTION – Eastern Colombia to Guyana.

Colombia: Vichada (*Maipures*). **Venezuela:** Amazonas (*Maraca*); Bolívar (*Auyán Tepui – Guayaraca, Cuchivero, Río Caura, Roraima*). **Guyana.**

TAXONOMY – The male genitalia justifies the new combination. **TYPE:**

Pyrrhopyge amythaon orino Evans, 1951. The holotype male with the following labels: / Type – *orino* Evans/ Maipures, Orinoco, [Vichada, Colombia] Dec. 98. (Cherrie)./ Rothschild Bequest B.M. 1939-1/. NHMUK.

***Pyrrhopyge josephina pollio* Evans, 1951, comb. n.**

Pls. 26–27, Figs. 14 (holotype ♂ of *Pyrrhopyge amythaon pollio*) (d, v)

DISTRIBUTION – Southeastern Peru.

Peru: Puno (*Chaquimayo*).

TAXONOMY – The male genitalia justify the new combination. **TYPE:**

Pyrrhopyge amythaon pollio Evans, 1951. Holotype male with the following labels: / Type – *pollio* Evans/ Chaquimayo, [Puno] S. Peru. 2500-3000 ft. 8-10 '10. H. & C. Watkins 1121/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Pyrrhopyge divisor* O. Mielke & Casagrande, sp. n.**

Pls. 26–27, Figs. 15 (holotype ♂), 16 (paratype ♂) (d, v); Text-Fig. 18 (♂ genitalia)

DISTRIBUTION – Eastern Peru to northwestern Brazil (Acre).

Peru: Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m*); Loreto (*Iquitos – Palo Seco*). **Brazil:** Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*).

DESCRIPTION – Male: “*bixae* form”. Forewing length 21–22 mm. Ground colour black with some bluish iridescence. Head, palpi and anal tuft red. Fringes white on forewing, from midway M_3 – CuA_1 to tornus. Wings dorsally lighter blue than in other species of the group, similar to *P. infantilis infantilis*. Forewing with outer margin slightly excavated in CuA_2 –2A. Hindwing with outer margin slightly excavated from M_2 to 2A, ventrally with the basal area light bluish white from costal margin to the inner margin, just reaching the end of the cell, interrupted by the dark veins (larger at 2A) and outwardly curved. Genitalia very characteristic; apophyses of tegumen until the end of uncus, straight and pointed; uncus bifid V-shaped; harpe with a wide and blunt expansion, upwards, and constricted at the base.

Female: unknown.

TAXONOMY – A small species of the “*bixae* form”, only possible to identify by an examination of the male genitalia. The straight apophyses of the tegumen, the bifid V-shaped uncus and the typical valva are diagnostic. The bifid uncus aspect with a very broad slit is also shared with *P. josephina* **stat. rest.** and *P. sergius*, suggesting a close relationship.

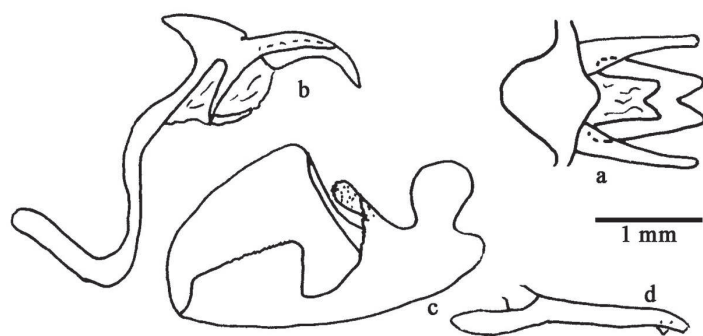
ETYMOLOGY – The specific name refers to the Parque Nacional da Serra do Divisor, the type locality, a large national park in Acre and

close to the Peruvian border.

TYPES:

Holotype male with the following labels: / Holotypus/ 23-30-VIII-2014, Rio Moa, Pq. [Parque] Nac[ional]. Serra do Divisor (Séde), Mâncio Lima, Acre, Brasil, [O.] Mielke, Casagrande, Carneiro, Dias, Dolibaina, Siewert & Salik leg. 7° 26' 52" S, 73° 39' 55" [W]/ Holotypus *Pyrrhopyge divisor* O. Mielke & Casagrande. O. Mielke & Casagrande det. 2017/ gen[itália] prep[arada]., [O.] Mielke 2017/ DZ 37.616/. DZUP.

Paratypes: 4 ♂, same data as the holotype, DZ 29.424, DZ 37.334, DZ 37.398, DZ 37.506 (DZUP); 1 ♂, VIII-2016, Río Itaya, Palo Seco, Iquitos, Loreto, Peru, J. J. Ramírez H. leg., OM 81.104 (OM); 1 ♂, 26-X-2018, Quebrada Quitacalzón, Cosñipata Valley, Cusco, Peru, 1100 m, 13°01' S, 71°30' W, O. Mielke leg. (MUSM); 1 ♂, XI-2011, Iquitos, Loreto, Peru, 100 m, ex coll. Büche (EB).



Text-Fig. 18. *Pyrrhopyge divisor* sp. n. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (Peru, Loreto, Iquitos, Palo Seco, Itaya, VIII-2016, J. J. Ramírez H. leg., paratype, OM 81.104).

***Pyrrhopyge mina* Bell, 1931, stat. rest.**

Three subspecies.

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia, northern Venezuela, Guyana and central-northern and southwestern Brazil.

***Pyrrhopyge mina mina* Bell, 1931, stat. rest.**

Pls. 26–27, Figs. 19, 21–24 (♂), 20 (♀) (d, v); Pls. 30–31, Figs. 19–20 (♂) (d, v)

Syn.: *Pyrrhopyge sergius bolius* Bell, 1947

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia and central-northern and southwestern Brazil.

Peru: Cusco (Cosñipata Valley – Pilcopata 500 m to Quebrada Quitacalzón 1,100 m, Quincemil); Madre de Dios (Puerto Maldonado, Salvación, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 (near Atalaya, Cusco)); Puno (Chaquimayo, Sandia, San Gaban); San Martín (Bellavista). **Bolivia:** La Paz (Caranavi 500–1,000 m, Ixiamas, Mapiri, Río Chimate, Río Zongo, Tumapasa); Santa Cruz (Buenavista, Sara). **Brazil:** Acre (Senador Guimard – Reserva Catuaba); Rondônia (Cacaulândia); Mato Grosso (Cáceres, Chapada dos Guimarães, Diamantino, Pantanal); Goiás (Mineiros, Parque Nacional das Emas); Distrito Federal (Brasília); Paraná (Querência do Norte).

The citations **Argentina:** Santiago del Estero (Evans 1951, Hayward 1973) are probably incorrect.

TAXONOMY – Since Evans (1951), the taxon was identified as *Pyrrhopyge sergius josephina* Draudt, 1920, but this taxon is actually a very close related species to *P. amythaon* Bell, 1931 (see comments at this species). The oldest available name is *Pyrrhopyge mina* stat. rest., so this taxon has its species status reinstated.

The male and female genitalia do not differ from that of *P. sergius*.

Two phenotypes occur in this subspecies, the “phidias form” and the “bixae form”, according to DNA barcode; the latter never

mentioned in literature. In the “bixae form” the blue white band on the ventral hindwing rarely reaches the discocellular veins and narrows towards the costal margin. The “phidias form” occurs together with the “bixae form” at least in Peru, Cusco, Cosñipata Valley, Pilcopata 500 m and Chontachaca 950 m. The latter phenotype also occurs in Brazil, Acre (Mâncio Lima and Senador Guimard).

The DNA barcode minimum divergences are: between *P. mina* stat. rest. (n= 7) and *P. sergius sergius* (n= 5) is 3.06%, and between *P. mina* stat. rest. and *P. michael* sp. n. is 2.60%, so *P. mina* stat. rest. has its status reinstated. The DNA barcode includes the holotypes of *P. mina* stat. rest., *P. sergius bolius* and *P. sergius ganus*.

TYPES:

Pyrrhopyge mina Bell, 1931. Holotype male with the following labels: / Type/ Bolivia/ 813/ G416/ *Pyrrhopyge mina* Bell Holotype ♂/. AMNH.

Pyrrhopyge sergius bolius Bell, 1947. Holotype male with the following labels: / Holotype/ Rio Chimate [sic, recte Chimate], [La Paz] Boliv[ia]. Apr. 12 26, 1900f/ G1720/ *Pyrrhopyge sergius* ssp. *bolius* Bell Holotype ♂/ F 41451 Acc. 27264/. AMNH.

***Pyrrhopyge mina ganus* Bell, 1947, comb. n.**

Pls. 26–27, Figs. 17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Northern Venezuela and Guyana.

Venezuela: Aragua (Choroní, Maracay, Parque Nacional Henry Pittier – Rancho Grande, Romerito). **Guyana.**

The citations **Colombia**, **Ecuador**, and **Peru:** Huánuco; Pasco (Pozuzo) (Evans 1951) are probably incorrect.

TAXONOMY – The DNA barcode of the holotype reveals that this taxon is related to *P. mina* stat. rest. The divergence between *P. mina* stat. rest. (n= 7) and *P. mina ganus* comb. n. (n= 1) is 0.60%, reason for the new combination.

TYPE:

Pyrrhopyge sergius ganus Bell, 1947. Holotype male with the following labels: / Holotype/ Br[itish]. Guiana/ Collector Dr. C. L. Pollard/ G 1262/ *Pyrrhopyge sergius* ssp. *ganus* Bell Holotype ♂/. AMNH.

***Pyrrhopyge mina previsto* O. Mielke, Brockmann & C. Mielke, ssp. n.**

Pls. 28–29, Figs. 1 (holotype ♂) (d, v); Text-Fig. 19 (♂ genitalia)

DISTRIBUTION – Southeastern Peru.

Peru: Ucayali (*Previsto*).

DESCRIPTION – Male forewing length 24 mm. A subspecies of *Pyrrhopyge mina* stat. rest. of the “sergius form” very similar to *P. jola* sp. n.. Head, palpi and anal tuft red. Dorsal fore and hindwings, and ventral forewing unmarked; ventral hindwing with a white marginal white band at the external margin between Sc+R₁ and 2A–3A, interrupted by the dark veins, reaching nearly 2 mm in M₃–CuA₁ and CuA₁–CuA₂, where basal margin of the white band convex, not straight as in *P. jola* sp. n.. Fringes white, at the forewing between M₂ and tornus, and at the hindwing between Sc+R₁ and the anal angle. The male genitalia are of the “*Pyrrhopyge sergius* group” and very similar to *Pyrrhopyge sergius andronicus*: the flanges of the tegumen are slightly curved inwards at the end, slightly curved upwards at the end of the basal third, and then more or less straight.

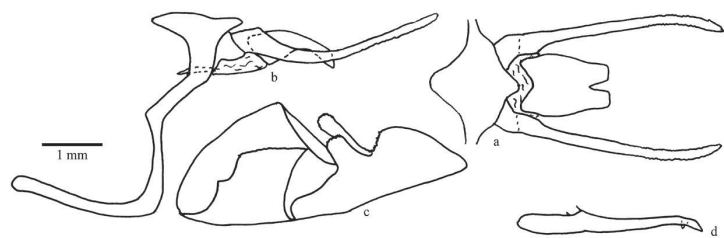
The DNA barcode divergence between *Pyrrhopyge mina previsto* ssp. n. (n= 1) and *P. mina* stat. rest. (n= 7) is 1.07%, between *P. mina previsto* ssp. n. and *P. sergius sergius* (n= 5) 3.98%, between *P. mina previsto* ssp. n. and *P. jola* sp. n. (n= 4) 3.49%, between *P. mina previsto* ssp. n. and *P. michael* sp. n. (n= 24) 3.67%, and between *P. mina previsto* ssp. n. and *P. phylleia* (n= 14) 3.82%.

REMARKS – Only known from the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ Peru [Ucayali] Previsto [9°3'S, 75°38'W] 18.IX.1975 ex coll. P. Kueppers, Karlsruhe via F. Stumpe an mich/ Holotypus *Pyrrhopyge mina previsto* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C.

Mielke det. 2019/ Sequenced for Barcode at Biodiversity Institute of Ontario HESP-EB 01 119/ DNA sample ID: NVG-15081H05 c/o Nick V. Grishin/. MSFR.



Text-Fig. 19. *Pyrrhopyge mina previsto* **ssp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

Pyrrhopyge sergius Hopffer, 1874

Five subspecies.

DISTRIBUTION – Western, central and eastern Colombia to western Ecuador and to central-eastern Peru, southwestern Venezuela, Suriname, French Guiana and northern Brazil.

TAXONOMY – The DNA barcodes of many of the *Pyrrhopyge* species show that *P. sergius*, with two subspecies (*P. sergius sergius* and *P. sergius andronicus*), is closely related to *P. mina* **stat. rest.**, *P. michael* **sp. n.** and *P. jola* **sp. n.**, comprising a complex of four species.

The DNA barcode minimum divergences are: between *P. sergius sergius* (n= 5) and *P. mina* **stat. rest.** (n= 7) – 3.06%, between *P. sergius sergius* and *P. michael* **sp. n.** (n= 24) – 2.29%, and between *P. sergius sergius* and *P. jola* **sp. n.** (n= 4) – 2.36%, between *P. jola* **sp. n.** and *P. michael* **sp. n.** (n= 14) – 1.90%, between *P. jola* **sp. n.** and *P. mina* **stat. rest.** – 2.70%, and between *P. mina* **stat. rest.** and *P. michael* **sp. n.** – 2.60%.

The lectotype of *Pyrrhopyge sergius*, and the holotypes of *P. andronicus*, *P. mina* **stat. rest.**, *P. sergius ganus* and *P. sergius bolius* were included in the barcode analysis. Other subspecies of *P. sergius* (*P. sergius andros*, *P. sergius selina* and *P. sergius semana*) were not barcoded.

The male genitalia of the species of the “*P. sergius* species group” are variable, mainly in the shape of the harpe, which is triangular and enlarged (Text-Fig. 21 – *P. sergius sergius*) or more triangular and elongated (Text-Figs. 20 – *P. sergius sergius*, holotype, 22 – *P. sergius andronicus*); also by the apophyses of the tegumen which are evenly curved upwards (Text-Figs. 20 – *P. sergius sergius*, holotype, 21 – *P. sergius sergius*, 25 – *Pyrrhopyge jola* **sp. n.**), slightly angled (Text-Fig. 22 – *P. sergius andronicus*) or nearly straight, and by the dorsal and internal projection at the base of the harpe which is straight in *P. sergius* (Text-Figs. 21, 22) and *P. mina* **stat. rest.** (Text-Fig. 19 – *P. mina previsto* **ssp. n.**), or slightly curved to the base as in *P. michael* **sp. n.** (Text-Fig. 23). The female genitalia of *Pyrrhopyge sergius*, *P. mina* **stat. rest.** and *P. michael* **sp. n.** are similar as illustrated (Fig. 24 – *P. michael* **sp. n.**).

Pyrrhopyge sergius sergius Hopffer, 1874

Pls. 28–29, Figs. 2 (lectotype ♂), 3 (♂), 4 (♀) (d, v); Text-Figs. 20–21 (♂ genitalia)

DISTRIBUTION – Central-eastern Peru.

Peru: Junín (*Chanchamayo*, *San Ramón*); Pasco (*Pozuzo*); San Martín (*Tocache*); all identified by DNA barcode.

Some more records are available, but specimens were not examined (some may be *P. michael* **sp. n.**): Cusco (*Aguas Calientes*, *Cashiriari*, *Illapani Viejo*, *Machu Picchu*, *Marcapata*, *Quillabamba*, *Río Urubamba*); Huánuco (*Puente Cayumba*); Junín (*La Merced*, *Río Colorado*, *Río Anapati Valley*); Pasco (*Huancabamba*, *Pichis Road*).

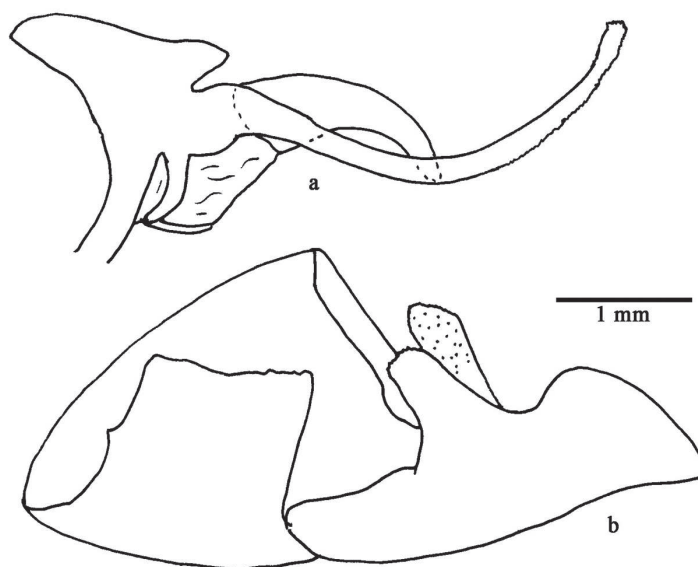
TAXONOMY – As this subspecies of the “*sergius* form” is very similar to *Pyrrhopyge michael* **sp. n.**, the male genitalia are figured. The apophyses of the tegumen are slightly curved upwards and inwards at the distal point, the harpe is straight with the inner arm at the base

not curved slightly to the base of the valva, as in *P. michael* **sp. n.** The female genitalia of both species are also similar (Fig. 24 – *P. michael* **sp. n.**).

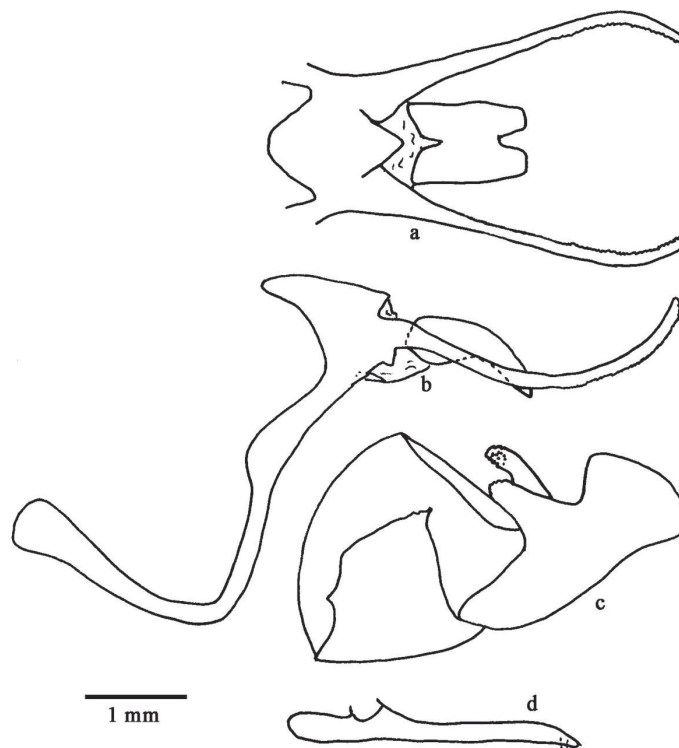
P. sergius sergius is very similar to the allopatric “*sergius* form” of *P. michael* **sp. n.**

TYPE:

Pyrrhopyge sergius Hopffer, 1874. The original description mentions an unspecified number of males from Chanchamayo, [Junín, Peru], Thamm leg. To ensure the correct identification of *Pyrrhopyge sergius*, one syntype male is **herein designated as the lectotype**: this specimen with the following labels: / Lectotypus/ Chanchamayo [Junín] Peru Staudgr/ 19786/ *Sergius* Hpfr * Stett. Ent. Ztg. 1874. P 369/ Genital-Unters. Nr. 4722 Zool. Mus. Berlin/ gen[itália]. prep[arada]. [O.] Mielke 1979/; another label will be added: / Lectotypus *Pyrrhopyge sergius* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published.



Text-Fig. 20. *Pyrrhopyge sergius sergius*. Male genitalia: a) tegumen and uncus, lateral view; b) right valva, internal view (lectotype).



Text-Fig. 21. *Pyrrhopyge sergius sergius*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (Peru, Junín, San Ramón, 5 Km SE Vitoc, Río Tulumayo, 19-X-1989, 1100 m, O. Mielke & Casagrande leg., OM 23.585).

***Pyrrhopyge sergius andros* Evans, 1951**

Pls. 28–29, Figs. 5 (holotype ♂), 6 (♂) (d, v)

DISTRIBUTION – Central and eastern Colombia to southwestern Venezuela.

Colombia: Caquetá (*Río Orteguaza*); Cundinamarca (*Bogotá*).

Venezuela: Amazonas (*Cerro Yutagé*).

TYPE:

Pyrrhopyge sergius andros Evans, 1951. Holotype male with the following labels: / Type – *andros* Evans/ Colombie Reçu du Frère Apollinaire Marie de Bogota/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Pyrrhopyge sergius andronicus* Bell, 1931**

Pls. 28–29, Figs. 7 (♂), 8 (♀) (d, v); Text-Fig. 22 (♂ genitalia)

DISTRIBUTION – Colombia to western and eastern Ecuador.

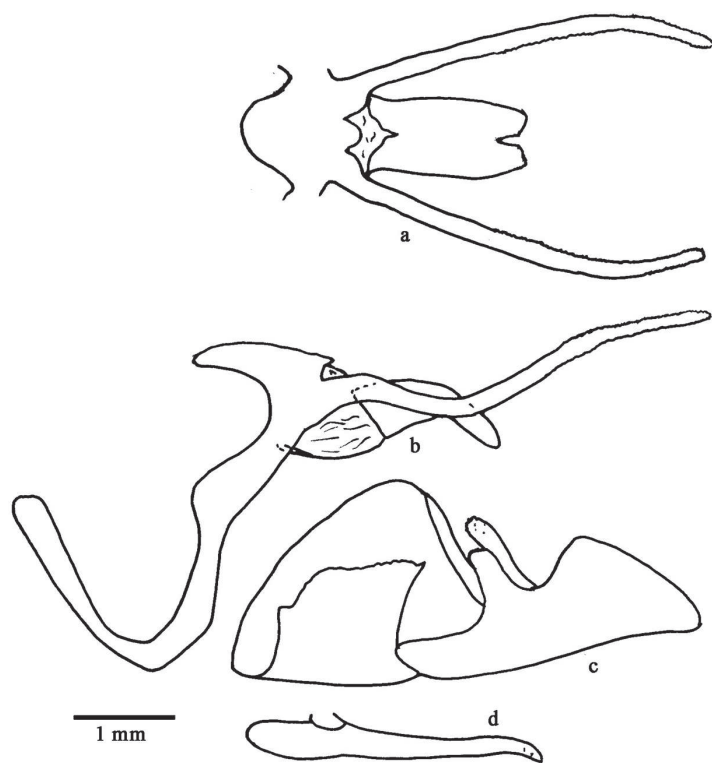
Colombia. Ecuador: Loja (*San Francisco*); Morona-Santiago (*Canari, Macas, Pablo Sexto, Sucúa*); Napo (*Cotundo, Misahualli, Río Napo, Tola, Yanahurco*); Pastaza (*Mera*); Pichincha (*Pacto*); Tungurahua (*Yunguilla*); Zamora-Chinchi (Zamora).

The citations **Argentina:** Misiones (*Puerto Libertad* – as *Puerto Bemberg*) (Hayward 1940a, 1941, 1948, 1973 and Canals 2003) are probably incorrect.

TAXONOMY – The DNA barcode minimum divergence between *P. sergius sergius* (n= 5) and *P. sergius andronicus* (n= 4), including the holotype is 1.67%.

TYPE:

Pyrrhopyge andronicus Bell, 1931. Holotype male with following labels: / Type/ Macas [Morona-Santiago] Ecuador/ I59/ G383/ *Pyrrhopyge andronicus* Bell Holotype ♂/. AMNH.



Text-Fig. 22. *Pyrrhopyge sergius andronicus*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (Ecuador, Napo, Yanahurco, 5-IX-1978, 550, Lafebre leg., OM 37.060).

***Pyrrhopyge sergius semana* Evans, 1951**

Pls. 28–29, Figs. 9 (♂), 10 (♀) (d, v)

DISTRIBUTION – Suriname, French Guiana and northern Brazil (Amazonas and Pará).

Suriname: Brokopondo (*Brownsberg*); Paramaribo; Sipaliwini (*Bakhuis Gebergte, Suriname*). **French Guiana:** Cayenne (*Fourgassier, Montagne de Kaw, Soula, Tonaté*); Saint-Laurent-du-Maroni (*Saint-*

Laurent-du-Maroni). **Brazil:** Amazonas (*Fonte Boa, São Gabriel da Cachoeira*); Pará (*Óbidos*).

TYPE:

Pyrrhopyge sergius semana Evans, 1951. Holotype male with the following labels: / Type – *semana* Evans/ St. Laurent [du Maroni], Maroni River, French Guiana. July-Sept. 1915./. NHMUK.

***Pyrrhopyge sergius selina* Evans, 1951**

Pls. 28–29, Figs. 11 (♂), 12 (♀) (d, v)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (*Belém*).

The citations **Colombia:** Putumayo (Salazar 1995) and East Cordillera (Andrade 2002) are probably incorrect.

TYPE:

Pyrrhopyge sergius selina Evans, 1951. Holotype male with the following labels: / Type – *selina* Evans/ Para, [Brazil]/ Adams Bequest. B.M. 1912-399./. NHMUK.

***Pyrrhopyge michael* O. Mielke, Brockmann & C. Mielke, sp. n.**

Pls. 28–29, Figs. 13 (holotype ♂), 14 (allotype ♀), 15, 17–18 (paratype ♂), 16, 19 (paratype ♀) (d, v); Text-Figs. 23 (♂ genitalia), 24 (♀ genitalia)

DISTRIBUTION – Northern to central-eastern Peru and northern Brazil (Amazonas and Acre).

Peru: Amazonas (*Chachapoyas – Yambrasbamba* – “*bixae* form”, *Huambo* – “*bixae* form”, *Rodríguez de Mendoza* – “*bixae* form” and “*phidias* form” with a few blue white scales at the costal margin); Huánuco (*Carpish, Tingo Maria* – “*sergius* form”); Junín (*Río Perené* – “*sergius* form”); Loreto (*Iquitos* – “*bixae* form”); San Martín (*Juanjuí* – “*bixae* form”, “*sergius* form” and “*phidias* form”, *Rioja – Mina de Sal* – “*bixae* form”, *Moyobamba* – “*bixae* form”, *Rioja – San Agostin 1,000 m to El Afluente 1,300 m* – “*bixae* form”, *Santa Cruz* – “*bixae* form”). **Brazil:** Amazonas (*São Paulo de Olivença* – “*bixae* form”); Acre (*Porto Walter* – “*bixae* form”).

The following localities mentioned by Evans (1951) under *Pyrrhopyge sergius andronicus* probably refer to this species: **Peru:** Cajamarca; Loreto (*Caballococha, Río Marañon, San Antonio, Upper Río Putumayo*); San Martín (*Yumbatos*).

DESCRIPTION – Male: forewing length 23–28 mm. Species similar to other species of the “*phidias* group”. Ground colour dark black. Head, palpi and anal tuft red. Dorsal fore and hindwings, and ventral forewing without markings; ventral hindwing of the “*phidias* form”, or of the “*sergius* form” (holotype and allotype) or “*bixae* form”. In the “*sergius* form”, the ventral hindwing with the outer margin wide, light blue (4–5 mm) between Sc+R₅ until 2A and very small (ca. 1 mm) between 2A–3A, inwardly sometimes irregular and with dark veins; very similar to *P. sergius sergius*. In the “*bixae* form”, the ventral hindwing with the basal blue white band narrow, not reaching the end of the discal cell, and narrower at the costal margin. Fringes white, on the forewing from M₂–M₃ to tornus, and at the hindwing from Sc+R₁ to 2A–3A, wider at the hindwing.

Genitalia similar to those of *P. sergius sergius*, but the apophyses of the tegumen are sometimes more angled upwards and inwards at the end, and inner arm at the base of the harpe slightly curved to the base (straight in *P. sergius*).

Female: forewing length 28–32 mm. Similar to male, but with rounder wings. Genitalia similar to *P. sergius sergius*.

The DNA barcode minimum divergences between close related species are: *P. michael* sp. n. (n= 24) and *P. sergius sergius* (n= 5) – 2.60%, between *P. michael* sp. n. and *P. mina mina* stat. rest. – 2.60%, between *P. michael* sp. n. and *P. phylleia* – 1.99%, and between *P. jola* sp. n. and *P. michael* sp. n. (n= 24) – 1.90%. There are no difference in the DNA barcode between specimens of the “*sergius* form”, the “*bixae* form” and the “*phidias* form”. The three phenotypes are sympatric at least in Juanjuí, San Martín, Peru, and the “*bixae* form and *phidias* forms” in Huambo and Rodríguez de Mendoza, Amazonas, Peru.

A correct identification is only possible by a DNA barcode examination.

ETYMOLOGY – The specific name is in honour of Michael Büche, a good friend who provided us with many Peruvian butterflies. TYPES:

Holotype (“*sergius* form”) male with the following labels: / Holotypus/ Tingo Maria, Huánuco, Peru, 670 m, Büche leg./ gen[itá]lia. prep[arada]. [O.] Mielke 2017/ Holotypus *Pyrrhopyge michael* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 35.718 (OM-DZUP).

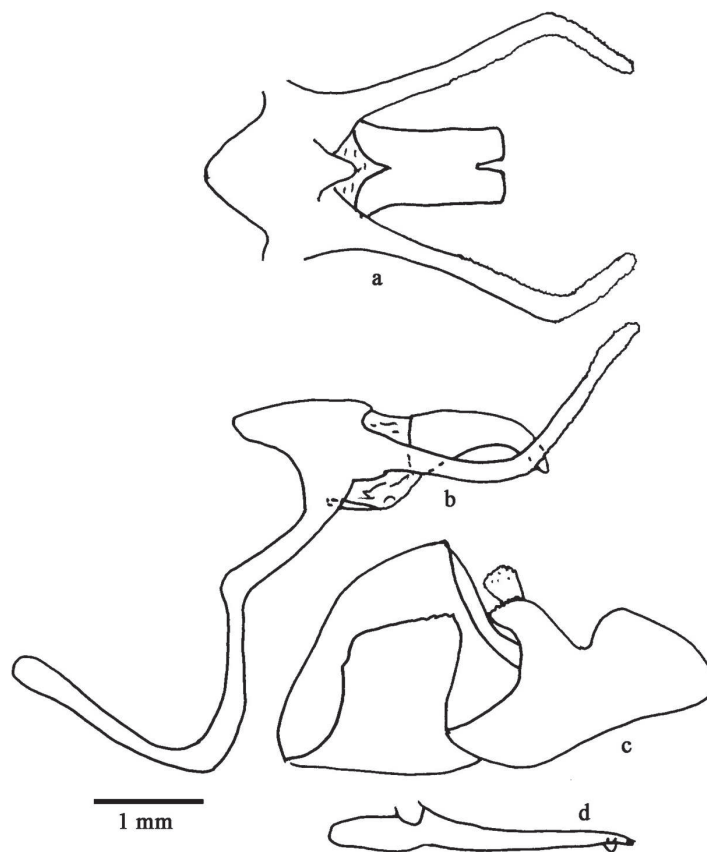
Allotype (“*sergius* form”) female with the following labels: / Allotypus/ III-1997 Tingo Maria, [Huánuco] Peru, M. Büche leg./ gen[itá]lia. prep[arada]. [O.] Mielke 2017/ Allotypus *Pyrrhopyge michael* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ BC-OM [barcoded for O. Mielke] OM 47.078 (OM-DZUP).

Paratypes (“*sergius* form”): 2 ♂, V-1964, 1 ♂, XI-1964, 1 ♂, V-1965, 1 ♂, 23-X-1980, 2 ♂, 20-II-1982, Tingo Maria, Huánuco, Peru, OM 7.063, OM 7065, OM 7.064, OM 10.840, OM 26.315, OM 18.379, OM 18.380 (OM); 1 ♂, 27-VIII-1991, 1 ♀, 6-IX-1991, 1 ♂, 26-X-1991, 1 ♂, 4-IV-1992, 1 ♂, 16-V-1992, 1 ♂, 20-II-1993, Tingo Maria, Huánuco, Peru, 670 m, Büche leg., OM 35.635, OM 35.585, OM 35.647, OM 35.499, OM 35.907, OM 35.876 (OM); 3 ♂, 16-26-X-1971, 7 ♂, 27-X-1971, 30 ♂, XI-1971, Rondos, Tingo Maria, Huánuco, Peru, 850 m, Villegas leg., DZ 40.524–40.563 (DZUP); 1 ♂, I-1981, Río Perené, Junín, Peru, 550 m, OM 71.308 (OM); 1 ♂, 22-X-2002, Juanjuí, San Martín, Peru, 280 m, OM 60.266 (OM); 1 ♂, Peru, OM 35.247 (OM); 1 ♂ 1 ♀, 1992, Tingo Maria, Huánuco, Peru, Rojas Villegas leg., EB 01 0120, EB 01 111 (EB); 1 ♂, 16-XII-1986, 1 ♂, 1-II-1987, 1 ♂, 11-II-1987, 1 ♂, 22-XII-1987, 1 ♀, IV-2010, Tingo Maria, Huánuco, Peru, via M. Büche, EB 01 112, EB 01 118 [aberration], EB 01 246, EB 01 247 (EB); 1 ♂, 6-7-IV-2019, Nuevo Jean, San Martín, ex coll. J. Ramirez Hernandez (EB); 1 ♂, 4-X-2014 (Genitalia-160138), 1 ♂, 2-X-2015 (Genitalia-160154), 3 ♂, 2-X-2014, 1-X-2015, 26-I-2018, Tingo Maria, Huánuco, Peru, J. Uehara leg. (CJU); 1 ♂, XII-2007, 1 ♂ 2 ♀, IX-2015, Tingo Maria, Huánuco, Peru (CJU).

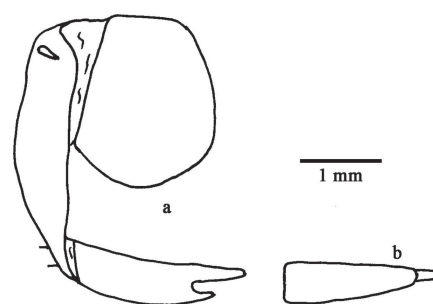
Paratypes (“*bixae* form”): 1 ♂, 22-X-2002, Huambo, Amazonas, Peru, 1000 m, OM 60.300 (OM); 1 ♂, X-2009, 1 ♀, I-2016, Picuroyacu, río Amazonas, Iquitos, Loreto, Peru, J. J. Hernández leg., OM 80.477, OM 79.286 (OM); 1 ♂, V-2017, Villa Hermosa, Yamborasbamba, Chachapoyas, Amazonas, Peru, J. Pintado leg., OM 86.530 (OM); 1 ♂, XII-2014, Moyobamba, San Martín, Peru, 800 m, C. Tello leg., OM 77.256 (OM); 1 ♂, 25-VIII-2002, 1 ♂, 21-X-2002, 1 ♂, 22-X-2002, 1 ♂, 10-XI-2002, Juanjuí, San Martín, Peru, 280 m, OM 60.481, OM 61.849, OM 60.410, OM 60.211 (OM); 1 ♂, 24-II-2003, Juanjuí, San Martín, Peru, OM 63.624 (OM); 1 ♂, 5-VII-2012, 2 ♂, 29-VII-2012, Pañasapa, Juanjuí, San Martín, Peru, C. Calderon leg., OM 73.182, OM 73.192, OM 73.212 (OM); 1 ♂, XI-2016, 4 ♂, V-2017, 1 ♂, VII-2017, El Afluente, Rioja, San Martín, Peru, 1300 m, J. Pintado leg., OM 83.334, OM 86.537, OM 86.544, OM 86.550, OM 86.568, OM 86.569 (OM); 1 ♂, Naranjillo, Rioja, San Martín, Peru, 1000 m, Tafur Novoa leg., OM 68.703 (OM); 2 ♂, Naranjos, Rioja, San Martín, Peru, 1200 m, Tafur Novoa leg., OM 68.601, OM 68.643 (OM); 1 ♂, I-2007, 1 ♂, II-2007, Naranjos, Rioja, San Martín, Peru, 1300 m, J. M. Zevallos leg., OM 68.292, OM 60.050 (OM); 1 ♂, X-2016, 2 ♂, XI-2016, San Agostin, Rioja, San Martín, Peru, 1000 m, J. Pintado leg., OM 83.529, OM 83.489, 83.519 (OM); 1 ♂, IX-2016, 1 ♂, IX-2017, Santa Cruz, San Martín, Peru, 1200 m, J. Pintado leg., OM 81.317, OM 86.538 (OM); 1 ♂, III-1961, São Paulo de Olivença, Amazonas, Brazil, OM 11.640 (OM); 1 ♂, Porto Walter, Acre, Brazil, OM 36.171 (OM); 1 ♂, 25-V-1994, 1 ♂, 2-VIII-1994, 1 ♂, 10-VIII-1994, 1 ♂, 10-VI-1995, 1 ♂, 10-X-1995, 1 ♂, 26-VIII-1996, 1 ♂, 15-VI-1997, 2 ♂, 12-X-1997, 1 ♂, 18-XII-1997, 1 ♂, 2-IX-1999, Rodríguez de Mendoza, Amazonas, Peru, ex coll. F. König, EB 01 275, EB 01 279, EB 02 569, EB 02 570, EB 02 572, EB 02 573, EB 14 065, EB 14 079, EB 14 085, EB 14 087, EB 14 088 (EB); 2 ♂, VI-VII-2007, Mina de Sal, Rioja, San Martín, Peru, ex coll. R. Sturm, EB 00 249, EB 00 248 (EB); 1 ♀, 1-X-1998, 2 ♂, 17-XII-1998, 1 ♂, 11-XI-2000, 1 ♂, 26-XII-2001,

Juanjuí, San Martín, Peru, EB 14 046, EB 14 054, EB 14 055, EB 14 069, EB 14 057 (EB).

Paratypes (“*phidias* form”): 1 ♂, XII-1999, Juanjuí, San Martín, Peru, EB 01 256 (EB); 1 ♀, 2-III-2002, Río Huambo, 40 km W Juanjuí, San Martín, 1400 m, Peru, ex coll. F. König, EB 00 457 (EB); 1 ♂, 12-X-1997, Rodríguez de Mendoza, Peru, EB 14 079 (EB).



Text-Fig. 23. *Pyrrhopyge michael* **sp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (Peru, Huánuco, Tingo Maria, 20-II-1993, 670 m, M. Büche leg., OM 35.876, paratype).



Text-Fig. 24. *Pyrrhopyge michael* **sp. n.** Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (allotype).

Pyrrhopyge jola O. Mielke, Brockmann & C. Mielke, **sp. n.**

Pls. 28–29, Figs. 20 (holotype ♂) (d, v); Text-Fig. 25 (♂ genitalia)

DISTRIBUTION – Central-eastern to southern Peru.

Peru: Cusco (*Cosñipata Valley – Chontachaca 950 m to Mirador 1,720 m, Marcapata*); Junín (*Río Colorado*); Pasco (*Huancabamba, Parque Nacional Yanachaga Chemillén, Puerto Davis, Río Pichis*).

DESCRIPTION – Male “*sergius* form”. Forewing length 23–27 mm. Species similar to other species of the group. Ground colour black with some bluish iridescence. Head, palpi and anal tuft red. Dorsal fore and hindwings, and ventral forewing without markings, ventral hindwing with a marginal light blue-white band, not exceeding

3 mm, at the external margin between Sc+R₁, or M₁, and 2A–3A, interrupted by the dark veins. Fringes white, on the forewing from M₂–M₃ to tornus, and at the hindwing from Sc+R₁ to 2A–3A, wider at the hindwing.

Female: unknown.

TAXONOMY – A species morphologically very similar in its “*sergius* form” to *P. sergius*, *P. mina* **stat. rest.** and *P. michael* **sp. n.**, but differs from all of them by the DNA barcode. The DNA barcode minimum divergence between *P. jola* **sp. n.** (n= 4) and *P. sergius sergius* (n= 5) is 2.36%, between *P. jola* **sp. n.** and *P. mina mina* **stat. rest.** (n= 7) is 2.70% and between *P. jola* **sp. n.** and *P. michael* **sp. n.** (n= 24) is 1.90%. The species is also very similar morphologically and genetically by the DNA barcode to *P. phylleia*; the minimum divergence is 0.89%, but the wing colour ornamentation is very distinct, including white fringes on the forewing only from M₂–M₃ until the tornus, while the yellow fringes of *P. phylleia* are from the apex until the tornus. Also *P. jola* **sp. n.** is sympatric with *P. mina mina* **stat. rest.** in the Cosñipata Valley, Cusco, Peru, between Chontachaca 950 m and Quebrada Quitacalzón 1,100 m.

Pyrrhopyge jola **sp. n.** is easily distinguished in this complex by the ventral hindwing with a marginal light blue-white band, not exceeding 3 mm, between Sc+R₁ or M₁, to 2A–3A, interrupted by the dark veins.

ETYMOLOGY – The specific name is in honour of Ernst Brockmann’s wife, Jolanthe (“Jola”).

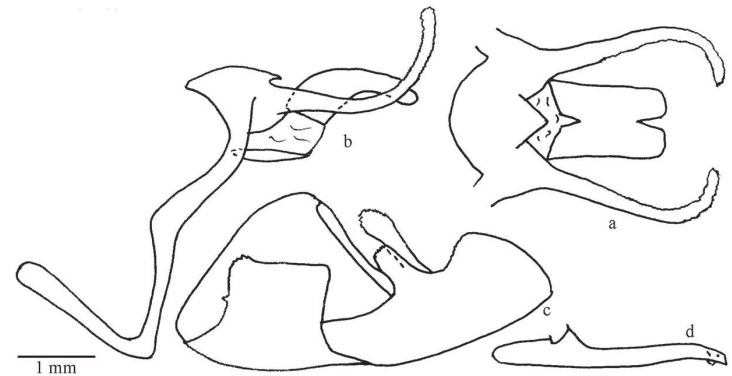
TYPES:

Holotype male with the following labels: / Holotypus/ 30-X-2013 [Quebrada] Santa Isabel, Cosñipata Valley, Cusco, Peru, 13° 02’ S, 71° 31’ W, 1200 m. O. Mielke leg./ Holotypus *Pyrrhopyge jola* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/. MUSM.

Paratypes: 7 ♂, 4–8-XI-2017, Chontachaca, Cosñipata Valley, Cusco, Peru, 950 m, 13°02’ S, 71°28’ W, Huanan V. leg. OM 84.665, OM 84.685, OM 84.705, OM 84.715, OM 84.735 (5 ♂ OM, 2 ♂ EB); 2 ♂, 8-XI-2007, 1 ♂, 12-II-2011, 1 ♂, 22-IX-2014, Quebrada Quitacalzón, Cosñipata Valley, Cusco, Peru, 1100 m, 13°01’ S, 71°30’ W, G. Lamas leg. (MUSM); 1 ♂, 29-I-2010, 1 ♂, 25-IX-2011, Quebrada Quitacalzón, Cosñipata Valley, Cusco, Peru, 1100 m, 13°01’ S, 71°30’ W, L. Gibson leg. (MUSM); 2 ♂, 2-II-2013, 6 ♂, 19-X-2013, 2 ♂, 2-XI-2017, 2 ♂, 10-XI-2017, 2 ♂, 27-X-2018, Quebrada Quitacalzón, Cosñipata Valley, Cusco, Peru, 1100 m, 13°01’ S, 73°30’ W, O. Mielke leg., OM 74.646, OM 74.843, OM 75.559, OM 75.649, OM 75.889, OM 75.899, OM 75.959, OM 76.039, OM 84.777, OM 84.951, OM 84.967, OM 85.001, OM 87.952, OM 87.976 (OM); 1 ♂, 4-X-2011, Quebrada Santa Isabel, Cosñipata Valley, Cusco, Peru, 1200 m, 13° 02’ S, 71° 31’ W, G. Lamas leg. (MUSM); 2 ♂, 19-X-2013, 1 ♂, 30-X-2013, 1 ♂, 22-X-2016, 3 ♂, 24-X-2016, 2 ♂, 1-XI-2017, 2 ♂, 6-XI-2018, Quebrada Santa Isabel, Cosñipata Valley, Cusco, Peru, 1200 m, 13°02’ S, 71°31’ W, O. Mielke leg., OM 75.709, OM 75.799, OM 76.530, OM 82.976, OM 82.309, OM 82.468, OM 82.478, OM 84.608, OM 84.638, OM 87.467, OM 87.498 (OM); 1 ♂, 6-XI-2001, 1 ♂, 7-XI-2001, 1 ♂, 9-XI-2001, 1 ♂, 20-X-2013, San Pedro, Cosñipata Valley, Cusco, Peru, 1400 m, 13°03’ S, 71°33’ W, G. Lamas leg. (MUSM); 11 ♂, 5-10-XI-2001, 2 ♂, 19-X-2013, 1 ♂, 20-X-2013, San Pedro, Cosñipata Valley, Cusco, Peru, 1400 m, O. Mielke leg., OM 53.922, OM 53.929, OM 53.936, OM 54.097, OM 54.237, OM 54.245, OM 54.251, OM 54.252, OM 54.259, OM 75.554, OM 76.570 (OM, 2 ♂, EB); 2 ♂, 23-IX-2011, 1 ♂, 29-X-2013, 1 ♂, 30-X-2013, San Pedro, Cosñipata Valley, Cusco, Peru, 1400 m, 13°03’ S, 71°33’ W, S. Kinyon leg. (MUSM); 1 ♂, 30-X-2013, San Pedro, Cosñipata Valley, Cusco, Peru, 1400 m, 13°03’ S, 71°33’ W, M. McInnis leg. (MUSM); 1 ♂, 5-XI-2012, 3 ♂, 20-X-2013, Quebrada Unión, Cosñipata Valley, Cusco, Peru, 1600 m, O. Mielke leg., OM 73.302, OM 75.543, OM 75.603, OM 75.817 (OM); 1 ♂, 22-X-2013, Puente Unión, Cosñipata Valley, Cusco, Peru, 1600 m, 13°03’ S, 71°32’ W, G. Lamas leg. (MUSM); 1 ♂, 2-XI-2012, Mirador, Cosñipata Valley, Cusco, Peru, 1720 m, 13°04’ S, 71°33’ W, S. Kinyon leg. (MUSM); 1 ♂, Marcapata, Cusco, Peru, OM 11.997 (OM); 1 ♂, Río Pichis, Pasco, Peru (MUSM); 1 ♂, 28-X-1994, 2

♂, 1-XI-1994, 1 ♂, 10-XI-1994, Quebrada Samantoshoari, 5 km SE Puerto Davis, Pasco, Peru, 210 m, 09°59’ S, 74°40’ W, J. Grados & J. Carrasco leg. (MUSM); 6 ♂, 16-IX-2007, Pampa Pescado, Parque Nacional Yanachaga Chemillén, Pasco, Peru, 400–450 m, 10°22’33” S, 75°14’36” W, J. Grados & S. Carbonel leg. (MUSM); 1 ♂, II-1903, Río Colorado, Junín, Peru, 2500 ft., Watkins & Tomlinson leg., R. Oberthür Coll. Brit. Mus. 1931-136, ex coll. British Museum Natural History, OM 36.214 (specimen identified as *Pyrrhopyge sergius sergius* by Evans) (OM); 1 ♂, Huancabamba, Pasco, Peru, 6000–10000 ft., Boettger leg., ex coll. Hamilton Druce, 1919, ex coll. J. J. Joicey Coll. B. M. 1925-451, ex col. British Museum Natural History, OM 36.091 (specimen identified as *Pyrrhopyge sergius sergius* by Evans) (OM).

The last two specimens from Peru were from the duplicate collection of NHMUK, seen by Evans (1951) and presented by R. I. Vane-Wright to OM in 1976.



Text-Fig. 25. *Pyrrhopyge jola* **sp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (paratype OM 76.570).

Pyrrhopyge caribe Orellana, [2010]

Two subspecies.

DISTRIBUTION – Eastern Colombia and Venezuela.

Pyrrhopyge caribe caribe Orellana, [2010]

Pls. 28–29, Figs. 21 (holotype ♂), 22 (paratype ♀) (d, v)

DISTRIBUTION – Eastern Colombia to Venezuela.

Colombia: Meta (*Villavicencio*). **Venezuela:** Aragua (*Choroní, El Castaño – near Maracay, Maracay – El Limón and El Castaño, Pao de Zárata, Pozo del Diablo – near Maracay, Quebrada Yuma*); Bolívar (*between Parupa and Kavanayén*); Carabobo (*Bejuma, Cuesta Yuma*); Distrito Capital (*Caracas, El Valle*); Lara (*Barquisimeto, San Pedro*); Táchira (*Río Doradas*); Trujillo (*Campo Elias, La Gira de Betijoque, Rafael Rangel*); Vargas (*Via Oritopo – Osma*).

TYPE:

Pyrrhopyge caribe caribe Orellana, [2010]. Holotype male from El Castaño, cr. Maracay, Aragua, Venezuela, 500 m, 13-XI-1967, W. Szumkowski leg. MIZA.

Pyrrhopyge caribe camacho Orellana, [2010]

Pls. 28–29, Figs. 23 (holotype ♂) (d, v)

DISTRIBUTION – Southern Venezuela.

Venezuela: Bolívar (*between Santa Elena de Uairén and Ikabarú*).

REMARKS – Only known from the holotype.

TYPE:

Pyrrhopyge caribe camacho Orellana, [2010]. Holotype male with the following labels: / Venezuela Bolivar entre Sta Elena de Uairén – Ikabarú 1000 m 3 Sep. 2000 A. Orellana & R. Hesterberg/ Prep Genit Hesp 00032/ AO092 foto/. The original description mentions the altitude as 850 m and the collector only A. Orellana. (AO).

Pyrrhopyge eduardo* O. Mielke, Brockmann & C. Mielke, *sp. n.

Pls. 28–29, Figs. 24 (holotype ♂) (d, v); Text-Fig. 26 (♂ genitalia)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: La Paz (*Caranavi area*).

DESCRIPTION – Male: “*phidias* form”. Forewing length 26 mm. Ground colour black with some bluish iridescence. Head, palpi, cheeks, collar and anal tuft red. Fringes white, at the forewing until the apex and at the hindwing from Sc+R₁ until anal angle. Forewing with outer margin not excavated in CuA₂–2A. Genitalia very similar to that of *Pyrrhopyge sarpedon*, but differs by the shorter distal process and the larger dorsal process of the valva. The wide apophyses of tegumen and the uncus narrow and bifid, U-shaped are characteristic. Only one male is known.

Female: unknown.

TAXONOMY – *Pyrrhopyge eduardo* *sp. n.* shows a similar male genitalia of that of *P. sarpedon*, however the former doesn't bear the white semi-hyaline band on the forewing, while is present in the latter. Both species are allopatric with a well distinct geographical distribution.

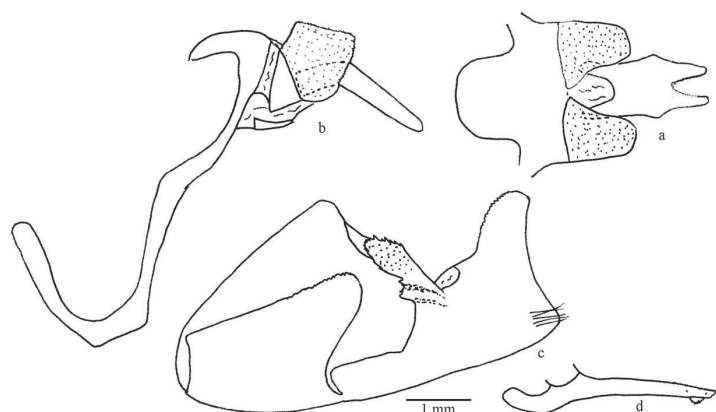
ETYMOLOGY – The specific name is in honour of Eduardo Jorge Costa Mielke, the second son of Olaf H. H. Mielke.

REMARKS – Only known from the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ X-XI-2002 Caranavi area La Paz, Bolivia 800 – 1700 m. C. Tello leg./ gen[italia] prep[arada]., [O.] Mielke 2014/ Holotypus *Pyrrhopyge eduardo* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 61.203/. OM-DZUP.

Paratypes: 1 ♂, Estrella, 1300 m, Caranavi, Bolivia, 23-X-2007, J. Uehara leg. (210174; CJU); 1 ♂, Estrella 1300–1500 m, Caranavi, Bolivia, 26-X-2007, J. Uehara leg. (CJU)



Text-Fig. 26. *Pyrrhopyge eduardo* *sp. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

***Pyrrhopyge steyermarki* Orellana, Costa & Grishin, 2021**

Pl. 76, Figs. 12 (holotype ♂) (d, v)

DISTRIBUTION – Southern Venezuela.

Venezuela: Bolívar (*Auyán Tepui – Guayraca–El Danto*).

REMARKS – Only known from the holotype.

TYPE:

Pyrrhopyge steyermarki Orellana, Costa & Grishin in Costa *et. al.*, 2021. Holotype male with the following labels: / Venezuela, Bolívar, Auyán Tepui, Guayraca–El Danto, 1.400 m, 04-IV-2015, M. Costa leg./ Auyntepui, entre [between] Guayraca y [and] El Danto, 1400 m, 4-4-2015/ *Pyrrhopyge steyermarki*, Holotipo ♂, Orellana, Costa & Grishin des. 2021/. Genit. prep. AO 2018-001; DNA sample NVG-19047E03. MIZA.

***Pyrrhopyge charybdis* Westwood, 1852**

Two subspecies.

DISTRIBUTION – Central to southern Brazil, northeastern Argentina and Uruguay.

***Pyrrhopyge charybdis charybdis* Westwood, 1852**

Pls. 30–31, Figs. 1 (♂), 2 (♀) (d, v)

DISTRIBUTION – Central to southern Brazil, northeastern Argentina and Uruguay.

Brazil: Distrito Federal (*Brasília*); Bahia (*Rio de Contas – Pico das Almas*); Minas Gerais (*Belo Horizonte, Carmo do Rio Claro, Catas Altas – Caraça, Conceição dos Ouros, Diamantina, Itajubá, Itamonte, Passa Quatro, Poços de Caldas, Virginia*); Espírito Santo (*Santa Cruz, Santa Teresa*); Rio de Janeiro (*Itatiaia, Nova Friburgo*); São Paulo (*Campos do Jordão, Delfim Moreira, São Bento do Sapucaí, São José do Barreiro – Bocaina, São Paulo*); Paraná (*Candói, Castro, Curitiba, Faxinal do Céu, Foz do Iguaçu, Guarapuava, Inácio Martins, Lapa, Mangueirinha, Palmas, Palmeira, Palotina, Piraquara, Tibagi, Turvo*); Santa Catarina (*Bom Jardim da Serra, Fraiburgo, Joaçaba, Mafra, Monte Castelo, Santa Cecília, Curitiba, São Bento do Sul – Rio Vermelho, São Joaquim, Urubici*); Rio Grande do Sul (*Bom Jesus, Canela, Guarani, Osório, Pareci Novo, Pelotas, Porto Alegre, Santa Maria, São Francisco de Paula, São José dos Ausentes*). **Uruguay:** Durazno (*La Paloma*); Rivera (*Cerro Chato*); Tacuarembó. **Argentina:** Buenos Aires (*Isla Martín García*); Corrientes; Entre Ríos; Formosa; Misiones.

The citations **Colombia:** Cesar (*Manauare*) (Godman & Salvin 1880), **Bolivia:** La Paz (*Chulumani, Coroico*) and Cochabamba (Weeks 1901, 1905), **Colombia, Venezuela and Trinidad and Tobago:** Port of Spain (Kaye 1904, 1921), **Venezuela** (Longstaff 1908), **Venezuela:** Bolívar (*Suapure*) (Weeks 1911), **Venezuela:** Distrito Capital (*Caracas*), **Belize:** Western District (Davis 1928), **Brazil:** Amazonas (*Manaus*) (Bell 1931), **Colombia** (Hayward 1947, 1948), **Venezuela:** Distrito Capital (*Caracas*) (Forster 1949), **Brazil:** Pernambuco (Evans 1951), **Colombia:** east Andes (Sturm 1958), **Brazil:** Paraíba (Cavalcante 1983), **Colombia:** Meta (*Villavicencio*) (Sturm 2001), **Suriname** (de Jong 1983), and **Brazil:** Mato Grosso (Palo Jr. 2017) are incorrect.

TYPE:

Pyrrhopyga [*sic*] *charybdis* Westwood, 1852. The original description mentions an unspecified number of specimens without data in NHMUK [Brazil, Minas Gerais – on the label of the lectotype]. To ensure the correct identification of *Pyrrhopyga charybdis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Type – *charybdis* D. W. H./ Minas Geraes [Brazil] Hewitson Coll. 79-69. *Pyrrhopyga charybdis*. 2./ This specimen from Minas Geraes ex Hew. Coll. Is in my opinion [??] the TYPE of *charybdis* W. D. H. It agrees [??] with the figure. N.D. Riley 14/X/30/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga charybdis* Westwood, 1852 O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge charybdis semita* Evans, 1951**

Pls. 30–31, Figs. 3 (♂), 4 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Rio de Janeiro (*Angra dos Reis, Cabo Frio, Petrópolis, Rio de Janeiro, Teresópolis*); São Paulo (*Cananéia – Ilha do Cardoso, Guarujá, Santos, São Bernardo, São Paulo – Alto da Serra e Ipiranga*); Paraná (*Morretes – Alto da Serra 800 m, Pontal do Paraná*); Santa Catarina (*Blumenau, Corupá, Dalbérgia, Imbituba, Joinville, Massaranduba, São Bento do Sul – Rio Natal, São Francisco do Sul*).

The citation **Bolivia** (Palo Jr. 2017) is incorrect.

REMARKS – Inhabitant of lowlands along the coast.

TYPE:

Pyrrhopyge charybdis semita Evans, 1951. Holotype male with the following labels: / Type – *semita* Evans/ Theresopolis [Rio de Janeiro State, Brazil]/. NHMUK.

***Pyrrhopyge cressoni* Bell, 1932**

Pls. 30–31, Figs. 5–6 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia.

Ecuador: Pastaza (*Mera*). **Peru:** Cusco (*Cosñipata Valley* – *Quebrada Quitacalzón 1,100 m, Quincemil, Saringabeni*); Huánuco (*Tingo Maria*); Loreto (*Río Putumayo*); Madre de Dios (*Tambopata Reserve*); Puno (*Parque Nacional Bahuaia Sonene*); San Martín (*Achinamiza, Juanjui*). **Bolivia:** La Paz (*Caranavi, Madidi*); Santa Cruz (*Buenavista*).

TYPE:

Pyrrhopyge cressoni Bell, 1932. Holotype male with the following labels: / Buenavista 75 Km. N. W. of Santa Cruz, Boliva [*sic*] 450 m. alt. 1926–27 (Jose Steinbach)/ Slide Phil. 30/ Genitalia slide No. 1978/ Type *Pyrrhopyge cressoni* E. L. Bell 7753./ *Pyrrhopyge cressoni* Bell Holotype ♂ / ANSP, transferred to CMP.

***Pyrrhopyge amyclas* (Cramer, 1779)**

Two subspecies.

DISTRIBUTION – Colombia to northern Peru and to French Guiana, Trinidad and Tobago and northern Brazil.

***Pyrrhopyge amyclas amyclas* (Cramer, 1779)**

Pls. 30–31, Figs. 9 (neotype ♂ of *Papilio amyclas* and neotype of *Tamyris laonome*), 13 (♂), 14 (♀) (d, v)

Syn.: *Hesperia amiatus* Fabricius, 1793

Tamyris laonome Swainson, 1821

DISTRIBUTION – Colombia to French Guiana and Trinidad and Tobago.

Colombia. Venezuela. Trinidad and Tobago: Diego Martin (*Bellevue*). **Guyana:** Demerara-Mahaica (*Demerara River, Georgetown*). **Suriname:** Coronie; Marovijne (*Anapaike, Galibi*); Nickerie (*Wageningen*); Para (*Gelderland*); Paramaribo (*Beekhuizen, Nieuw Amsterdam, Zorg em Hoop*); Sipaliwini; Wanica (*Lelydorp*). **French Guiana:** Cayenne (*Cayenne, Farnous, Macouria – La Marguerite, Tonate*).

The citations **Peru** and **Bolivia** (Bell 1946); **Brazil:** Rio de Janeiro (Prittwitz 1868) and **Bolivia:** Santa Cruz (*Santa Cruz de la Sierra*) (Burmeister 1878) are probably incorrect.

TYPES:

Papilio amyclas Cramer, 1779. The original description mentions an unspecified number of specimens from Suriname. No syntypes were found in NBC or NHMUK, where many of Cramer's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Papilio amyclas*, mentioned as *Pyrrhopyge amyclas amyclas* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Paramaribo [Suriname] July, 1892/ Rothschild Bequest B.M.1939-1./ ex coll. Brit. Mus. Nat. Hist./ Neotypus *Papilio amyclas* Cramer, 1779 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 36.164/. OM-DZUP. Specimen from the duplicate collection of NHMUK, seen by Evans (1951) and presented by R. I. Vane-Wright to OM in 1976.

Hesperia amiatus Fabricius, 1793. The original description mentions an unspecified number of specimens from America in collection Drury. No syntypes were found in the above mentioned collections, but one is in the Fabricius collection in ZMUC. To ensure the correct identification, this syntype male (without abdomen) is **herein designated as the lectotype**; this specimen with the following label: / *amiatus*!; two other labels will be added: / Lectotypus/ Lectotypus *Hesperia amiatus* Fabricius, 1793, O. Mielke, Brockmann & C. Mielke det. 2019/. ZMUC.

Tamyris laonome Swainson, 1821. The original description mentions the holotype (one specimen mentioned) female from South America. No syntypes were found in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure

the correct identification of *Tamyris laonome*, mentioned as a synonym of *Pyrrhopyge amyclas amyclas* by Evans (1951), a male is **herein designated as the neotype**; the specimen is the same as the neotype of *Papilio amyclas* and the following additional label has been added: / Neotypus *Tamyris laonome* Swainson, 1821 O. Mielke, Brockmann & C. Mielke det. 2015/. OM-DZUP.

***Pyrrhopyge amyclas denticulata* Herrich-Schäffer, 1869**

Pls. 30–31, Figs. 12 (neotype ♂), 15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to northern Peru and northern Brazil.

Ecuador: Napo (*Río Napo*). **Peru:** Loreto (*Allpahuayo Mishana, Iquitos, Lower Río Ucayali, Pebas, Quebrada Polis*). **Brazil:** Roraima (*Alto Alegre – Ilha de Maracá*); Amazonas (*Barcelos, Humaitá, Januareté, Manaus, Maués, Parintins, Rio Jurua, Rio Negro, São Paulo de Olivença, Tefé*); Pará (*Itaituba, Juruti, Monte Cristo, Óbidos, Rio Arapiuns, Santarém*); Rondônia (*Porto Velho*).

The citations **Brazil** (São Paulo) and **Bolivia** (La Paz) (Palo Jr. 2017) are incorrect.

TYPE:

Pyrrhopyga [*sic*] *denticulata* Herrich-Schäffer, 1869. The original description mentions an unspecified number of specimens from Tropical America. No syntypes were found in MfN or ANSP, where the Herrich-Schäffer's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Pyrrhopyga denticulata*, mentioned as *Pyrrhopyge amyclas denticulata* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Ilha de Maracá Alto Alegre, RR [Roraima, Brazil] 24-31/VIII/1987 [O.] Mielke & Casagrande [leg.]/ *Pyrrhopyge amyclas denticulata* H.-Sch., 1869 O. Mielke det. 1989/ Neotypus *Pyrrhopyga denticulata* Herrich-Schäffer, 1869 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.255/. DZUP.

***Pyrrhopyge erazoe* Orellana, [2010]**

Pls. 30–31, Figs. 10 (♂), 11 (holotype ♀) (d, v)

DISTRIBUTION – Eastern Ecuador to southern Venezuela.

Venezuela: Bolívar (*Auyán Tepui*). **Ecuador:** Morona-Santiago (*Macas, Palora – Parque Nacional Sangay*); Napo (*Archidona*).

TYPE:

Pyrrhopyge erazoe Orellana, [2010]. Holotype female from Guayaraca, Auyán Tepui, Bolívar, Venezuela, 1100 m, 19-IV-1956, F. Fernández-Y. & C. J. Rosales leg. MIZA.

***Pyrrhopyge phylleia* Hewitson, 1874**

Pls. 30–31, Figs. 17 (♂), 18 (♀), 21 (lectotype ♂ of *Pyrrhopyge punctata*) (d, v); Text-Fig. 27 (♂ genitalia)

Syn.: *Pyrrhopyge hades* Mabille, 1903

Pyrrhopyge fleximargo Mabille & Boulet, 1908

Pyrrhopyge punctata Röber, 1925, **syn. n.**

Pyrrhopyge phylleia delos Evans, 1951, **syn. n.**

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Puno (*Río Inambari to Limbani, Sandia, San Juan del Oro*). **Bolivia:** Beni; Cochabamba (*Chapare, Cristal Mayu*); La Paz (*Caranavi, Chulumani, Coroico, Farinas, Irupana, Mapiri, Río Zongo, Valle del Zongo – Huaylipaya*); Santa Cruz (*Estrella*).

The citations **Ecuador:** Loja (*Loja*) (Campos 1927, Williams & Hayward 1944) are probably incorrect.

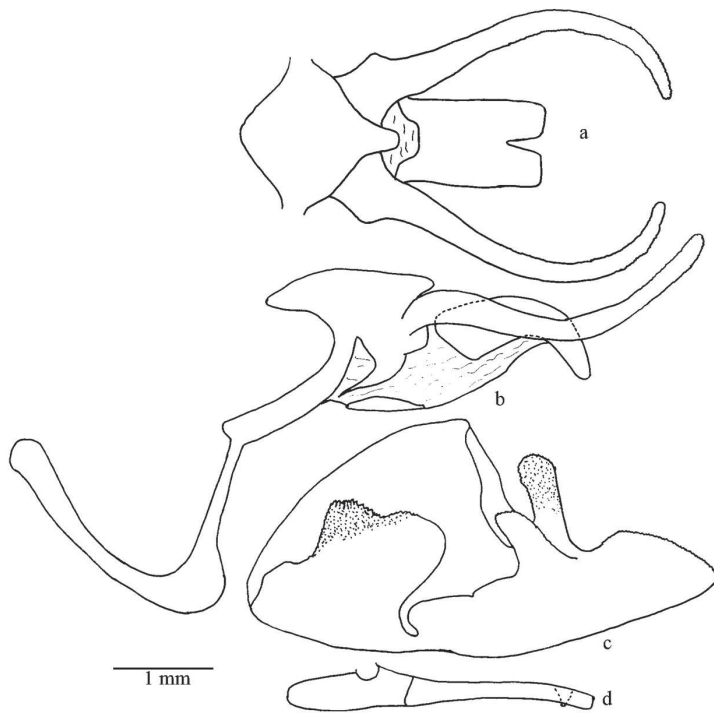
TAXONOMY – *Pyrrhopyge punctata* is a variation with an orange spot at the end of the discal cell on the ventral hindwing; also there are no red spots on the patagia, as mentioned by Evans (1951) for *P. phylleia delos*; the genitalia (Fig. 29) of the lectotype was dissected and

confirms the new synonymy. Specimens with patagia entirely black and with patagia more or less well-developed spot on either side, the only difference given by Evans (1951), are sympatric (Bolivia: La Paz – Chulumani); so *P. phylleia delos* is considered a new synonym of *P. phylleia*. For other taxonomic comments, see under *P. sergius*.

Male genitalia similar to those of *P. sergius sergius*, but the apophyses of the tegumen in *P. phylleia* normally well curved evenly upper the uncus and more curved inwards distally; the distal part of the harpe always triangular and wide. Female genitalia of both species don't differ.

The DNA barcode minimum divergences are: between *P. phylleia* (n= 15) and *P. sergius sergius* (n= 5) – 2.60%, between *P. phylleia* and *P. mina mina stat. rest.* (n= 7) – 2.75%, and between *P. phylleia* and *P. michael sp. n.* (n= 24) – 1.99%.

Pyrrhopyge phylleia is distinguished in this species group (“*Pyrrhopyge amyclas* group”, Evans 1951) only by the male genitalia from *P. arax* and *P. caranavi sp. n.*



Text-Fig. 27. *Pyrrhopyge phylleia*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (lectotype of *Pyrrhopyge punctata*).

TYPES:

Pyrrhopyga [sic] phylleia Hewitson, 1874. The original description mentions an unspecified number of specimens from Bolivia, Buckley leg. To ensure the correct identification of *Pyrrhopyga phylleia*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Type/ Bolivia, Hewitson Coll. 79-69. *Pyrrhopyga phylleia*. 4./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga phylleia* Hewitson, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge hades Mabilie, 1903. The original description mentions an unspecified number of specimens from Bolivia. To ensure the correct identification of *Pyrrhopyge hades*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T / *hades* Stgr. Boliv[ia]./ *Hades* Stgr. Bolivia/ Ex musaeo P. Mabilie 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge hades* Mabilie, 1903, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge fleximargo Mabilie & Boulet, 1908. Holotype (one specimen is mentioned) male with the following labels: / Type/ Pérou meridional 1906 Herm. Rolle Coll. Boulet Museu Paris/. MNHN. The original description mentions Bolivia as type locality, but the label of the holotype says “Pérou meridional” as provenance, this specimen

is in the drawer under a label “*Pyrrhopyge fleximargo*”. The original description agrees with the specimen.

Pyrrhopyge punctata Röber, 1925. The original description mentions an unspecified number of specimens from Río Zongo, [La Paz], Bolivia, 750 m, A. H. Fassl leg. To ensure the correct identification of *Pyrrhopyge punctata*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Río Zongo [sic] [La Paz] Bolivia 750 m Coll. Fassl/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge punctata* Röber, 1925. O. Mielke, Brockmann & C. Mielke det. 2019/. ZfB. **Syn. n.**

Pyrrhopyge phylleia delos Evans, 1951. Holotype male with the following labels: / Type – *delos* Evans/ R. Inambari to Limbani, [Puno] S. E. Peru, III.04. (G. Ockenden)/ Rothschild Bequest B.M.1939-1./ NHMUK. **Syn. n.**

Pyrrhopyge haemon Godman & Salvin, 1893

Pls. 30–31, Figs. 22, 24 (♂), 23 (♀) (d, v)

DISTRIBUTION – Costa Rica to Panama.

Costa Rica: Cartago (*Alto Belén, Orosi, Parque Nacional Tapantí*); Guanacaste (*San Jerónimo*), Heredia (*San Isidro*); San José (*Bajo La Hondura*); Province? (*San Germain*). **Panama:** Chiriquí (*Fortuna Dam*); Veraguas.

TYPE:

Pyrrhopyge haemon Godman & Salvin, 1893. Holotype (one specimen is mentioned) female with the following labels: / Typus/ Costarica 89 [??]/ *haemon* G. u. S. Type/ Zool. Mus. Berlin/. MfN.

Pyrrhopyge arax Evans, 1951

Pls. 32–33, Figs. 1 (♂), 2 (♀) (d, v)

DISTRIBUTION – Northern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley – Chontachaca 950 m to San Pedro 1,400 m*), Marcapata, Quillabamba; Junín (*Río Satipo, Shimá*); Loreto (*Iquitos*); Puno (*Carabaya, Limbani, Oroya – Río Inambari, Yahuar Mayo*). **Bolivia:** Cochabamba (*Chapare*); La Paz (*Caranavi, Chulumani, Puente Villa, San Carlos*).

TYPE:

Pyrrhopyge arax Evans, 1951. Holotype male with the following labels: / Type – *arax* Evans/ Bolivie. ex Germain./ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Pyrrhopyge pelota Plötz, 1879

Pls. 32–33, Figs. 3 (♂), 4 (♀) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia, northern (Rondônia) to southern Brazil, Paraguay and northeastern Argentina.

Peru: Madre de Dios (*Pampas del Heath*). **Bolivia:** Beni (*San Borja*); Santa Cruz. **Brazil:** Rondônia (*Vilhena*); Mato Grosso (*Pantanal*); Goiás (*Leopoldo Bulhões, Vianópolis*); Distrito Federal (*Brasília*); Minas Gerais (*Araguari, Belo Horizonte, Carmo do Rio Claro, Conceição d'Aparecida, Conceição dos Ouros, Diamantina, Lagoa Santa, São Roque de Minas, Sete Lagoas, Uberaba, Uberlândia*); Rio de Janeiro; São Paulo (*Campinas, Moji Guaçu, Rio Claro, Santana do Parnaíba, Santos, São Carlos*); Paraná (*Castro, Jaguariaíva, Palotina, Ponta Grossa – Vila Velha, Tibagi*); Santa Catarina (*São Bento do Sul – Rio Vermelho*); Rio Grande do Sul (*Southeast, Southwest*). **Paraguay:** Central; Cordillera (*Piribebuy – Salto Pirareta*); Guairá (*Villarica*); Paraguari (*Sapucaí*). **Argentina:** Chaco (*Fontana*); Corrientes (*Bella Vista, Goya, Ituzaingó, Paso de los Libres, San Cosma*); Entre Ríos (*Liebig*); Formosa; Misiones (*Puerto Iguazú*).

The citations **Colombia** (Hayward 1947, 1948, Saini *et al.* 1986) are incorrect.

TYPE:

Pyrrhopyga [sic] pelota Plötz, 1879. The original description mentions an unspecified number of specimens from Brazil in Mus.

Berol. [MfN]. To ensure the correct identification of *Pyrrhopyge pelota*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Holotypus/ Brasil. Sello./ *Pelota* Plötz*/ 4900/ *Pelota* Plötz Stett. Ent. Zeit. 1879. p. 535/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge pelota* Plötz, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyge caranavi* O. Mielke, Brockmann & C. Mielke, *sp. n.

Pls. 32–33, Figs. 5 (holotype ♂) (d, v); Text-Fig. 28 (♂ genitalia)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: La Paz (*Caranavi*).

DESCRIPTION – Male: forewing length 25–26 mm. A species morphologically very similar to *Pyrrhopyge phylleia* and *P. arax*. Head, palpi, collar, patagia laterally, and anal tuft red, fine marginal area of both wings in both sides dark yellow, finer than in *P. phylleia* and *P. arax*, on dorsal and ventral hindwing, and fringes dark yellow, until the apex on forewing. Apophyses of the tegumen long, reaching the end of the uncus, thick, angled and spined at distal part, uncus divided by a very narrow slit, valva with a short and rounded harpe.

Female: unknown.

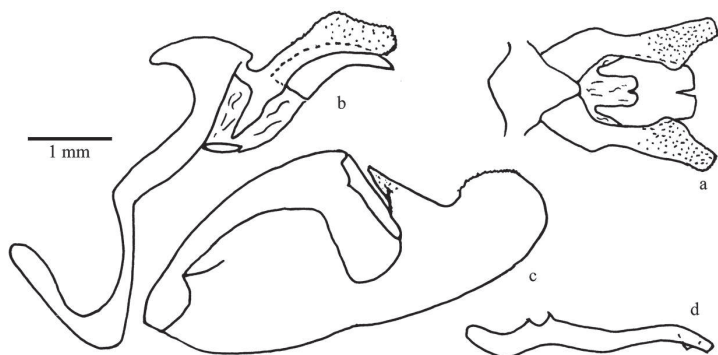
TAXONOMY – A species differing from *P. phylleia* and *P. arax* by the shorter and thicker apophyses of the tegumen, the different shapes of the uncus and of the valva.

ETYMOLOGY – The specific name refers to Caranavi, a city in La Paz province, Bolivia, where the two known specimens were collected.

TYPES:

Holotype male with the following labels: / Holotypus/ Caranavi, Bolivia 600-1000 m, IX-X-1988. Tello leg./ Holotypus *Pyrrhopyge caranavi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2018/ gen[italia]. prep[arada]., [O.] Mielke 2018/ 19.870/. OM-DZUP.

Paratype: 1 ♂, same data as the holotype, 19.891 (OM).



Text-Fig. 28. *Pyrrhopyge caranavi* *sp. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

***Pyrrhopyge papius* Hopffer, 1874**

Two subspecies.

DISTRIBUTION – Northwestern, western, central and eastern Colombia to central-eastern Bolivia and northwestern Venezuela.

***Pyrrhopyge papius papius* Hopffer, 1874**

Pls. 32–33, Figs. 6 (♂), 7 (♀) (d, v)

DISTRIBUTION – Northwestern, western, central and eastern Colombia to central-eastern Bolivia and northwestern Venezuela.

Colombia: Antioquia (*Bello*, *Frontino*, *Rionegro*); Boyacá (*Arcabuco*, *Muzo*); Caquetá (*Sucre*); Cauca (*Popayán*, *Suárez*);

Cundinamarca (*Alban*, *La Aguadita*, *Chirijara to Guayabetal*, *Choachi*, *Monterredondo*); Meta (*Río Negro*); Tolima (*Cañon del Tolima*, *Honda*, *La Victoria*); Valle del Cauca (*Cali*, *Calima*, *Lomitan*, *Río Aguacatal*, *Villa Carmelo*). **Ecuador:** Cotopaxi (*Angamarca*); Loja (*Loja*, *San Francisco*); Morona-Santiago (*Huamboya*, *San Isidro – Parque Nacional Sangay*); Napo (*Baeza*, *Cotundo*, *Puerto Misahualli*, *Río Jondachi*, *Talac*, *Tena*, *Yanayacu Biological Station*); Pastaza (*Abitagua*, *Mera*, *Puyo*, *Río Pastaza*); Tungurahua (*Ambato*, *Baños*, *El Rosario*, *El Topo*, *Hacienda La Mascota*, *Hacienda Santa Inés*, *Mapoto*, *Río San Francisco*, *Río Topo*, *Topo*, *Yunguilla*); Zamora-Chinchipe (*Río Numbala*, *Zamora*). **Peru:** Amazonas (*Chachapoyas – Yambasbamba*, *Oso Perdido 2,200 m*, *Río Nieva 1,500 m to Alto Nieva 1,930 m*, *Rodríguez de Mendoza*, *Santa Maria de Nieva*); Cajamarca (*Río Tabaconas*); Cusco (*Aguas Calientes*, *Machu Picchu*); Huancavelica (*Pomacochas 2,000 m*); Huánuco (*Tingo Maria*); Junín (*Andamarca*, *Chanchamayo*, *La Merced*, *Río Colorado*, *San Ramón*, *Satipo*); Pasco (*Huancabamba*, *Pichis Road*, *Pozuzo*, *Tambo Eneñas*); San Martín (*Juanjui*, *Moyobamba*, *Rioja – San Agostin 1,000 m to Jorge Chávez 1,400 m*, *Río Serranoyacu*, *Santa Cruz*, *Santa Rosa del Mirador*, *Tarapoto*). **Bolivia:** Beni (*Moxos*). **Venezuela:** Táchira (*El Reposo*, *Parque Nacional Tamá*, *Via Chorro del Indio*, *Via Delicias*).

TYPE:

Pyrrhopyge papius Hopffer, 1874. The original description mentions two males from Neu-Granada [Colombia] and Moxos [Beni, Bolivia], Pavon leg. To ensure the correct identification of *Pyrrhopyge papius*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Lectotypus/ Moxos [Beni, Bolivia] Pavon/ 4898/ *Papius* Hpfr*/ 1:9./; another label will be added: / Lectotypus *Pyrrhopyge papius* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published.

***Pyrrhopyge papius pasca* Evans, 1951**

Pls. 32–33, Figs. 8 (♂) (d, v); Pl. 76, Figs. 8 (♀) (d, v)

DISTRIBUTION – Northwestern, western and central Colombia.

Colombia: Antioquia (*Cocorna*, *El Retiro*); Caldas (*Manizales*, *Pensilvania*); Quindío (*Circasia*); Risaralda (*Montezuma*, *Pereira*, *Pueblo Rico*); Valle del Cauca (*Darién – Lago Calima*).

TYPE:

Pyrrhopyge papius pasca Evans, 1951. Holotype male with the following labels: / Type – *pasca* Evans/ Colombia (Cauca) Distrito de Pereira [Risaralda] Roman M. Valencia 1886/ R. Oberthür Coll. Brit. Mus. 1931-136/ *papius*. (respectus) *pasca* Nov: ♂ Colombia (Cauca) (BM). 22/2 Colombia (Cauca) Indigo blue above: no thoracic side tufts./ NHMUK.

***Pyrrhopyge melanomerus* Mabilie & Boullet, 1908**

Two subspecies.

DISTRIBUTION – Eastern Ecuador to southeastern Peru.

***Pyrrhopyge melanomerus melanomerus* Mabilie & Boullet, 1908**

Pls. 32–33, Figs. 9 (neotype ♂) (d, v)

Syn.: *Pyrrhopyge tarapotoensis* Bell, 1931

DISTRIBUTION – Central-eastern to southeastern Peru. Likely in Bolivia.

Peru: Junín (*Angamarca*, *Chanchamayo*, *La Merced*, *Perené*, *Río Perené*, *San Ramón*, *Satipo*); Pasco (*Pozuzo*); Puno (*Carabaya*); San Martín (*Tarapoto*). **Bolivia** (?).

TYPES:

Pyrrhopyge melanomerus Mabilie & Boullet, 1908. Holotype (one specimen is mentioned) male from Bolivia in collection Dognin. The holotype was not found in MNHN or USNM, where parts of Dognin's collection were deposited, or even in IRSC, NHMUK or

MfN, where many of Mabilles types are deposited, or in any museum or collection mentioned above (see “Abbreviations of the collections”). The type locality is mentioned as Bolivia, but no specimens were found for this country. To ensure the correct identification of *Pyrrhopyge melanomerus*, mentioned as *Pyrrhopyge melanomerus* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ 13-X-1989 Rio Tulumayo 6 KM SE Vitoc, S[an]. Ramon, Junín, Peru, 1100m, [O.] Mielke & Casagrande leg./ *Pyrrhopyge melanomerus* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2015/ OM 23.374/. OM-DZUP.

Pyrrhopyge tarapotoensis Bell, 1931. Holotype male with the following labels: / Type/ Tarapoto, [San Martín], Peru/ 390/ G424/ *Pyrrhopyge tarapotoensis* Bell Holotype ♂/. AMNH.

Pyrrhopyge melanomerus patma Evans, 1951

Pls. 32–33, Figs. 10 (♂), 11 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Peru.

Ecuador: Loja (*Loja*); Napo (*Cotundo*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Huambo, Rodríguez de Mendoza*); Huánuco (*Tingo Maria*); Pasco (*Pozuzo, Río Palcazu*); San Martín (*Moyobamba, Rioja – San Agostin 1,000 m to El Afluente 1,300 m*).

TYPE:

Pyrrhopyge melanomerus patma Evans, 1951. Holotype male with the following labels: / Type – *patma* Evans/ Zamora, [Zamora-Chinchipe], Ecuador, 3–4000 ft. (O. T. Baron. / Rothschild Bequest B.M. 1939-1/. NHMUK.

Pyrrhopyge afluente O. Mielke, Brockmann & C. Mielke, *sp. n.*

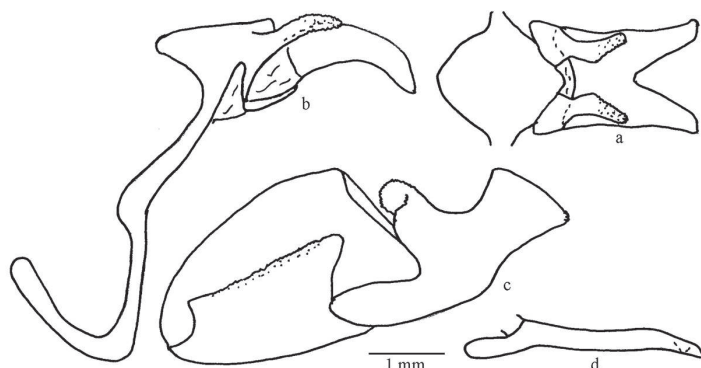
Pls. 32–33, Figs. 12 (holotype ♂) (d, v); Text-Fig. 29 (♂ genitalia)

DISTRIBUTION – Eastern Ecuador to Central-eastern Peru.

Ecuador: Pastaza (*Mera*). **Peru:** San Martín (*Rioja – El Afluente 1,300 m*).

DESCRIPTION – Forewing length 26 mm. Ground colour black. Patagia laterally, fore coxae with a few scales, tegulae, the lateral parts of dorsal metathorax, posteriorly to tegulae, and anal tuft red. Fringes white, on forewing until apex. Genitalia very characteristic; apophyses of tegumen angled and very short, not reaching the uncus division; uncus bifid, division broadly V-shaped, valva with a large harpe, broadening distally and the dorso-proximal process broad and short, until the middle of the ampulla. Only three males are known.

Female: unknown.



Text-Fig. 29. *Pyrrhopyge afluente* *sp. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

TAXONOMY – A species of the “*Pyrrhopyge papius* species group” (Evans 1951), only possible to identify by an examination of the male genitalia. The very short and angled apophyses of tegumen not reaching the uncus slit, the bifid uncus with a very broad division V-shaped and the very typical valva distinguish this species from all

others of this group.

ETYMOLOGY – The specific name refers to El Afluente, the type locality of the species.

TYPES:

Holotype male with the following labels: / Holotypus/ Peru, San Martín, Rioja, El Afluente, IX-2016, 5°39'48" S, 77°41'41" W, 1320 m, J. Pintado leg./ gen[italia]. prep[arada]., [O.] Mielke 2016/ Holotypus *Pyrrhopyge afluente* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 81.580/. OM-DZUP.

Paratypes: 1 ♂, same data as the holotype, OM 81.490 (OM); 1 ♂, IX-2017, El Afluente, Rioja, San Martín, Peru, 1300 m, J. Pintado leg. (MUSM); 2 ♂, IX-X-2012, Mera, 1200 m, Pastaza, Ecuador (Genitalia 160172, 160038; CJU).

Pyrrhopyge decipiens Mabilles, 1903

Pls. 32–33, Figs. 13–14 (♂) (d, v)

DISTRIBUTION – Northwestern, western and eastern Colombia to eastern Bolivia.

Colombia: Antioquia (*Medellín, Guarne*); Quindío (*La Concha, Vereda*); Putumayo. **Ecuador:** Morona-Santiago (*San Isidro – Parque Nacional Sangay*); Pastaza (*Abitagua, Mera*); Tungurahua (*Río Topo*). **Peru:** Amazonas (*Cordillera del Condor – Falso Paquisha, Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Buenos Aires 2,400 m*); Huánuco (*Río Huallaga, Tingo Maria*); Junín (*Chanchamayo, Perené, Río Perené, San Ramón, Satipo – Anapate Valley, Shimá*); Madre de Dios (*Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Huancabamba, Pozuzo*); Puno (*Carabaya, Chaquimayo, Yahuarmayo*); San Martín (*Rioja – San Agostin 1,000 m to El Afluente 1,300 m*). **Bolivia:**

TYPE:

Pyrrhopyge decipiens Mabilles, 1903. The original description mentions an unspecified number of specimens from Ecuador and Bolivia. To ensure the correct identification of *Pyrrhopyge decipiens*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Type Mabilles/ *decipiens* M. Bolivia/ *P. decipiens* Mab./ *Ponina* [??] Bol[ivia]./ Ex museo P. Mabilles 1925/ Ex Oberthür Coll. Brit. Mus. 1927-3./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge decipiens* Mabilles, 1903, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge frona Evans, 1951

Pls. 32–33, Figs. 15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia.

Ecuador: Morona-Santiago (*Palora – Parque Nacional Sangay*). **Peru:** Cusco (*Quincemil*); Huánuco (*Tingo Maria*); Junín (*Perené, Río Colorado, San Ramón, Satipo, Shimá*); Pasco (*Pozuzo*); San Martín (*Juanjuí, Rioja*). **Bolivia:** Cochabamba (*Carmen Pampa*); La Paz (*Caranavi*).

TYPE:

Pyrrhopyge frona Evans, 1951. Holotype male with the following labels: / Type – *frona* Evans/ Inca, Perene, [Junín], Peru, 4000 ft. 8-10. '02. H. Watkins/ Adams Bequest B.M. 1912-399/. NHMUK.

Pyrrhopyge placeta Evans, 1951

Pls. 32–33, Figs. 17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Southeastern Brazil (Minas Gerais and Espírito Santo).

Brazil: Minas Gerais (*Caxambu*); Espírito Santo (*Santa Leopoldina, Santa Teresa*).

TYPE:

Pyrrhopyge placeta Evans, 1951. Holotype female with the following labels: / Type – *placeta* Evans/ Espírito-Santo Brasil. Ex coll. Fruhstorfer/ Rothschild Bequest B.M. 1939-1/. NHMUK.

***Pyrrhopyge sanpedro* O. Mielke & Lamas, sp. n.**

Pls. 32–33, Figs. 19 (holotype ♂) (d, v); Text-Fig. 30 (♂ genitalia)

DISTRIBUTION – Southeastern Peru.

Peru: Cusco (*Cosñipata Valley – San Pedro 1,400 m*).

DESCRIPTION – Male: Forewing length 25–26 mm. Ground colour black. Fore coxae, patagia laterally, tegulae, and anal tuft red; a little red also on lateral parts of dorsal metathorax, posteriorly to tegulae. Forewing with a spot of few scarce dark yellow scales at the base of Cu_1 – Cu_2 , dorsally smaller than ventrally, and in the holotype also ventrally in the adjacent half of the discal cell. Fringes white, on the forewing from R_5 (missing above this vein) to 2A and on the hindwing a little longer, from $Sc+R_1$ to the centre of 2A–3A. Genitalia with uncus division as a narrow slit; apophyses of tegumen slender and shorter than uncus; harpe broad, with two dorso-distal lobes, the proximal thick and the distal upwards, distally slightly concave.

Female: unknown.

TAXONOMY: A species very similar in colouration to all species of the “*Pyrrhopyge papius* species group” (Evans 1951), but differing by the presence of a spot of dark yellow scales at the base of Cu_1 – Cu_2 on the ventral forewing. The male genitalia presents also the uncus with a narrow slit, as in *P. papius*, the only species of this group with such shape, but differs from this species by the shape of the valva. In *P. sanpedro* sp. n. the distal concavity is shallow, the disto-dorsal process is directed dorsally and the disto-proximal process is thick and blunt, while in *P. papius* the distal concavity is deep, the disto-dorsal process is directed distally and then upwards, and the dorso-proximal process is pointed (see figure in Evans 1951).

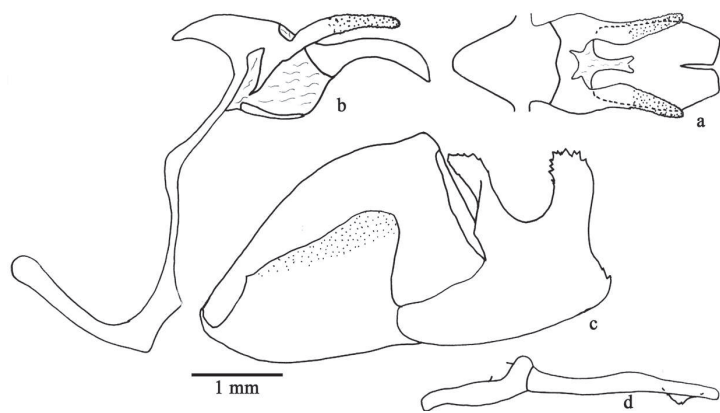
ETYMOLOGY – The specific name refers to the type locality, San Pedro, a small locality in the Cosñipata Valley, Cusco, Peru.

REMARKS – Only known from two specimens from the type locality.

TYPES:

Holotype male with the following labels: / Holotypus/ Peru. CU [Cusco]. San Pedro 1400m 1303/ 7133 3.II.2013 K. Kertell [leg.]/ Holotypus *Pyrrhopyge sanpedro* O. Mielke & Lamas. O. Mielke & Lamas det. 2017/. MUSM.

Paratype: 1 ♂, 10-II-2011, San Pedro, Cosñipata Valley, Cusco, Peru, 1400 m, 13°03' S, 71°33' W, S. Kinyon leg. (MUSM).



Text-Fig. 30. *Pyrrhopyge sanpedro* sp. n. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (paratype).

***Pyrrhopyge schausi* Bell, 1931**

Pls. 32–33, Figs. 20 (♂) (d, v)

DISTRIBUTION – Western, central and eastern Colombia to central-eastern Peru.

Colombia: Caldas (*Manizales*); Cauca; East Cordillera (*east side*).

Ecuador: Loja (*Loja*); Morona-Santiago (*Macas*); Pastaza (*Río Pastaza, San Jorge*); Pichincha (*Quito*); Tungurahua (*Ambato, Baños, Colonia Azuaya, El Rosario, El Topo*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Pongo*

del Rentema); Huancavelica (*Pomacochas 2,000 m*); San Martín (*Rioja – El Afluente 1,200 m to Jorge Chávez 1,450 m, Santa Cruz 1,200 m*).

TYPE:

Pyrrhopyge schausi Bell, 1931. Holotype male with the following labels: / Type/ Ecuador/ *Pyrrhopyge schausi* Bell Holotype ♂/. AMNH.

***Pyrrhopyge sadia* Evans, 1951**

Pls. 32–33, Figs. 21–23 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Peru.

Ecuador: Loja (*Loja*); Pastaza (*Sarayacu*); Pastaza – Tungurahua (*Baños to Puyo*); Zamora-Chinchipe (*Chicaña, Zamora*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Cordillera del Condor – Falso Pakisha, Rodríguez de Mendoza*); San Martín (*Rioja – Naranjos 1,000 m to Jorge Chávez 1,450 m, San Miguel de Shanusi*).

TYPE:

Pyrrhopyge sadia Evans, 1951. Holotype male with the following labels: / Type – *sadia* Evans/ Sarayacu [Pastaza] Eastern Side of Ecuador C. Buckley 1879/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Pyrrhopyge martena* Hewitson, [1869]**

Pls. 32–33, Figs. 24 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Peru.

Ecuador: Chimborazo (*Riobamba*); Loja (*Loja, San Francisco*); Morona-Santiago (*El Retiro – Parque Nacional Sangay, Gualaquiza, Macas*); Pastaza (*Mera, Pastaza, San Jorge, Sarayacu*); Tungurahua (*Ambato, Baños, El Topo, Hacienda Santa Inés, Río San Francisco, Río Topo*); Zamora-Chinchipe (*Zamora*). **Peru:** San Martín (*Rioja*).

TYPE:

Pyrrhopyga [*sic*] *martena* Hewitson, [1869]. The original description mentions an unspecified number of males from Ecuador in collection Hewitson. To ensure the correct identification of *Pyrrhopyga martena*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Ecuador. Hewitson Coll. 79-69. *Pyrrhopyga martena*. 4./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga martena* Hewitson, [1869], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge badassa* Hewitson, 1866**

Pls. 34–35, Figs. 1 (♂), 2 (♀) (d, v)

Syn.: *Pyrrhopyge rubricor* Mabilbe & Boulet, 1908

DISTRIBUTION – Western Colombia to western Ecuador, central and eastern Colombia to central-eastern Peru.

Colombia: Boyacá (*Muzo*); Caquetá (*road Altamira – Florencia, Sucre*); Cauca (*Popayán*); Cundinamarca (*Bogotá*); Meta (*Upper Río Negro, Villavicencio*). **Ecuador:** Cotopaxi (*Angamarca*); Loja (*Loja*); Morona-Santiago (*El Retiro*); Pastaza (*Canelos, Mera, Río Pastaza*); Tungurahua (*Ambato, Baños, El Topo, Hacienda La Mascota, Río San Francisco, Río Topo, Yunguilla*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Pomacochas*); Cajamarca (*Charapi, Chinchipe, Manchara*); Junín (*La Merced*); Loreto (*Nauta*).

The citation **French Guiana** (Bell 1931) is probably incorrect.

TAXONOMY – Evans (1951) characterized *Pyrrhopyge badassa* with five subspecies, even considering that the sympatry of *P. h. badassa* and *P. h. pseudobadassa* and the latter with *P. h. halma* (La Merced, Junín and Carabaya, Puno), respectively in Peru. In addition to the chromatic aspects, he also mentioned some differences in the genitalia, however it shows a high degree of variability making them diagnostically less helpful, with the exception of *P. h. hanga* in which the apophyses of the tegumen are larger than in the other taxa and the distal point of the harpe is more blunt. Among the specimens examined there is also a geographic overlap between *P. h. hanga* and *P. h. halma* in San Pedro, Cosñipata Valley, Cusco, Peru, and between *P.*

h. hanga, *P. h. henna* (a new synonym of *Pyrrhopyge olivacea* **stat. rev.**) and *P. h. pseudohadassa* in Previsto, Ucayali, Peru (about 35 km east of Tingo Maria) and in Tingo Maria, Huánuco, Peru. The only allopatric taxon is *P. h. hadassa*.

The analysis of the DNA barcode of the five subspecies, indicates large distances between the taxa, which together with their sympatry justifies the change of the status of all subspecies to species. The minimum divergences are: between *P. hadassa* (n= 2) and *P. halma* **stat. n.** (n= 2) – 6.7%, between *P. hadassa* and *P. hanga* **stat. n.** (n= 2) – 6.1%, between *P. hadassa* and *P. olivacea* **stat. rev.**, (n= 4) – 1.4%, between *P. hadassa* and *P. pseudohadassa* **stat. rest.** (n= 2) – 5.8%; between *P. halma* **stat. n.**, and *P. hanga* **stat. n.** – 4.3%, between *P. halma* **stat. n.** and *P. olivacea* **stat. rev.** – 6.7%, between *P. halma* **stat. n.**, and *P. pseudohadassa* **stat. rest.** – 1.8%; between *P. hanga* **stat. n.**, and *P. olivacea* **stat. rev.** – 6.7%, between *P. hanga*, **stat. n.**, and *P. pseudohadassa* **stat. rest.** – 4.0%, and between *P. olivacea* **stat. rev.**, and *P. pseudohadassa* **stat. rest.** – 6.4%.

TYPES:

Pyrrhopyga [**sic**] *hadassa* Hewitson, 1866. The original description mentions an unspecified number of specimens in the collection Hewitson, without data [Ecuador – on the label of the lectotype]. To ensure the correct identification of *Pyrrhopyga hadassa*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Ecuador Hewitson Coll. 79-69. *Pyrrhopyga hadassa* 4/ Type H T /; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga* [**sic**] *hadassa* Hewitson, 1866. O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge rubricor Mabilie & Boulet, 1908. The original description mentions four [?] males from Ecuador and Bolivia in MNHN (collection Boulet) and collection Mabilie. To ensure the correct identification of *Pyrrhopyge rubricor*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Env. Loja Equateur 1890 Donckier Coll. Boulet Museum Paris/ *Pyrrhopyge rubricor* Mab. & Boull., Type. O. Mielke det. 1996/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge rubricor* Mabilie & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Pyrrhopyge halma Evans, 1951, **stat. n.**

Pls. 34–35, Figs. 3 (♂) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Morro Leguia 2,100 m*); Puno (*Carabaya, Oroya – Río Inambari, Sandia, San Gaban, Yahuar Mayo*). **Bolivia:** Cochabamba (*Charapaya, Cochabamba, Cristal Mayu, El Limbo, San Jacinto, Yungas del Espíritu Santo*); La Paz (*Caranavi, Chaco, Chulumani, Farinas, Parque Nacional Madidi 1,500–1,800 m, Río Sarampiuni, Río Zongo, Yungas*).

TAXONOMY – This species belongs to the “*Pyrrhopyge hadassa* species group” (Evans 1951). The new status is confirmed; see comments under *Pyrrhopyge hadassa*.

TYPE:

Pyrrhopyge hadassa halma Evans, 1951. Holotype male with the following labels: / Type – *halma* Evans/ Bolivia Yungas & La Paz 1000 m ex H. Rolle 1902/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Pyrrhopyge hanga Evans, 1951, **stat. n.**

Pls. 34–35, Figs. 4 (♂) (d, v)

DISTRIBUTION – Central-eastern to southeastern Peru.

Peru: Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to Quebrada Santa Isabel 1,200 m*); Junín (*El Porvenir, Satipo*); Pasco (*Pozuzo*); Ucayali (*Previsto*).

TAXONOMY – This species belongs to the “*Pyrrhopyge hadassa* species group” (Evans 1951). The new status is confirmed; see

comments under *Pyrrhopyge hadassa*.

TYPE:

Pyrrhopyge hadassa hanga Evans, 1951. Holotype male with the following labels: / Type – *hanga* Evans/ Pozuzo [**sic**], Huanuco [**recte** Pasco], [Peru] 800-1000 m. (W. Hoffmanns)/. NHMUK.

Pyrrhopyge olivacea Röber, 1925, **stat. rev.**

Pls. 34–35, Figs. 5 (♂) (d, v); Text-Fig. 31 (♂ genitalia)

Syn.: *Pyrrhopyge hadassa henna* Evans, 1951, **syn. n.**

DISTRIBUTION – Northern Peru to central-eastern Bolivia.

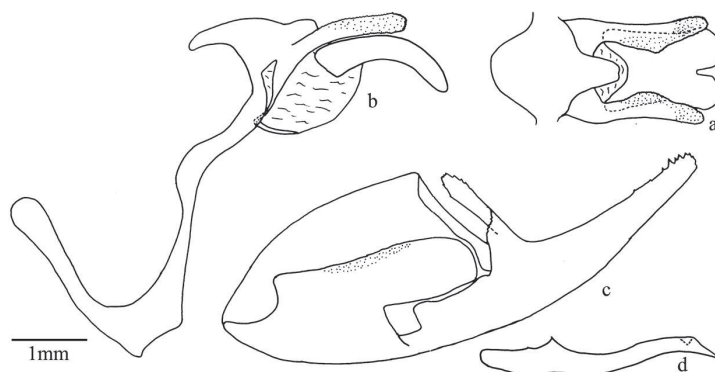
Peru: Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Huambo, Molinopampa, Rodríguez de Mendoza*); Huánuco (*Tingo Maria*); Loreto (*Contamana*); San Martín (*Juanjui, Moyobamba, Pedro Ruiz Gallo, Rioja – Jordania 900 m to Jorge Chávez 1,450 m and Mina de Sal, Santa Cruz*). **Bolivia:** Beni.

TAXONOMY – *Pyrrhopyge olivacea* **stat. rev.** belongs to the “*Pyrrhopyge hadassa* species group” (Evans 1951) and was never properly recognized. Evans (1951) synonymized it with *Pyrrhopyge telassina telassina* (described herein as a new species – *Pyrrhopyge leticia* **sp. n.**) which was accepted by several authors (O. Mielke 2004, 2005). Gerardo Lamas (Lima, Peru) found a male syntype in SMT, which is recognized here as the senior synonym of *Pyrrhopyge hadassa henna* Evans, 1951. The genitalia of the lectotype was dissected and is illustrated here. The species has its status revalidated, explained under *Pyrrhopyge hadassa*.

TYPES:

Pyrrhopyge olivacea Röber, 1925. The original description mentions an unspecified number of specimens from Peru. To ensure the correct identification of *Pyrrhopyge olivacea*, one syntype male found in SMT is **herein designated as the lectotype**; this specimen with the following labels: / Syntypus/ Peru/ *Pyrrhopyge tenuis* Mab. u. Boull./ Staatl. Museum für Tierkunde Dresden/ Stauding. & Bang-Haas Dresden. Ankauf 1961/ Syntype ♂ *Pyrrhopyge olivacea* Röber, 1925 [label by Gerardo Lamas]/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge olivacea* Röber, 1925, O. Mielke, Brockmann & C. Mielke det. 2019/. SMT.

Pyrrhopyge hadassa henna Evans, 1951. Holotype male with the following labels: / Type – *henna* Evans/ Pérou [Amazonas] Huambo M. de Mathan IV^e Trim. 1889/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK. **Syn. n.**



Text-Fig. 31. *Pyrrhopyge olivacea* **stat. rev.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view, d) aedeagus, lateral left view (lectotype).

Pyrrhopyge pseudohadassa Mabilie & Boulet, 1908, **stat. rest.**

Pls. 34–35, Figs. 6 (♂), 7 (♀) (d, v)

Syn.: *Pyrrhopyge tenuis* Mabilie & Boulet, 1908, **praeocc.** (Staudinger, 1888), **syn. n.**

DISTRIBUTION – Central-eastern to southeastern Peru.

Peru: Cusco (*Aguas Calientes, Illapani Viejo, Machu Picchu*); Huánuco (*Tingo Maria*); Junín (*Andamarca, Chanchamayo, La Merced, Perené, Río Colorado, Río Perené, San Ramón, Satipo, Shimá*); Pasco (*Huancabamba, Pichis Road, Pozuzo*); Puno (*Carabaya*); San Martín (*Tocache*); Ucayali (*Previsto*).

The citations **Colombia** (Hayward 1947), **Bolivia** (Mabille & Boullet 1908, Bell 1931, Gareca *et al.* 2006) and **Brazil**: Amazonas (Mabille & Boullet 1908, Bell 1931) are probably incorrect.

TAXONOMY – This species belongs to the “*Pyrrhopyge hadassa* species group” (Evans 1951). The species has its status restored explained under *Pyrrhopyge hadassa*. The examination of the lectotype of *Pyrrhopyge tenuis* Mabille & Boullet, 1908, confirms the new synonymy.

TYPES:

Pyrrhopyge pseudohadassa Mabille & Boullet, 1908. The original description mentions four males and one female from Amazonas [Peru], Bolivia and Peru in MNHN (collection Boullet) and collection Mabille. To ensure the correct identification of *Pyrrhopyge pseudohadassa*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Lectotype / Type/ Amazone supérieur [Peru] 1904 O. Staudinger Coll. Boullet Museum Paris/ *Pyrrhopyge pseudohadassa* M. & B. Lectotype O. Mielke det. 1996/ gen. prep. [O.] Mielke, 1996/. MNHN. Although the lectotype labels were attached in 1996 by O. Mielke, the designation was never published. The type locality is probably Peru.

Pyrrhopyge tenuis Mabille & Boullet, 1908. The original description mentions four males from Peru in MNHN (collection Boullet) and in collection Mabille. To ensure the correct identification of *Pyrrhopyge tenuis*, a preoccupied name by Staudinger (1888), one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Chanchamayo [Junín] Pérou 1882 H. Martinet Coll. Boullet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge tenuis* Mabille & Boullet, 1908, O. Mielke, Brockmann & C. Mielke 2014 det./. MNHN. There are two more male specimens in MNHN from Huancabamba, Peru, herein designated as paralectotypes; they will be similarly labelled as the lectotype. **Syn. n.**

Pyrrhopyge cosnipata O. Mielke, E. Brockmann & C. Mielke, **sp. n.**

Pls. 34–35, Figs. 8 (holotype ♂) (d, v); Text-Fig. 32 (♂ genitalia)

DISTRIBUTION – Southeastern Peru.

Peru: Cusco (*Cosnipata Valley – Quebrada Santa Isabel to San Pedro 1,200 – 1,400 m*).

DESCRIPTION – Male: Forewing length 25–28 mm. Ground colour black. Fore coxae, patagia laterally, tegulae, and anal tuft red; a little red also on lateral parts of dorsal metathorax, posteriorly to tegulae. Fore and hindwings without markings, in one paratype (OM 80.083) there are scarce red scales at base of the dorsal forewing between CuA₁ and 2A and some red scales in the distal half of cell on ventral hindwing, fringes yellow, at forewing from R₅ to tornus and at hindwing, a little larger, from Sc+R₁ to centre of 2A–3A.

Species very similar in colouration to *P. olivacea* **stat. rev.** with which it can easily be confused, differing however by the yellow colour restricted to the fringes on the ventral side of the hindwing (faintly reaching the wing in *P. olivacea* **stat. rev.**). The male differs from *P. olivacea* **stat. rev.** by the uncus with a smaller and narrower terminal median slit and by the harpe of the valva wide with a wider dorso-basal projection, with a wide distal tip dorsally curved to an almost right angle, leaving a distal concavity (in *P. olivacea* **stat. rev.**, the harpe thin with narrower dorso-basal projection, and with distal tip straight, or slightly curved upwards). Both species are sympatric in the Quebrada Santa Isabel (1,200 m), and probably in the Quebrada Quitacalzón (1,100 m) and San Pedro (1,400 m) in the Cosnipata Valley, Cusco, Peru.

Female: unknown.

ETYMOLOGY – The specific name refers to the type locality, Cosnipata Valley, Cusco, Peru.

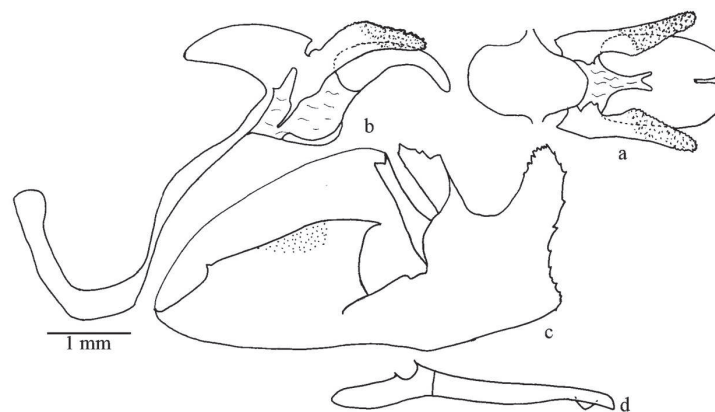
REMARKS – Only known from the type locality.

TYPES:

Holotype male with the following labels: / Holotypus/ Peru. Cusco. Cosnipata Valley. Quebrada Santa Isabel. 13° 02' S. 71° 31' W. 1.200m. 30-III-2016 O. Mielke leg./

gen[itália]. prep[arada]., [O.] Mielke 2014/ Holotypus *Pyrrhopyge cosnipata* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke. det. 2017/. MUSM.

Paratypes: 1 ♂, X-1993, [Quebrada Santa Isabel], Cosnipata Valley, Cusco, Peru, 1200 m, 13°02' S, 71°31' W, Büche leg., OM 35.022 (OM); 1 ♂, 2-6-XI-2012, San Pedro, Cosnipata Valley, Cusco, Peru, 1400 m, O. Mielke leg., OM 73.342 (OM); 1 ♂, 7-XI-2008, Paucartambo to Pilcopata Rd., Bridge [Quebrada Santa Isabel], Cosnipata Valley, Cusco, Peru, 1194 m, 13°02' S, 71°31' W, L. Gibson leg.; collection Loran Gibson; 1 ♂, 30-III-2016, Quebrada Santa Isabel, Cosnipata Valley, Cusco, Peru, 1200 m, 13° 02' S, 71° 31' W, O. Mielke leg., OM 80.083 (OM); 2 ♂, 2-XI-2016, San Pedro, Cosnipata Valley, Cusco, Peru, 1400 m, 13°03' S, 71°30' W. L. Gibson leg., OM 82.681 (OM, MUSM); 1 ♂, 9-XI-2017, Quebrada Quitacalzón, Cosnipata Valley, Cusco, Peru, 1100 m, 13°01' S, 71°30'W, B. Dempwolf leg., OM 85.204 (OM).



Text-Fig. 32. *Pyrrhopyge cosnipata* **sp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view, d) aedeagus, lateral left view (Peru, Cusco, Cosnipata Valley, Quebrada Santa Isabel, 1200 m, X-1993, Büche leg., paratype, OM 35.022).

Pyrrhopyge telassa Hewitson, 1866

Pls. 34–35, Figs. 9 (♂) (d, v)

DISTRIBUTION – Colombia to western and eastern Ecuador.

Colombia: Oriental Cordillera. **Ecuador:** Chimborazo (*Riobamba*); Imbabura (*Lita*); Loja (*Loja, San Francisco*); Morona-Santiago (*El Retiro, Huamboya, Macas, Santiago de Méndez*); Napo (*Misahualli*); Pastaza (*Mera, Río Pastaza*); Tungurahua (*Ambato, Baños, El Rosario, El Topo, Hacienda Santa Inés, Mapoto, Río Blanco, Río Topo, Yunguilla*); Zamora-Chinchipe (*road Loja to Zamora, Zamora*).

The citations **Peru** (Mabille 1878) and **Bolivia**: La Paz and Cochabamba (Weeks 1901, 1905), **Peru**: Madre de Dios and **Brazil**: Rondônia (Cacaulândia) (Garwood *et al.* 2009) are probably incorrect.

TAXONOMY – Evans (1951), based on chromatic aspects of the wings and the fore coxae, mentioned four subspecies of this species, although two of them are sympatric in Huancabamba, Perené and La Merced (*P. telassa phaeax* and *P. telassa silex*). *Pyrrhopyge telassa silex* is a new synonym of *P. telassina*, a species of the *telassa* species complex. The DNA barcode minimum divergence among all taxa is 2.1%, which warrants all being given species status. The minimum divergences are: between *P. telassa* (n= 2) and *P. croceimargo* **stat. rest.** (n= 3) – 3.8%, between *P. telassa* and *P. phaeax* **stat. rest.** (n= 4) – 3.1%, between *P. telassa* and *P. telassina* (n= 2) – 2.1%, between *P. croceimargo* **stat. rest.**, and *P. phaeax* **stat. rest.** – 4.1%, between *P. croceimargo* **stat. rest.**, and *P. telassina* – 5%, and between *P. phaeax* **stat. rest.**, and *P. telassina* – 4.1%.

TYPE:

Pyrrhopyga [sic] telassa Hewitson, 1866. The original description mentions an unspecified number of females [the fig. is male] without data [Ecuador – on the label of the lectotype] in collection Hewitson. To ensure the correct identification of *Pyrrhopyga telassa*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T / Ecuador. Hewitson Coll. 79-69. *Pyrrhopyga telassa* . 3 . / typi[?] *Telassa* Hew./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga telassa* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Pyrrhopyge telassina* Staudinger, 1888**

Pls. 34–35, Figs. 10 (♂) (d, v)

Syn.: *Pyrrhopyge cleopas* Mabille, 1903, **nom. nud.** (ICZN, Art. 12.1)

Pyrrhopyge cleopas Mabille & Boulet, 1908, **syn. n.**

Pyrrhopyge telassa silex Evans, 1951, **syn. n.**

DISTRIBUTION – Northeastern Peru to central-eastern Bolivia.

Peru: Amazonas (*Chachapoyas* – Yambrasbamba, *Rodríguez de Mendoza*); Cusco (*Cosñipata Valley* – *Chontachaca* 950 m); Huánuco (*Tingo Maria*); Junín (*Chanchamayo*, *La Merced*, *Perené*, *Shimá*); Pasco (*Huancabamba*, *Pozuzo*); San Martín (*Rioja* – *Mina de Sal*, *Santa Cruz*). **Bolivia**: Beni; La Paz (*Caranavi*).

TAXONOMY – This species belongs to the “*Pyrrhopyge telassa* species complex” and is a new senior synonym of *Pyrrhopyge cleopas* Mabille & Boulet, 1908 and *Pyrrhopyge telassa silex* Evans, 1951. The genitalia of the lectotypes of *P. telassina*, *P. cleopas* and the [holo]type of *P. telassa silex* were examined and no differences were found. The apophyses of the tegumen are “spined, angled, short, not reaching the uncus division” as described by Evans (1951) for *P. telassa*, so very different from the “*P. telassina* species complex”, where this structure is “seen from side, slender, smooth, not angled, extending well beyond the uncus division” (Evans 1951). This species is sympatric with *P. croceimargo* **stat. rest.** at least in Chontachaca 950 m, Cosñipata Valley, Cusco, Peru.

TYPES:

Pyrrhopyge telassa var. *telassina* Staudinger, 1888. The original description mentions an unspecified number of males from Chanchamayo, [Junín], Peru, and Bolivia. To ensure the correct identification of *Pyrrhopyge telassa* var. *telassina*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Lectotypus/ Chanchamayo [Junín, Peru], Thamm/ Coll. Staudinger/ *Pyrrhopyge telassina* Stgr., 1888, Lectotypus, O. Mielke det. 1979/ gen. prep. [O.] Mielke 1979/ Genital-Unters. Nr. 4717, Zool. Mus. Berlin/; another label will be added: / Lectotypus *Pyrrhopyge telassa* var. *telassina* Staudinger, 1888, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. There are three more males, two from Chanchamayo and one from Bolivia, herein designated as paralectotypes; they will be similarly labelled as the lectotype. Although the lectotype and paralectotypes labels were attached in 1979 by O. Mielke, the designation was never published.

Pyrrhopyge cleopas Mabille, 1903. There is no description; a **nomen nudum**.

Pyrrhopyge cleopas Mabille & Boulet, 1908. The original description mentions one male and one female (collection Boulet), and nine males and one female from Bolivia and Peru in MNHN, and three males in collection Mabille. Twelve specimens (all males) in MNHN were found in two drawers below two labels “*Pyrrhopyge cleopas*” (eight specimens) and “*Pyrrhopyge croceimargo* trans. ad *cleopas*” (four specimens). To ensure the correct identification of *Pyrrhopyge cleopas*, the first specimen (a male) below the label “*Pyrrhopyge cleopas*” is **herein designated as the lectotype**; this specimen with the following label: / Huancabamba [Pasco] Nord Pérou 1904 Herm. Rolle Coll. Boulet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge cleopas* Mabille & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2015/. MNHN. **Syn. n.**

Pyrrhopyge telassa silex Evans, 1951. Holotype male with the following labels: / Type – *syrrax* [sic] Evans/ Inca, Perene, [Junín] Peru. 4000 ft. 8-10. '02. H. Watkins/ Adams Bequest. B.M. 1912.-339/. NHMUK. **Syn. n.**

Pyrrhopyge phaeax* Hopffer, 1874, **stat. rest.*

Pls. 34–35, Figs. 11 (♂), 12 (♀) (d, v)

Syn.: *Pyrrhopyge telassa* var. *tenuis* Staudinger, 1888

DISTRIBUTION – Northeastern to southeastern Peru.

Peru: Amazonas (*Huambo*, *Rodríguez de Mendoza*); Cajamarca (*Charapi*); Cusco (*Agua Calientes*, *Machu Picchu*); Huánuco (*Carpish Pass*, *Tingo Maria*); Junín (*Andamarca*, *Chanchamayo*, *La Merced*, *Perené*, *San Ramón*, *Satipo*); Pasco (*Huancabamba*, *Oxapampa*, *Pozuzo*); San Martín (*Roque*, *Tocache*); Ucayali (*Previsto*).

The citation **Colombia** (Biezanko & Ruffinelli 1962) is probably incorrect.

TAXONOMY – This species belongs to the “*telassa* species complex”. The species has its status reinstated, due to the DNA barcode minimum divergence of 3.95% between *P. telassa* (n= 2) and this taxon (n= 4) and the chromatic aspects of the wings (see Evans 1951).

TYPES:

Pyrrhopyge phaeax Hopffer, 1874. The original description mentions an unspecified number of males from Chanchamayo, [Junín, Peru], Thamm leg. To ensure the correct identification of *Pyrrhopyge phaeax* **stat. rest.**, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / 16217/ *phaeax* Hpf. */ Chanchamayo, [Junín] Peru, Theod. Müller/ gen. prep. [O.] Mielke 1979/ Genital-Unters. Nr. 4716, Zool. Mus. Berlin/ Lectotypus /; two other labels will be added: / Lectotypus / Lectotypus *Pyrrhopyge phaeax* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyge telassa var. *tenuis* Staudinger, 1888. The original description mentions an unspecified number of males from Chanchamayo, [Junín], Peru. To ensure the correct identification of *Pyrrhopyge telassa* var. *tenuis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Lectotypus/ Chanchamayo [Junín, Peru] Thamm/ Coll. Staudinger/ *Cleopas* Mab. *Phaeax* Hopff. v. *tenuis* Stgr./ [O.] gen. prep. Mielke 1979/ Genital-Unters. Nr. 4715, Zool. Mus. Berlin/; another label will be added: / Lectotypus *Pyrrhopyge telassa* var. *tenuis* Staudinger, 1888, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published.

The genitalia of both lectotypes were examined.

Pyrrhopyge croceimargo* Mabille & Boulet, 1908, **stat. rest.*

Pls. 34–35, Figs. 13–14 (♂) (d, v)

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley* – *Chontachaca* 950 m to *Wayquecha* 2,700 m, *Quincemil*); Huánuco (*Tingo Maria*); Madre de Dios (*Upper Río Madre de Dios* – *Amazonia Lodge* (near *Atalaya*, Cusco)); Pasco (*Pozuzo*); Puno (*Chaquimayo*, *Río Inambari*, *San Gaban*); San Martín. **Bolivia**: Cochabamba (*El Limbo*, *Cristal Mayu*); La Paz (*Caranavi*, *Chulumani*, *Coroico*, *Farinas*, *Mapiri*, *Parque Nacional Madidi* 1,500–1,800 m, *Puente Villa*, *Río Zongo*, *Sarampiuni*, *Valle del Zongo* – *Huaylipaya*).

The citations **Colombia** (Hayward 1947) and **Ecuador** (Bell 1931) are probably incorrect.

TAXONOMY – This species belongs to the “*telassa* species complex” and has its status reinstated, because the DNA barcode minimum divergence between *P. telassa* (n= 2) and this taxon (n= 3) is 3.8%. This species is sympatric with *P. telassina*, at least in Chontachaca 950 m, Cosñipata Valley, Cusco, Peru.

TYPES:

Pyrrhopyge croceimargo Mabille & Boulet, 1908. The original description mentions three males and one female from Bolivia in MNHN (collection Boulet) and in collection Mabille. To ensure the correct identification of *Pyrrhopyge croceimargo* **stat. rest.**, one syntype male is **herein designated as the lectotype**, this specimen with the following labels: / Lectotype/ Type/ Bolivie 1904 Herm. Rolle Coll. Boulet. Museum Paris/ *Pyrrhopyge croceimargo* M. & B. Lectotype. O. Mielke det. 1996/. MNHN. Although the lectotype labels were attached in 1996 by O. Mielke, the designation was never published. The genitalia of the lectotype were examined.

Pyrrhopyge leticia* O. Mielke, Brockmann & C. Mielke, *sp. n.

Pls. 34–35, Figs. 15 (holotype ♂) (d, v)

Pyrrhopyge telassina [misidentification]; Evans, 1951. **Cat. Amer. Hesp. 1**, p. 25.

DISTRIBUTION – Northern to central-eastern Peru.

Peru: Huánuco (*Tingo Maria*); Junín (*Chanchamayo*, *El Porvenir*, *La Merced*, *Perené*, *San Ramón* – *Vitoc*, *Satipo*, *Shimá*); Loreto; Pasco (*Pozuzo*); San Martín (*Huayabamba*).

TAXONOMY – Evans (1951) characterized *Pyrrhopyge leticia* **sp. n.** (as *Pyrrhopyge telassina*) with three subspecies, which are allopatric and have differences in their male genitalia: *Pyrrhopyge telassina telassina* Staudinger, 1888 (herein considered a species of the *telassa* complex, and a new senior synonym of *Pyrrhopyge telassa silex*), *P. t. tagra* Evans, 1951 (a junior synonym of *Pyrrhopyge mendax* Mabille & Boulet, 1908, **stat. rest.**) and *P. t. shiva* Evans, 1951 (here considered as full species status, **stat. n.**). The analysis of the DNA barcode of the three subspecies indicates large distances between the taxa, which together with the characteristics of the male genitalia justifies the change of status of all the subspecies to species. The minimum divergences are: between *P. leticia* **sp. n.** (n= 2) and *P. shiva* **stat. n.** (n= 2) – 2.9%, between *P. leticia* **sp. n.** and *P. mendax* **stat. rest.** (= *tagra* Evans) (n= 3) – 2.0%, and between *P. mendax* **stat. rest.** (= *tagra* Evans) and *P. shiva* **stat. n.** – 2.9%.

As there is no available name for the *Pyrrhopyge telassina sensu* Evans, it is hereby described as a new species. Evans's (1951) description of *Pyrrhopyge telassina telassina* applies to the newly described species.

DESCRIPTION – Belongs to the “*mendax* species complex” (= *P. telassina sensu* Evans 1951). Species very similar to *P. shiva* **stat. n.** and *P. mendax* **stat. rest.** as for size, colour and male genitalia; female genitalia not examined. *Pyrrhopyge leticia* **sp. n.** is distinguished from *P. mendax* **stat. rest.** by the apophyses of the tegumen not twisted, narrower and smooth (twisted, thicker and rough in *P. mendax* **stat. rest.**) and from *P. shiva* **stat. n.** by the shorter apophyses of the tegumen and the narrower valva (longer apophyses and larger valva in *P. shiva* **stat. n.**). The marginal and submarginal yellow bands of the wings are highly variable; wings usually equally dark as in *P. mendax* **stat. rest.** and lighter than in *P. shiva* **stat. n.** Only an examination of the genitalia and / or verification of the DNA barcode will ensure a correct identification.

Female: unknown.

ETYMOLOGY – The specific name is in honour of Leticia Mielke Merlin, granddaughter of Olaf H. H. Mielke and niece of Carlos G. C. Mielke.

TYPES:

Holotype male with the following labels: / Holotypus/ 19-X-1989 Rio Tulumayo, 5 KM SE Vitoc, S.[an] Ramón, Junin, Peru, 1100 m, Mielke & Casagrande leg./ gen. prep. [O.] Mielke 2013/ Holotypus *Pyrrhopyge leticia* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 23.477. OM-DZUP.

Paratypes: 14 ♂, same data as the holotype, OM 23.303, OM 23.309, OM 23.321, OM 23.367, OM 23.345, OM 23.357, OM 23.363, OM 23.375, OM 23.420, OM 23.459, OM 23.465, OM 23.492, OM 23.523, OM 23.603 (OM); 1 ♂, Huayabamba, San Martín, Peru, OM 40.832 (OM); 3 ♂, 17-X-1938, 25-X-1938, 14-

III-1939, Satipo, Junín, Peru, Paprzycki leg., OM 9.400, OM 9.403, OM 9.396 (OM); 2 ♂, 9-XIII-1986, 3.IX.1987, Huánuco, Tingo Maria, M. Büche leg. EB01 076, EB01 077 (EB); 2 ♂, 6.X.2014, Shimá, Junín, Peru, S. Kise leg. (CJU); 1 ♂, 2.IX.1999, Shimá, Junín, Peru, J. Uehara leg. (CJU).

Pyrrhopyge mendax* Mabille & Boulet, 1908, *stat. rest.

Pls. 34–35, Figs. 16 (♂) (d, v)

Syn.: *Pyrrhopyge telassina tagra* Evans, 1951, **syn. n.**

DISTRIBUTION – Northern to central-eastern Peru.

Peru: Amazonas (*Rodríguez de Mendoza*); Huánuco (*Tingo Maria*); San Martín (*Juanjui* – *Río Huambo*).

TAXONOMY – The species has its status reinstated according to comments under *Pyrrhopyge leticia* **sp. n.** The holotype of *P. mendax* **stat. rest.** was examined and the genitalia dissected, resulting in the new synonymy of *Pyrrhopyge telassina tagra* with *Pyrrhopyge mendax* **stat. rest.**

TYPES:

Pyrrhopyge mendax Mabille & Boulet, 1908. Holotype (one specimen is mentioned) male with the following labels: / Type/ Holotype/ Pérou 1904 O. Staudinger, Coll. Boulet Museum Paris/ gen. prep. O. Mielke 1996/ *Pyrrhopyge mendax* M. & B. Holotype. O. Mielke det. 1996/. MNHN.

Pyrrhopyge telassina tagra Evans, 1951. Holotype male with the following labels: / Type – *tagra* Evans/ Pérou [Amazonas] Huambo M. de Mathan IV^e Trim. 1889/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK. **Syn. n.**

Pyrrhopyge shiva* Evans, 1951, *stat. n.

Pls. 34–35, Figs. 17 (♂) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley* – *Chontachaca 950 m to San Pedro 1,400 m*); Puno (*Chaquimayo*, *Oroya*). **Bolivia:** Cochabamba (*Cristal Mayu*); La Paz (*Caranavi*, *San Ernesto*).

TAXONOMY – This species belongs to the “*mendax* species complex”. A new status is given according to comments under *Pyrrhopyge leticia* **sp. n.**

TYPE:

Pyrrhopyge telassina shiva Evans, 1951. Holotype male with the following labels: / Type – *shiva* Evans/ S. Ernesto, [La Paz] Bol. [ivia], 1000 m. 68° W., 15° S. IX.00 (Simons)./ Rothschild Bequest B.M.1939-1./ NHMUK.

***Pyrrhopyge terra* Evans, 1951**

Pls. 34–35, Figs. 18 (♂) (d, v)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: Cochabamba; La Paz (*Caranavi*, *Coroico*, *Río Zongo*).

TYPE:

Pyrrhopyge terra Evans, 1951. Holotype male with the following labels: / Type – *terra* Evans/ Boliv[ia]./ J.J.Joicey Coll. B.M.1925-451./ NHMUK.

***Pyrrhopyge creona* H. Druce, 1874**

Pls. 34–35, Figs. 19 (♂) (d, v)

Syn.: *Pyrrhopyga* [*sic*] *aurora* Mabille, 1891

DISTRIBUTION – Northern to central-eastern Peru.

Peru: Amazonas (*Abra Pardo Miguel*, *Alto Nieva 1,930 m*, *Chachapoyas* – *Yambrasbamba*, *Huambo*, *Nueva Esperanza*, *Oso Perdido*

2,200 m, Pedro Ruiz Gallo, Pomacochas, Rodríguez de Mendoza, Santa Rosa); San Martín (Huayabamba, Juanjuí, Moyobamba, Rioja – Naranjos 1,000 m to Jorge Chávez 1,450 m, Santa Cruz 1,200 m).

The citations **Brazil** (Kirby 1879, Mabille 1891, Mabille & Vuillot 1893), **Bolivia** (Mabille & Boullet 1908, Mabille 1912, Draudt 1921), **Brazil**: Amazonas (*Manicoré*) (Evans 1951) and **Brazil**: Amazonas (Palo Jr. 2017) are incorrect.

TYPES:

Pyrrhopyga [*sic*] *creona* H. Druce, 1874. The original description mentions an unspecified number of specimens from Peru in Museum Druce. To ensure the correct identification of *Pyrrhopyga creona*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Peru Druce Coll./ *P. Creona* H. Druce Type/ Godman-Salvin Coll. 1912.-23. *Pyrrhopyge creona*, Druce/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga creona* H. Druce, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [*sic*] *aurora* Mabille, 1891. The original description mentions an unspecified number of specimens from Peru. To ensure the correct identification of *Pyrrhopyga aurora*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Coll. Staudinger/ *Aurora* Mab. n. spec. (Mab.)/ *Pyrrh. aurora* Mab. Type/ *Aurora* Mab., *Creona* Druce/ *creona* Druce/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga aurora* Mabille, 1891, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyge crista Evans, 1951

Pls. 34–35, Figs. 20 (holotype ♂) (d, v)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: La Paz (*Apolobamba, Valle del Zongo – Huaylipaya*).

TYPES:

Pyrrhopyge crista Evans, 1951. Holotype male with the following labels: / Type – *crista* Evans/ Apolobamba [La Paz], Bolivia/ Godman-Salvin Coll. 1912.-23. *Pyrrhopyge kelita*, Hew./ NHMUK.

Pyrrhopyge kelita Hewitson, [1869]

Pls. 34–35, Figs. 21 (♂), 22 (♀) (d, v)

Syn.: *Pyrrhopyge kelita* var. *tristis* Mabille & Boullet, 1908

DISTRIBUTION – Ecuador to central-eastern Bolivia.

Ecuador. Peru: Cusco (*Cosñipata Valley – Pilcopata 550 m to Quebrada Buenos Aires 2,400 m*); Pasco (*Oxapampa*); Puno (*Carabaya, Oroya – Río Inambari, Sandia*). **Bolivia**: Beni (*Río Beni*); Cochabamba (*Chapare, Cristal Mayu*); La Paz (*Apolobamba, Caranavi, Chulumani, Coroico, Irupana, Tamampaya, Valle del Zongo – Huaylipaya, Yungas, Río Zongo*).

TYPES:

Pyrrhopyga [*sic*] *kelita* Hewitson, [1869]. The original description mentions an unspecified number of males from Apolobamba, [La Paz], Bolivia, collections Godman & Salvin and Hewitson. To ensure the correct identification of *Pyrrhopyga kelita*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Apolobamba [La Paz, Bolivia] Hewitson Coll. 79-69. *Pyrrhopyga kelita*. 4./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga kelita* Hewitson, [1869], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge kelita var. *tristis* Mabille & Boullet, 1908. The original description mentions an unspecified number of specimens from Bolivia and Ecuador. To ensure the correct identification of *Pyrrhopyge kelita* var. *tristis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Yungas de La Paz, Bolivie 1000 m. 1904 Herm. Rolle Coll. Boullet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge kelita* var. *tristis* Mabille & Boullet, 1908. O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Pyrrhopyge bouletti Le Cerf, 1922

Pls. 34–35, Figs. 23 (♂), 24 (♀) (d, v); Pls. 36–37, Figs. 1 (♂) (d, v)

DISTRIBUTION – Northern to central-northern Colombia and to northwestern Venezuela.

Colombia: Norte de Santander (*near Pamplona*); Santander (*Chichira*). **Venezuela**: Táchira (*Betania – via Páramo El Tamá, road Delicias – Rubio, El Tamá, via Delicias, El Reposo – via Delicias*).

TYPES:

Pyrrhopyge bouletti Le Cerf, 1922. Holotype male with the following labels: / Type/ Colombie [Norte de Santander] env de Pamplona 1920 P. Rochereau/ *Pyrrhopyge bouletti* Le Cerf ♂ Type F. Le Cerf 1922/. MNHN.

Pyrrhopyge arinas (Cramer, 1777)

Three subspecies.

DISTRIBUTION – Northern to southeastern Peru, southern Venezuela to French Guiana and northern Brazil.

Pyrrhopyge arinas arinas (Cramer, 1777)

Pls. 36–37, Figs. 3 (♂), 4 (♀) (d, v)

Syn.: *Pyrrhopyga* [*sic*] *pedaia* Hewitson, 1866

DISTRIBUTION – Southern Venezuela to French Guiana.

Venezuela: Amazonas (*Yavita*). **Suriname. French Guiana**: Cayenne (*Camp Patawa, Cayenne, Kaw road, Matoury, Roura*).

TYPES:

Papilio arinas Cramer, 1777. The original description mentions an unspecified number of specimens from Suriname in collection Alberti. To ensure the correct identification of *Papilio arinas*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Calkoen Surinam/ Museum Leiden *Yanguna arinas* Cramer det. V. R. Altena '41/ Museum Leiden, Holotype [error], *Papilio arinas* Cramer 1777/; two other labels will be added: / Lectotypus/ Lectotypus *Papilio arinas* Cramer, 1777, O. Mielke, Brockmann & C. Mielke det. 2019/. NBC.

Pyrrhopyga [*sic*] *pedaia* Hewitson, 1866. The original description mentions an unspecified number of males from Ega [Tefé], Amazonas, [Brazil]. To ensure the correct identification of *Pyrrhopyga pedaia*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ [Brazil] Amazons. Ega. Hewitson Coll. 79-69. *Pyrrhopyga arinas*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga pedaia* Hewitson, 1866. O. Mielke, Brockmann & C. Mielke det. 2104/. NHMUK.

Pyrrhopyge arinas erica O. Mielke, Brockmann & C. Mielke, *ssp. n.*

Pls. 30–31, Figs. 8 (holotype ♂) (d, v); Text-Fig. 33 (♂ genitalia)

DISTRIBUTION – Central-eastern Peru.

Peru: Junín (*Satipo*).

DESCRIPTION – Male: “*phidias* form”. Forewing length 28 mm. Ground colour black with some blue iridescence. Head, palpi, a few scales on collar, fore coxae and anal tuft red. Ventral forewing with a slight indication of a white band between discal cell and Cu_{A1}–Cu_{A2}. Fringes dark on forewing and white, slightly crenulated, on hindwing. Forewing with outer margin not excavated in Cu_{A2}–2A. Genitalia similar to that of *Pyrrhopyge arinas arinas*. Apophyses of tegumen angled, spined and reaching the centre of the downcurved and bifid uncus; uncus arms wide apart, U-shaped; valva with the harpe elongated, broad and slightly curved upwards, and dorso-basal process broad and short, until the middle of the ampulla. Only one male is known.

Female: unknown.

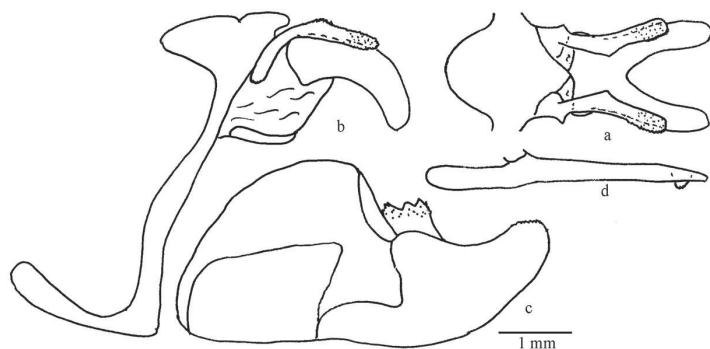
TAXONOMY – This subspecies is distinguished from the other two subspecies of *Pyrrhopyge arinas* due to the lack of a white band on the forewing.

ETYMOLOGY – The specific name is in honour of Erica Costa Mielke, the second daughter of Olaf H. H. Mielke.

REMARKS – Only known from the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ VII-VIII-1987 Satipo, Junín, 100 m, Peru, C. Tello leg./ gen[itália]. pre[parada]., [O.] Mielke 2011/ Holotypus *Pyrrhopyge arinas erica* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ 18.751/. OM-DZUP.



Text-Fig. 33. *Pyrrhopyge arinas erica* **ssp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

Pyrrhopyge arinas temenos (Bell, 1931)

Pls. 36–37, Figs. 2, 5 (♂) (d, v); Pl. 76, Figs. 9 (♀) (d, v)

DISTRIBUTION – Northern to southeastern Peru and northern Brazil (Amazonas and Acre).

Peru: Huánuco (*Tingo Maria*); Junín (*Aldea, La Merced, Perené, Satipo*); Loreto (*Iquitos*); Puno (*Inca*); San Martín (*Rioja, Sacanche*). **Brazil:** Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*); Amazonas (*Benjamin Constant, Tefé*).

TYPE:

Yanguna arinas form *temenos* Bell, 1931. Holotype male with the following labels: / Type H T/ Pérou [Junín], Chanchamayo La Merced C. O. Schunke Reçu Novembre 1904/ Genitalia Slide No. B. M. 3. *Yanguna*/ Br. Mus. Slide 3./ R. Oberthür Coll. Brit. Mus. 1931-136/ *Yanguna temenos* Bell Holotype ♂/ Very like 7 i but note basal joints of palpi, and cilia of fw/. NHMUK.

Pyrrhopyge mopsus (Bell, 1931)

Pls. 36–37, Figs. 6 (♂) (d, v)

Two subspecies.

DISTRIBUTION – Central-eastern to southeastern Peru.

Pyrrhopyge mopsus mopsus (Bell, 1931)

Pls. 36–37, Figs. 6 (♂) (d, v)

DISTRIBUTION – Central-eastern to southeastern Peru.

Peru: Junín (*Chanchamayo, La Merced, Shimá*); Madre de Dios (*Parque Nacional Bahuaja Sonene*).

TYPE:

Yanguna mopsus Bell, 1931. Holotype male with the following labels: / Type H T/ 1810 Chanchamayo [Junín] Peru 1500 m/ ex coll. Hamilton Druce, 1919./ J.J.Joicey Coll. B.M. 1925-451./ Genitalia Slide No. B.M. 1. *Yanguna*/ Br. Mus. Slide 1./ *Yanguna mopsus*. Bell Holotype ♂/ 4 Like *arinas* Cram. Band differently shaped. 4 others exactly, same in BM ? *arinas* sp./. NHMUK.

Pyrrhopyge mopsus elisa O. Mielke, Brockmann & C. Mielke, **ssp. n.**

Pls. 30–31, Figs. 7 (holotype ♂) (d, v); Text-Fig. 34 (♂ genitalia)

DISTRIBUTION – Southeastern Peru.

Peru: Madre de Dios (*Salvación*).

DESCRIPTION – Male: “*phidias* form”. Forewing length 25 mm. Ground colour black, with some blue iridescence. Head, palpi, collar, cheeks, fore coxae and anal tuft red. Fringes white (lost at the forewing apex). Forewing with outer margin not excavated in CuA₂–2A. Genitalia similar to that of *Pyrrhopyge mopsus mopsus*. Apophyses of the tegumen largely beyond the tip of the uncus, wide on the basal half and thin and curved inward on the distal half; uncus bifid with a tiny slit; valva with the harpe hook-shaped, dorso-basal process short and wide and costa crenulated ventrally; sacculus apparently divided into dorsal and ventral portions, based on the less sclerotized central area. Only one male is known.

Female: unknown.

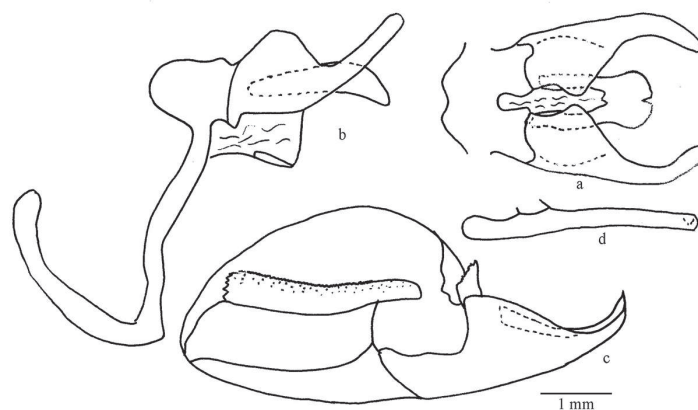
TAXONOMY – This subspecies is distinguished from *Pyrrhopyge mopsus mopsus* due to the lack of a white band on the forewing.

ETYMOLOGY – The specific name is in honour of Elisa Costa Mielke, the first daughter of Olaf H. H. Mielke.

REMARKS – Only known from the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ XI-1995 Salvación, Madre de Dios, Peru, M. Büche leg./ gen[itália]. pre[parada]., [O.] Mielke 2014/ Holotypus *Pyrrhopyge mopsus elisa* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ 20.446/. OM-DZUP.



Text-Fig. 34. *Pyrrhopyge mopsus elisa* **ssp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).

Pyrrhopyge cacaulandia O. Mielke, Brockmann & C. Mielke, **sp. n.**

Pls. 36–37, Figs. 7 (holotype ♂), 8 (allotype ♀) (d, v); Text-Figs. 35 (♂ genitalia), 36 (♀ genitalia)

DISTRIBUTION – Northern Brazil (Rondônia).

Brazil: Rondônia (*Ariquemes, Cacaúlândia*).

DESCRIPTION – Male: very similar to *Pyrrhopyge arinas arinas*. Forewing length 23–25 mm; outer margin not excavated, similar to *P. arinas temenos*. Ground colour black. Head, palpi, inner half of genae, collar, fore coxae and anal tuft light red. Forewing with three superimposed white and semi-hyaline spots in discal cell, between CuA₁–CuA₂ and CuA₂–2A; discal cell spot nearly square, convex proximally and concave distally, second elongated, convex proximally and concave distally and third triangular with base below second spot. Fringes on the forewing of ground colour and on hindwing white between M₁ and the upper half of 2A–3A. Genitalia very different from *P. arinas arinas*: apophyses of tegumen only reaching the base

of the uncus slit; uncus bifid, arms broad and parallel, not divergent, separated between them by a distance nearly one third of the width of each arm; harpe with ventral margin in a nearly right angle. Only two males are known.

Female: similar to male. Forewing length 29 mm. All three superimposed white and semi-hyaline spots more elongated. Genitalia very different from *P. arinas arinas*; sterigma dorsally longer than ventrally; lateral apophyses of sterigma divergent, at the sides of the sterigma and nearly the half sterigma size. Only one female is known.

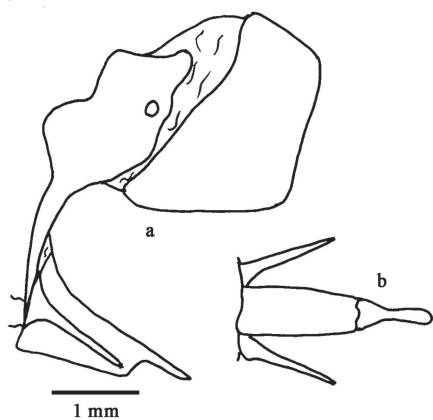
TAXONOMY – A species phenotypically very similar to *Pyrrhopyge arinas arinas*, but there are only some differences in the male and female genitalia. In *P. arinas* the apophyses of the tegumen until nearly the uncus tip; uncus bifid with the arms slender and divergent; harpe with the ventral margin curved, not in a nearly right angle; sterigma ventrally longer than dorsally; lateral apophyses of sterigma nearly parallel, below the sterigma, longer than half of the sterigma, and at the end slightly curved upwards. The only species in this group with a similar uncus is *P. mopsus*, but the apophyses of the tegumen are long and the valvae are pointed, very different (see Bell 1933 and Evans 1951). The presence of a short white streak in Sc-R, above the white cell spot, of *P. mopsus mopsus*, absent in *P. cacauplandia* **sp. n.**, is the only difference between the two species.

ETYMOLOGY – The specific name refers to Cacauplandia, a city in Rondônia, Brazil, where the only known female (allotype) was collected.

REMARKS – Only known from three specimens.



Text-Fig. 35. *Pyrrhopyge cacauplandia* **sp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus, uncus and gnathos, lateral left view; c) right valva, internal view; d) aedeagus, lateral left view (holotype).



Text-Fig. 36. *Pyrrhopyge cacauplandia* **sp. n.** Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma with ventral apophyses of sternum VIII, ventral view (allotype).

TYPES:

Holotype male with the following labels: / Holotypus/ 6-10-VIII-1978 Ariquemes, RO [Rondônia, Brazil,] Furtado leg./ gen[itália]. prep[arada]., [O.] Mielke 2010/ Holotypus *Pyrrhopyge cacauplandia* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 36.647/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ 20-VII-1996 Cacauplandia Rondônia [Brazil] [O.] Mielke leg./ gen[itália]. prep[arada]., [O.] Mielke 2017/ Allotypus *Pyrrhopyge cacauplandia*

O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/ OM 25.585/. OM-DZUP.

Paratype: 1 ♂, 6-10-VIII-1978, Ariquemes, Rondônia, Brazil, Furtado leg., OM 36.880 (OM).

Pyrrhopyge creusae (Bell, 1931)

Pls. 36–37, Figs. 9 (holotype ♂) (d, v)

DISTRIBUTION – Suriname and French Guiana.

Suriname: Sipaliwini. **French Guiana:** Cayenne (*Cacao, Cayenne, Montagne des Chevaux – Galion, Roura*); Saint-Laurent-du-Maroni (*Saint-Laurent-du-Maroni*).

TYPE:

Yanguna creusae Bell, 1931. Holotype male with the following labels: / Type H T/ [French Guiana] Cayenne C. Bar/ R. Oberthür Coll. Brit. Mus. 1931-136/ *Yanguna creusae* Bell Holotype ♂/. NHMUK.

Pyrrhopyge sarpedon (Bell, 1931)

Pls. 36–37, Figs. 10 (♂) (d, v); Pl. 76, Figs. 10 (♀) (d, v)

DISTRIBUTION – Southeastern Colombia to southeastern Peru and northern Brazil (Acre and Rondônia).

Colombia: Amazonas (*Letícia*). **Ecuador:** Napo (*Tena*). **Peru:** Amazonas; Cusco (*Quincemil*); Loreto (*Pebas*). **Brazil:** Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*); Rondônia (*Ariquemes, Cacauplandia*).

TYPE:

Yanguna sarpedon Bell, 1931. Holotype male with the following labels: / Type H T/ Pebas Amazonas [Loreto, Peru] M. de Mathan fin X^{brc}. & 1^{er}. Tr. 1880/ Genitalia Slide No. B.M. 2 *Yanguna* Br. Mus. Slide 2./ R. Oberthür Coll. Brit. Mus. 1931-136/ *Yanguna sarpedon* Bell Holotype ♂/ 1, possible a local race of 5 [??] 6/. NHMUK.

Pyrrhopyge crida Hewitson, 1871

Pls. 36–37, Figs. 11 (♂), 12 (♀) (d, v)

DISTRIBUTION – Western Mexico to western Ecuador and central Colombia.

Mexico: Chiapas; Colima; Oaxaca (*Chimalapa, Tutla*). **Nicaragua:** Chontales (*Santo Domingo*). **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste, Carillo*); Limón (*Barra del Colorado, Guapiles, Puerto Limón*). **Panama:** Veraguas (*Santa Fe*). **Colombia:** Boyacá (*Muzo, Otanche*); Cundinamarca (*Bogotá*). **Ecuador:** Esmeraldas (*Chuchuvi Valley*); Imbabura (*Lita, Paramba*); Pichincha (*Pacto*).

TYPE:

Pyrrhopyga [*sic*] *crida* Hewitson, 1871. The original description mentions an unspecified number of specimens from Chontales, Nicaragua, Th. Belt leg. To ensure the correct identification of *Pyrrhopyga crida*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Nicaragua. Hewitson Coll. 79-69. *Pyrrhopyga crida*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga crida* Hewitson, 1871, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge tatei (Bell, 1932)

Pls. 36–37, Figs. 13 (♀) (d, v); Pl. 76, Figs. 11 (holotype ♂) (d, v)

DISTRIBUTION – Southern Venezuela.

Venezuela: Amazonas (*Cerro Duida*); Bolívar (*Gran Sabana – El Jardín*).

REMARKS – Only few specimens are known. Endemic to the Venezuelan Pantepui.

TYPE:

Yanguna tatei Bell, 1932. Holotype male with the following labels: / Type/ Mt. Duida [Amazonas] Venezuela XII.12.1928/ Ac. 29500 Tate, No. 456/ G1177/ *Yanguna tatei* Bell Holotype ♂/. AMNH.

Gunayan O. Mielke, 2002

Three species

DISTRIBUTION – Colombia to southern Brazil.

***Gunayan rhacia* (Hewitson, [1875])**

Pls. 36–37, Figs. 14 (♂), 15 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Minas Gerais; Rio de Janeiro (*Rio de Janeiro*); São Paulo (*Ubatuba*); Santa Catarina (*Joinville*).

TYPE:

Pyrrhopyga [**sic**] *rhacia* Hewitson, [1875]. The original description mentions an unspecified number of males from Minas Geraes, [Brazil]. To ensure the correct identification of *Pyrrhopyga rhacia*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *Rhacia*. Hew. Minas Geraes [Brazil] Type/ Min. Ger. Rogers/ J.J.Joicey Coll. B.M.1925.-451./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga rhacia* Hewitson, [1875], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Gunayan rubricollis* (Sepp, [1841])**

Two subspecies.

DISTRIBUTION – Eastern Colombia to central-eastern Bolivia, Suriname, French Guiana and northern Brazil.

***Gunayan rubricollis rubricollis* (Sepp, [1841])**

Pls. 36–37, Figs. 17 (♂), 18–20 (♀), 21 (neotype ♀ of *Papilio rubricollis*) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *hadora* Hewitson, 1866

DISTRIBUTION – Eastern Colombia to central-eastern Bolivia, Venezuela, Suriname, French Guiana and northern Brazil.

Colombia: Cundinamarca (*Llanos de San Martín*); Putumayo (*Puerto Leguizamo*). **Ecuador:** Morona-Santiago (*Macas*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Pilcopata 550 m to Chontachaca 950 m*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo*); Loreto (*Contamana, Iquitos*); Madre de Dios (*Parque Nacional del Manu, Tambopata Reserve*). **Bolivia:** Cochabamba (*Chapare*); La Paz (*Caranavi*). **Suriname.** **French Guiana:** Cayenne (*Cayenne, Matoury, Montagne de Kaw*). **Brazil:** Amazonas (*Manicoré, Rio Santiago, São Paulo de Olivença, Tefé*); Pará (*Acará, Belém, Óbidos, Rio Arapiuns, Santarém, Santo Antonio do Tauá*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor, Senador Guiomard*); Rondônia (*Cacaulândia*).

TYPES:

Papilio rubricollis Sepp, [1841]. The original description mentions an unspecified number of specimens from Suriname. No syntypes were found in any museum or collection mentioned above (see “Abbreviations of the collections”) and according to Horn & Kahle (1936) the collection Sepp is lost. To ensure the correct identification of *Papilio rubricollis*, mentioned as *Pyrrhopyge rubricollis* by Evans (1951), the only available specimen, a female from Suriname, as indicated in the drawer before the specimen, is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Museum Leiden *Yanguna rubricollis* Sepp det. V. R. Altena ‘41/ Calkoen ?/ Museum Leiden/ RMNH.INS 9601678/; two other labels will be added: / Neotypus/ Neotypus *Papilio rubricollis* Sepp, [1841] O. Mielke, Brockmann & C. Mielke det. 2018/. NBC.

Pyrrhopyga [**sic**] *hadora* Hewitson, 1866. The original description mentions an unspecified number of males and females from Pará and Tefé (= Ega), Amazonas, Brazil. To ensure the correct identification of *Pyrrhopyga hadora*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Amazonas [Amazonas, Brazil]. Hewitson Coll. 79.-69. *Pyrrhopyga hadora*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga hadora* Hewitson, 1866. O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Gunayan rubricollis flemingi* (Bell, 1947), *stat. rev., stat. n.

Pls. 36–37, Figs. 16 (♂) (d, v)

DISTRIBUTION – Eastern Colombia and northwestern Venezuela.

Colombia: Meta (*Villavicencio*). **Venezuela:** Monagas (*Caripito*).

TAXONOMY – This taxon, described as a species and mentioned by Evans (1951) as a synonym of *Pyrrhopyge rhacia*, is here considered a subspecies of *Gunayan rubricollis flemingi* **stat. rev., stat. n.** The male genitalia are similar to the nominotypical subspecies as the DNA barcode (*G. rubricollis flemingi* **stat. rev., stat. n.** (n= 5) and *G. rubricollis rubricollis* (n= 1)), however it differs by the absence of the white band on the forewing.

Only the holotype, one male and one female are known, the last two from Villavicencio, Meta, Colombia, in OM and LCRO.

TYPE:

Yanguna flemingi Bell, 1947. Holotype male with the following labels: / Holotype/ Caripito. [Monagas] Venez[uela]. Ex. pupa VII-29-’42 Fleming/ *Yanguna flemingi* Holotype ♂ Bell/ G1931/. AMNH.

***Gunayan timaeus* (Bell, 1931)**

Pls. 36–37, Figs. 22–23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Central and eastern Colombia, eastern Peru and northern Brazil (Amazonas and Acre).

Colombia: Cundinamarca (*Bogotá*); Guaviare (*San José del Guaviare*). **Peru:** Junín (*Satipo*); Loreto (*Contamana, Iquitos, Pebas, Río Pacaya, Río Putumayo*); Madre de Dios (*Puerto Maldonado*); San Martín (*Achinamiza*). **Brazil:** Amazonas (*Ipixuna*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*).

TYPE:

Yanguna timaeus Bell, 1931. Holotype male with the following labels: / Type H T/ Pebas, [Loreto] E. Peru. J. Ha[?]xwall/ ♂/ Genitalia Slide No B.M. 5 *Yanguna* Godman-Salvin Coll. 1912.-23. *Yanguna cometes*, Cramer./ Brit. Mus. Slide # 5/ *Yanguna timaeus* Bell Holotype ♂/ Like *cometes*, but the colour of thorax & legs (ex. forelegs) beneath 3 others exactly same in BM “Bogota” + Pebas, Peru/. NHMUK.

Yanguna Watson, 1893

Nine species.

DISTRIBUTION – Mexico to central-eastern Brazil.

Yanguna blossomiae* Williams, 1931, *stat. rev.

Two subspecies.

DISTRIBUTION – Western and eastern Ecuador to southeastern Peru, French Guiana and northwestern Brazil.

Yanguna blossomiae blossomiae* Williams, 1931, *stat. rev.

Pls. 38–39, Figs. 1 (♂), 2 (♀) (d, v); Pls. 74–75, Figs. 10 (holotype ♀) (d, v); Text-Figs. 37 (♂ genitalia), 38 (♀ genitalia)

DISTRIBUTION – Western and eastern Ecuador to southeastern

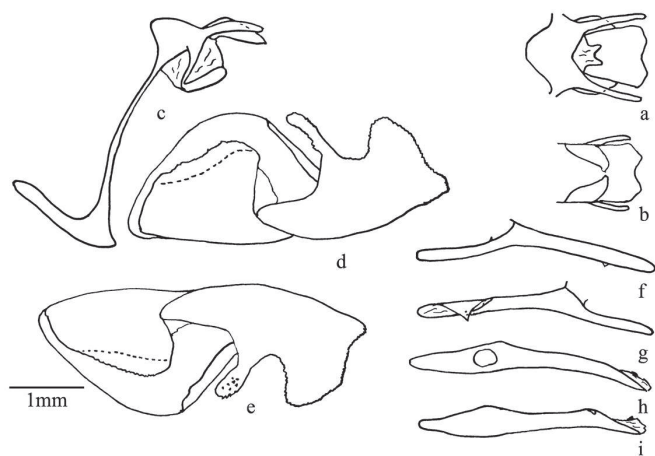
Peru and northwestern Brazil (Acre and Rondônia).

Ecuador: Chimborazo (*Alausi – Huigra*). **Peru:** Cusco (*Marcapata*); Huánuco (*Tingo Maria*); Loreto (*Iquitos*). **Brazil:** Acre (*Bujari, Marechal Thaumaturgo*); Rondônia (*Ariquemes, Ouro Preto d'Oeste*).

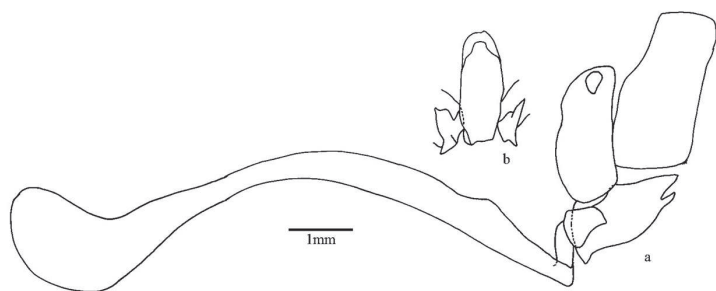
TAXONOMY – This species was considered a synonym of *Yanguna thelersa*, but it is here revalidated by the differences in the DNA barcode from *Y. thelersa* (n= 4) and *Y. blossomiae stat. rev.* (n= 3, including the holotype). The minimum divergence between the two species is 6.48%. The species is very similar to *Y. thelersa*, but differs by the dark fringes on both wings and by, at least, the male and female genitalia illustrated below.

TYPE:

Yanguna blossomiae Williams, 1931. Holotype (one male [*recte* female] is mentioned) with the following labels: / Huigra, [Chimborazo] Ecuador Alt. 4000 ft./ Type *Yanguna blossomae* [*sic*] R. C. Williams, Jr. 7588/ Ex Collection W.J.Coxey/ (W.J.Coxey) Mar., 1930 A.N.S. Lot 217/. ANSP, transferred to CMP.



Text-Fig. 37. *Yanguna blossomiae blossomiae stat. rev.* Male genitalia: a) tegumen and uncus, dorsal view; b) gnathos and uncus, ventral view; c) tegumen, saccus, uncus and gnathos, lateral left view; d) right valva, internal view; e) left valva, internal view; f) aedeagus, lateral left view; g) aedeagus, lateral right view; h) aedeagus, dorsal view; i) aedeagus, ventral view (18-20-IX-2003, 50 KM NO Bujari, Acre, Brazil, O.-C. Mielke leg., DZ 29.386, DZUP).



Text-Fig. 38. *Yanguna blossomiae blossomiae stat. rev.* Female genitalia: a) segment VIII, sterigma, bursa copulatrix and papilla analis, lateral left view; b) sterigma, ventral view (Brazil, Rondônia, Ouro Preto d'Oeste, 17-31-VIII-1987, C. Elias leg., DZ 3.477, DZUP).

Yanguna blossomiae galion O. Mielke, Brockmann C. Mielke, *ssp. n.*

Pls. 38–39, Figs. 3 (holotype ♂), 4 (allotype ♀) (d, v)

DISTRIBUTION – French Guiana.

French Guiana: Cayenne (*Cayenne – Montagne des Chevaux*).

DESCRIPTION – Male: forewing length 26 mm. Very similar to *Y. blossomiae blossomiae stat. rev.*, but differing by the more elongated forewing. Forewing with three very small apical spots in R_3 – R_4 , R_4 – R_5 and R_5 – M_1 , smaller than in *Y. b. blossomiae*.

Female: forewing length approximately 33 mm (the only known female has the apex destroyed). Apical spots a little larger than in male. The allotype was collected in copula with the holotype.

There is no difference in the DNA barcode between both subspecies; *Y. blossomiae galion ssp. n.* (n= 2).

ETYMOLOGY – The specific name refers to the city in French Guiana, where the holotype and allotype were collected.

REMARKS – Only known from four specimens from the type locality.

TYPES:

Holotype male with the following labels: / Holotypus/ Guiana Francesa, Montagne de [*sic*] Chevaux, 4°43'30" N, 52°24'37"W, Galion, 16-X-1988 J. Y. Gallard leg. incopula [with the allotype]/ gen[italia]. prep[arada]., [O.] Mielke 2013/ Holotypus *Yanguna blossomiae galion* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2020/ OM 71.427/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Guiana Francesa, Montagne de [*sic*] Chevaux, 4°43'30" N, 52°24'37"W, Galion, 16-X-1988 J. Y. Gallard leg. incopula [with the holotype]/ gen[italia]. prep[arada]., [O.] Mielke 2013/ Holotypus *Yanguna blossomiae galion* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2020/ OM 71.417/. OM-DZUP.

Paratypes: 2 ♂, same locality as the holotype, 25-X-1989, 29-X-1989, Brevignon leg., OM 71.447, OM 71.437 (OM); 1 ♂ same locality and collector as the holotype, 17-VIII-1997; EB).

Yanguna moa O. Mielke, Casagrande & Brockmann, *ssp. n.*

Two subspecies.

DISTRIBUTION – Central-eastern Peru, French Guiana and northern Brazil.

Yanguna moa moa O. Mielke, Casagrande & Brockmann, *ssp. n.*

Pls. 38–39, Figs. 5 (holotype ♂) (d, v); Text-Fig. 39 (♂ genitalia)

DISTRIBUTION – Central-eastern Peru and northern Brazil.

Peru: Junín (*Satipo – Río Anapate*); Huánuco (*Tingo Maria*); San Martín (*Tocache*). **Brazil:** Pará (*Rio Arapiuns*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*); Rondônia (*Vilhena*).

DESCRIPTION – Male: forewing length 26–28 mm. A species very similar externally to *Y. blossomiae blossomiae stat. rev.* and *Y. thelersa*. The male genitalia of these species quite different and are illustrated for a better comparison. The differences between the male genitalia are the shorter apophyses of the tegumen reaching only the 2/3 of the tip of the uncus; the more rounded and distally narrower uncus with two well separated distal projections; the appearance of asymmetric valvae with harpes of two valvae distally only slightly projected, as in *Y. blossomiae stat. rev.*, and each with two dorsal projections, at the right valva, one subterminal, narrower than that of left valva, and another proximal, directed basically and thicker than that of left valva, and at the left valva, one subterminal rounded and another proximal longer and curved ventrally; sacculus reaching the costa at right valva; aedeagus with a well-developed projection on right side, vesica without cornutus. Forewing with or without small white apical spots in R_3 – R_4 , R_4 – R_5 , R_5 – M_1 and white cell spot usually narrower at R than at CuA_1 . On the hindwing, dark orange spot at the base of the dorsal side normally larger and more rounded than in *Y. blossomiae blossomiae stat. rev.* and *Y. thelersa*, reaching the end of the discal cell.

Yanguna moa moa ssp. n. is also quite similar to *Y. cometes cometides*, from which it differs by the larger dark orange basal region of hindwing dorsally and by whitened fringes of same wing, while in *Y. cometes cometides* this dark orange region is reduced to a spot within discal cell and fringes white, and by male genitalia.

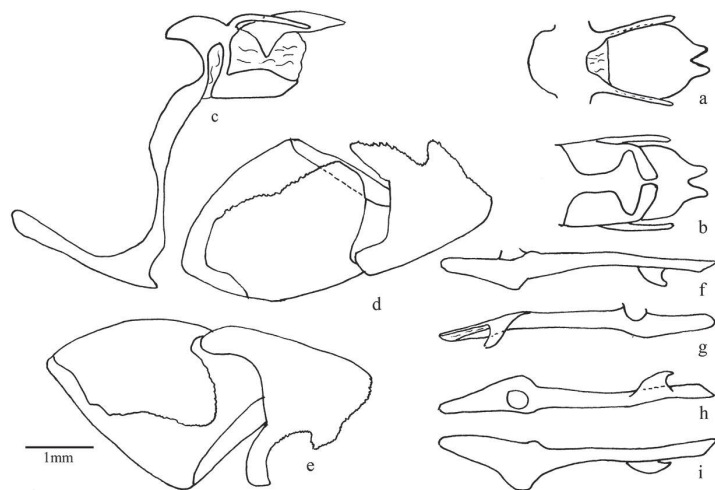
Female: unknown.

ETYMOLOGY – The specific name refers to the Rio Moa, Mâncio Lima – *Parque Nacional da Serra do Divisor*, Acre, Brazil, where one of the paratypes was collected.

TYPES:

Holotype male with the following labels: / Holotypus/ Tingo Maria, Huánuco, Peru, 670 m, IX-1990, Büche leg./ gen[itália]. pre[parada]., [O.] Mielke 2013/ Holotypus *Yanguna moa moa* O. Mielke, Casagrande & Brockmann. O. Mielke, Casagrande & Brockmann det. 2017/ OM 36.018/. OM-DZUP.

Paratypes: 1 ♂, 10-III-1987, Tingo Maria, Huánuco, Peru, 650 m, ex coll. Büche, HESP-EB 03 288 (EB); 1 ♂, 29-III-1987, Tingo Maria, Huánuco, Peru, 650 m, ex coll. Büche, HESP-EB 03 289 (EB); 2 ♂, IX-1990, Tingo Maria, Huánuco, Peru, 670 m, Büche leg., OM 35.984, OM 36.048 (OM); 1 ♂, IX-2001, Tingo Maria, Huánuco, Peru, 650 m, ex coll. Büche, HESP-EB 03 290 (EB); 1 ♂, IX-2013, Tingo Maria, Huánuco, Peru, J. J. Ramírez leg., OM 80.378 (OM); 2 ♂, III-2014, Tocache, San Martín, Peru, ex coll. D. González A. (EB); 4 ♂, Peru or Bolivia, ex coll. Claude Lestrade, Paris (EB); 1 ♂, 20-27-VI-2013, Rio Moa, Parque Nacional do Serra do Divisor, Mâncio Lima, Acre, Brazil, [O.] Mielke, Casagrande, Carneiro, Dias & Dolibaina leg., 7°26'52" S, 73°39'55" W, DZ 29.407 (DZUP); 1 ♂, Anapate valley, Satipo, Junín, Peru, 700 m, X-1996, Carlos Tello leg., HESP-EB 01 102 (EB); 1 ♂, Anapate valley, Satipo, Junín, Peru, 700 m, X-1986, HESP-EB 01 093 (EB); 1 ♂, Tocache, San Martín, Peru (EB).



Text-Fig. 39. *Yanguna moa moa* **ssp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) gnathos and uncus, ventral view; c) tegumen, saccus, uncus and gnathos, lateral view; d) right valva, internal view; e) left valva, internal view; f) aedeagus, lateral left view; g) aedeagus, lateral right view; h) aedeagus, dorsal view; i) aedeagus, ventral view (holotype).

Yanguna moa guiana* O. Mielke, Casagrande & Brockmann, **ssp. n.*

Pls. 38–39, Figs. 6 (holotype ♂) (d, v)

DISTRIBUTION – French Guiana.

French Guiana: Cayenne (*Montagne des Chevaux – Galion*).

DESCRIPTION – Male: forewing length 28 mm. Very similar to *Y. moa moa* **ssp. n.**, but differing by the more elongated forewing and by the strong concavity at the external margin of the hindwing. Forewing without apical spots. Only one male is known.

Female: unknown.

In the DNA barcode there is a minimum divergence of 1.53% between both subspecies: *Yanguna moa moa* **ssp. n.** (n= 2) and *Y. moa guiana* **ssp. n.** (n= 1).

ETYMOLOGY – The specific name refers to French Guiana, where the holotype was collected.

REMARKS – Only known from the holotype.

TYPE:

Holotype male with the following labels: / Holotypus/ Guiana Francesa Montagne des Chevaux 4°43'30" N, 52°24'30" W Galion, 14-VII-1989 J. Y. Gallard leg./ gen[itália]. prep[arada]., [O.] Mielke 2013/ Holotypus *Yanguna moa guiana* O. Mielke, Casagrande &

Brockmann. O. Mielke, Casagrande & Brockmann det. 2017/ OM 71.407/. OM-DZUP.

***Yanguna thelersa* (Hewitson, 1866)**

Pls. 38–39, Figs. 7 (♂), 8–10 (♀) (d, v); Text-Figs. 40 (♂ genitalia), 41 (♀ genitalia)

DISTRIBUTION – Western and eastern Ecuador to southeastern Peru, French Guiana and northern Brazil.

Ecuador: Chimborazo (*Huigra*); Napo (*Archidona, Río Napo*).

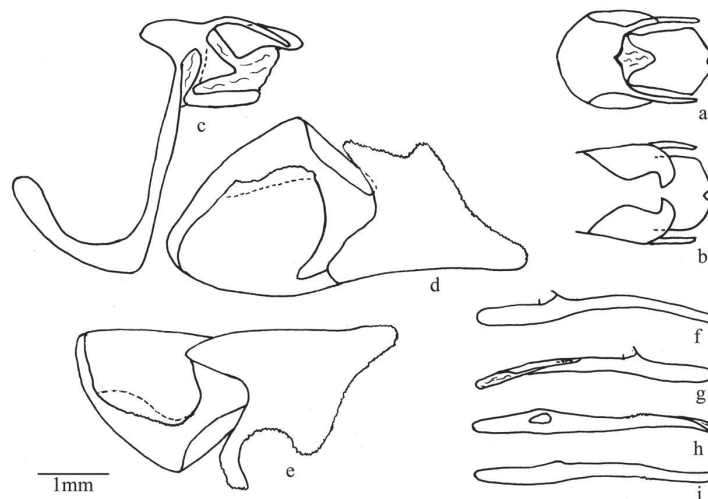
Peru: Cusco; Huánuco (*Tingo Maria*); Junín (*Chanchamayo, Río Colorado*); Loreto (*Água Blanca, Alto Río Contaya, Iquitos, Río Putumayo*); Madre de Dios (*Tambopata Reserve*); Pasco (*Chuchurras*); San Martín (*Tarapoto, Yumbatos*). **French Guiana:** Cayenne (*Montagne des Chevaux – Galion*). **Brazil:** Amazonas (*São Paulo de Olivença*); Pará (*Rio Arapiuns*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*); Rondônia (*Cacaulândia, Vilhena*); Maranhão (*Centro de Guilherme – Montes Áureos*).

TAXONOMY – A species very similar to *Yanguna blossomiae* **stat. rev.** and *Yanguna moa* **sp. n.** The male genitalia of these species are quite different and are illustrated for a better comparison. The differences are in the slightly shorter apophyses of the tegumen nearly reaching the uncus tip; in the more squared uncus with an small distal concavity; the appearance of the asymmetric valvae, with the harpes of both valvae distally projected and each with two dorsal projections, one median, short and stubby, and another proximal, triangular at the right valva, and long, slender and curved at the left valva, sacculus not reaching the costa; aedeagus with a very small projection on the right side, shorter than in *Y. blossomiae* **stat. rev.** and in *Y. moa* **sp. n.**, vesica without cornutus.

Forewing without white apical spots at least in males. The fringes said to be black at the upper hindwing (Evans 1951: 36), may be also greyish as in the similar species, *Y. blossomiae* **stat. rev.** and *Y. moa* **sp. n.**

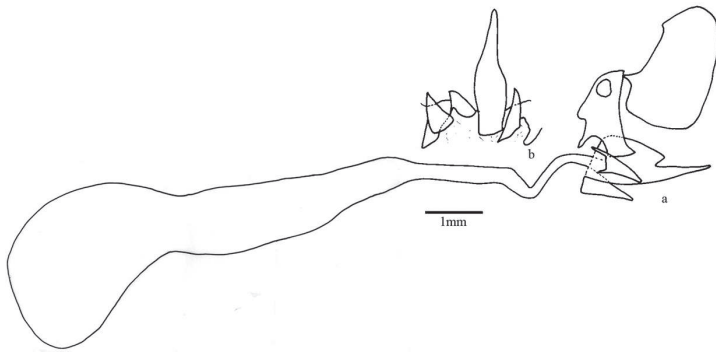
TYPE:

Pyrrhopyga [**sic**] *thelersa* Hewitson, 1866. The original description mentions an unspecified number of specimens from St. Paulo [de Olivença], Amazonas, [Brazil] in collection Hewitson. To ensure the correct identification of *Pyrrhopyga thelersa*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ St. Paulo. [São Paulo de Olivença, Amazonas, Brazil]. Hewitson Coll. 79-69. *Pyrrhopyga thelersa*. 4./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga thelersa* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.



Text-Fig. 40. *Yanguna thelersa*. Male genitalia: a) tegumen and uncus, dorsal view; b) gnathos and uncus, ventral view; c) tegumen, saccus, uncus and gnathos, lateral left view; d) right valva, internal view; e) left valva, internal view; f) aedeagus, lateral left view; g) aedeagus, lateral right view; h) aedeagus, dorsal view; i) aedeagus, ventral view.

g) aedeagus, lateral right view; h) aedeagus, dorsal view; i) aedeagus, ventral view (Brazil, Acre, Mâncio Lima, Parque Nacional da Serra do Divisor, sede, 20-27-VI-2013, [O.] Mielke, Casagrande, Carneiro, Dias & Dolibaina leg., DZ 28.784).



Text-Fig. 41. *Yanguna thelersa*. Female genitalia: a) segment VIII, sterigma, bursa copulatrix and papilla analis, lateral left view; b) sterigma, ventral view (Brazil, Pará, Rio Arapiuns, VIII-21980, Kesselring leg., OM 21.709).

Yanguna cometes (Cramer, 1779)

Four subspecies.

DISTRIBUTION – Central, eastern and southern Colombia to central-eastern Bolivia, northwestern and southern Venezuela, Suriname, French Guiana and northern Brazil.

Yanguna cometes cometes (Cramer, 1779)

Pls. 38–39, Figs. 11 (neotype ♂ of *Papilio cometes*), 12 (♀) (d, v)

DISTRIBUTION – Central-eastern Peru, Suriname, French Guiana and northern Brazil.

Peru: Pasco (*Pozuzo*). **Suriname.** **French Guiana:** Cayenne (*Camp Patawa, Larivot, Montagne des Chevaux – Galion, Roura, Tonate*); Saint-Laurent-du-Maroni (*Maroni River*). **Brazil:** Roraima (*Boa Vista*); Amazonas (*Benjamin Constant, Paricatuba*); Pará (*Óbidos*).

TYPE:

Papilio cometes Cramer, 1779. The original description mentions an unspecified number of specimens from Suriname. No syntypes were found in the NBC or NHMUK, where many of the Cramer's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Papilio cometes*, mentioned as *Pyrrhopyge cometes cometes* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / St. Laurent, Maroni River, French Guiana, July-Sept. 1915/ J. J. Joicey Coll. B.M. 1925-451/ *cometes* Cramer Det. W. H. Evans 28/2/1939/ Molecular 0247278501/ NHMUK 012824139/, and two other labels will be added: / Neotypus/ Neotypus *Papilio cometes* Cramer, 1779 O. Mielke, Brockmann & C. Mielke det. 2010/. NHMUK. Unfortunately, there is no available specimen from Suriname, the type locality.

Yanguna cometes angeli Orellana, [2010]

Pls. 38–39, Figs. 13 (holotype ♂), 14 (paratype ♀) (d, v)

DISTRIBUTION – Central and eastern Colombia to northwestern Venezuela.

Colombia: Cundinamarca (*Bogotá*); Meta (*Río Negro*). **Venezuela:** Táchira (*Río Frío*).

TYPE:

Yanguna cometes angeli Orellana, [2010]. Holotype male from Río Frío, Táchira, Venezuela, II-1990, 600 m, F. Romero leg. CFR.

Yanguna cometes staudingeri (Plötz, 1879)

Pls. 38–39, Figs. 15 (neotype ♂ of *Pyrrhopyga staudingeri*), 16 (♀) (d, v); Pls. 74–75, Figs. 11 (♀) (d, v)

DISTRIBUTION – Southern Colombia to central-eastern Peru and southern Venezuela.

Colombia: Putumayo (*Mocoa*). **Ecuador:** Morona-Santiago (*Macas*); Napo (*Tena*). **Peru:** Amazonas (*Huambo, Rodríguez de Mendoza*); Cajamarca; Huánuco (*Río Pachitea, Río Monzón, Tingo Maria*); Junín (*Anapati Valley, Chanchamayo, La Merced, Perené, Río Colorado, San Ramón, Satipo, Shimá*); Loreto (*Boquerón Padre Abad, Pebas, Upper Putumayo*); Pasco (*Huancabamba, Oxapampa, Pozuzo*); San Martín (*Huayabamba, Juanjui, Rioja – El Afluente 1,200 m, Santa Cruz 1,200 m*); Ucayali (*Pucallpa*). **Venezuela:** Bolívar (*Auyán Tepui*).

TYPE:

Pyrrhopyga [*sic*] *staudingeri* Plötz, 1879. The original description mentions an unspecified number of specimens without data. No syntypes were found in MfN or ZSM, where nearly all Plötz's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Pyrrhopyga staudingeri*, mentioned as *Pyrrhopyge cometes staudingeri* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Rondos, Tingo Maria, [Huánuco], Peru, 850 m, 27-X-1971 Villegas leg./ Neotypus *Pyrrhopyga staudingeri* Plötz, 1879 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 4629/. DZUP.

Yanguna cometes cometides Mabille & Boulet, 1908

Pls. 38–39, Figs. 17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Eastern Peru to central-eastern Bolivia and northwestern Brazil (Acre).

Peru: Cusco (*Consuelo 1,200 m, Cosñipata Valley – Chontachaca 950 m to Quebrada Quitacalzón 1,100 m, Quincemil*); Junín (*Perené*); Loreto (*San Juan de Polí*); Madre de Dios (*Parque Nacional Bahuaja Sonene, Shintuya, Tambopata Reserve*); San Martín (*Tarapoto*). **Bolivia:** La Paz (*Caranavi, Chulumani, Farinas, Iturralde, Río Zongo, Sarampiuni*). **Brazil:** Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor*).

The citations **Argentina** (Misiones) (Hayward 1932, 1935, 1941, 1948, 1973) are probably incorrect.

TYPE:

Yanguna staudingeri var. *cometides* Mabille & Boulet, 1908. Holotype male with the following labels: / Bolivie 1904 O. Staudinger Coll. Boulet Museum Paris/; two other labels will be added: / Holotypus/ Holotypus *Yanguna staudingeri* var. *cometides* Mabille & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Yanguna erebus (Plötz, 1879)

Pls. 38–39, Figs. 19 (holotype ♂), 20 (♂) (d, v)

DISTRIBUTION – Central-eastern Brazil (Bahia and Espírito Santo).

Brazil: Bahia; Espírito Santo (*Conceição da Barra*).

REMARKS – Only two males are known.

TYPE:

Erycides erebus Plötz, 1879. Holotype (one specimen is mentioned) male with the following labels: / 4904/ Bahia [Brazil] Sello/ 71: 17/ *erebus* Pl. Type/ *Rhacia* N., ? *Rhacia* Hew. Exot. Butt. V. Pyr. t S f.37/ MB 4905 type/ Lectotypus [error]/. MfN.

Yanguna cosyra (H. Druce, 1875)

Pls. 38–39, Figs. 21 (♂), 22–23 (♀) (d, v)

DISTRIBUTION – Mexico to western Ecuador, central Colombia and eastern Ecuador to Peru.

Mexico. **Guatemala:** Alta Verapaz (*Cubilguitz*). **Honduras:** Atlántida (*La Ceiba*). **El Salvador** (?). **Nicaragua:** Chontales (*Santo Domingo*). **Costa Rica:** Alajuela (*Road to Colonia Palmareña, San Cristóbal*); Cartago (*Turrialba*); Guanacaste (*Área de Conservación*

Guanacaste); Puntarenas (Coronado, Osa – Coronado, San Vito); San José (Mastatal). **Panamá:** Chiriquí (Bugaba); Colón (Gatun, Piña); Darién (Cana); Panamá (El Llano); Panamá Oeste (Cerro Campana); Veraguas. **Colombia:** Boyacá (Otanche); Cundinamarca (Bogotá); Boyacá (Muza); Risaralda (Montezuma); Valle del Cauca (Anchicaya). **Ecuador:** Esmeraldas (San Francisco, Durango, La Sabalera); Napo (Río Jondachi); Santo Domingo de los Tsáchilas (Alluriquín). **Peru.**

TYPE:

Pyrrhopyga [sic] *cosyra* H. Druce, 1875. The original description mentions an unspecified number of specimens from Bugaba, Veraguas, [Panamá] in collection Druce. To ensure the correct identification of *Pyrrhopyga cosyra*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ B.C.A.Lep.Rhop. *Yanguna cosyra*, Druce./ Bugaba, [Chiriquí] Panamá. Arce./ *Pyrrhopyga cosyra* H. Druce Type./ Type. Sp. figured./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga cosyra* H. Druce, 1875, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Yanguna tetricus Bell, 1931

Pls. 38–39, Figs. 24 (♂) (d, v)

DISTRIBUTION – Central-eastern Peru, Guyana and northern Brazil.

Peru: Junín (Satipo); Madre de Dios; San Martín (Moyobamba).

Guyana: Cayuni Mazaruni (Mont Roraima). **Brazil:** Roraima; Rondônia (Cacaulândia); Mato Grosso (Diamantino).

TYPE:

Yanguna tetricus Bell, 1931. Holotype male with the following labels: / Type H T/ Roraima. [Guyana] Crowley Bequest. 1901-78./ Roraima/ Genitalia Slide No. B.M. 4. *Yanguna* Brit. Mus. Slide 4./ *Yanguna tetricus* Bell Holotype ♂ / 4. apparently a *rubricollis* form. But note orange fore coxae, and black collar/. NHMUK.

Yanguna spatiosa (Hewitson, 1870)

Two subspecies.

DISTRIBUTION – Western Colombia to western Ecuador, eastern Ecuador to central-eastern Peru.

Yanguna spatiosa spatiosa (Hewitson, 1870)

Pls. 40–41, Figs. 1 (♂), 2 (♀) (d, v); Text-Figs. 42 (♂ genitalia), 43 (♀ genitalia)

DISTRIBUTION – Western and eastern Ecuador to central-eastern Peru.

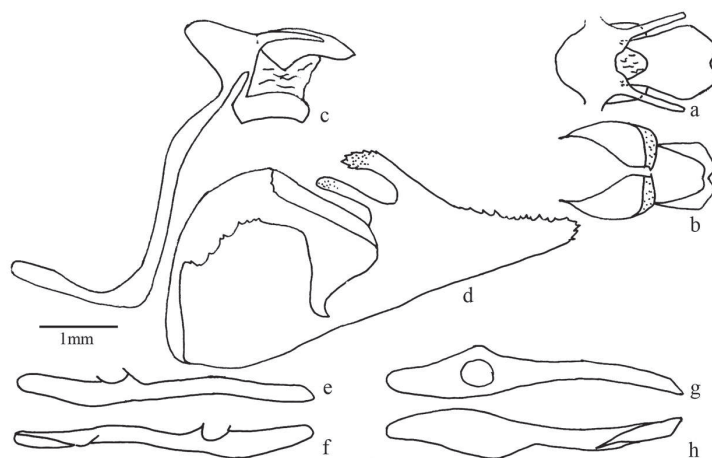
Ecuador: Esmeraldas (Chuchuvi); Loja (San Francisco); Morona-Santiago (Huamboya, Macas, Pablo Sexto); Pastaza (Canelos, Mera, Sarayacu); Santo Domingo de los Tsáchilas (Alluriquín); Tungurahua (Ambato, Baños, Hacienda La Mascota, Hacienda Santa Inés, Río Topo); Zamora-Chinchipe (Zamora). **Peru:** Amazonas (Alto Nieva 1,930 m, Cerro de Huambo, Molinopampa, Oso Perdido 2,200 m, Pomacochas 2,000 m, Rodríguez de Mendoza); San Martín (Huayabamba, Huicungo, Jorge Chávez, Juanjuí, Puente Nieva, Rioja – El Afluente 1,300 m, Santa Cruz 1,200 m).

The citations **Nicaragua** (Kirby 1879) and **Panamá:** Chiriquí (Staudinger 1888) are probably incorrect.

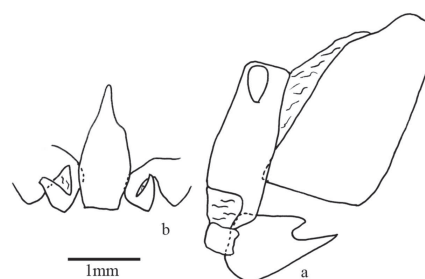
TAXONOMY – The male and female genitalia are illustrated for comparison with *Yanguna aspilos* *stat. rest.*, a similar species.

TYPE:

Pyrrhopyga [sic] *spatiosa* Hewitson, 1870. The original description mentions an unspecified number of specimens from St. Inés, [Tungurahua], Ecuador, Buckley leg. To ensure the correct identification of *Pyrrhopyga spatiosa*, one syntype female is **herein designated as the lectotype**, this specimen with the following labels: / Type/ *spatiosa*/ Ecuador. Hewitson Coll. 79-69. *Pyrrhopyga spatiosa*. 5./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga spatiosa* Hewitson, 1870, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.



Text-Fig. 42. *Yanguna spatiosa spatiosa*. Male genitalia: a) tegumen and uncus, dorsal view; b) gnathos and uncus, ventral view; c) tegumen, saccus, uncus and gnathos, lateral view; d) right valva, internal view; e) aedeagus, lateral left view; f) aedeagus, lateral right view; g) aedeagus, dorsal view; h) aedeagus, ventral view (Ecuador, Zamora-Chinchipe, between Loja and Zamora, Puente Zurita, 6-XII-2013, R. Westphal leg., 2000 m, OM 76.088).



Text-Fig. 43. *Yanguna spatiosa spatiosa*. Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (Ecuador, Tungurahua, Santa Inés, OM 41.247).

Yanguna spatiosa julian O. Mielke, Brockmann & C. Mielke, *ssp. n.*

Pls. 40–41, Figs. 4 (holotype ♂), 5 (allotype ♀) (d, v)

Yanguna spatiosa; Valencia M. *et al.* 2005: 202, pl. 41, fig. 6 (male dorsal, ventral).

DISTRIBUTION – Western Colombia.

Colombia: Chocó (Valencia M. *et al.* 2005); Risaralda (Valencia M. *et al.* 2005); Valle del Cauca (Aguacatal, Buenaventura – Río Dagua, Darién – Lago Calima).

The following localities probably refer to this subspecies.

Colombia: Chocó (El Palmar – Tabor, San Juan del Palmar); Cundinamarca (Guayabetal – Susumuco); Meta (Upper Río Negro); Risaralda (Chami, Montezuma, Otún); Tolima (San Antonio, Santa Margarita); Valle del Cauca (Buga, Carrapatos Valley, Farallones).

DESCRIPTION – This subspecies is very similar to *Yanguna spatiosa spatiosa* by the forewing length, body and wing colour, and male genitalia, but it differs by the presence of a band of the same colour as on the basal area of the dorsal hindwing (orange-red), nearly 3 mm wide, on the ventral hindwing, between the base of Sc+R₁–Rs, crossing the central area of the discal cell, until the extreme base of CuA₁–CuA₂; this band is narrower and more diffuse than in *Yanguna aspilos lara* *ssp. n.*

ETYMOLOGY – The specific name is in honour to our friend Julian Adolfo Salazar Escobar, Museo de Historia Natural, Universidad de Caldas, Colombia, for his contribution to the Colombian lepidopterology.

TYPES:

Holotype male with the following labels: / Holotypus/ 17-XII-1971, Lago Calima, NO [NW] de Cali, [Valle del Cauca], Colombia, 1500m, [K.] Brown [Jr.] leg./ gen[itá]lia. prep[arada]. [O.] Mielke 1990/ Holotypus *Yanguna spatiosa julian* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/ OM

21.713/. OM-DZUP.

Allotypus female with the following labels: / Allotypus/ Rio Dagua, [Buenaventura, Valle del Cauca], Colombia/ gen[itália]. prep[arada]. [O.] Mielke 2019/ Allotypus *Yanguna spatiosa julian* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/ OM 41.263/. OM-DZUP.

Paratypes: 1 ♂, Río Dagua, Buenaventura, Valle del Cauca, Colombia, OM 41.223 (OM); 1 ♂, Río Aguacatal, Valle del Cauca, Colombia, 2000 m, Fassl leg., OM 34.682 (OM).

Yanguna aspilos* Mabilles & Boulet, 1908, *stat. rest.

Three subspecies.

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia.

Yanguna aspilos aspilos* Mabilles & Boulet, 1908, *stat. rest.

Pls. 40–41, Figs. 3, 6 (♂), 9 (♀) (d, v); Text-Fig. 44 (♀ genitalia)

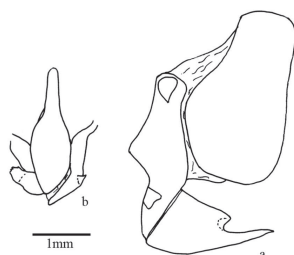
DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley – Quebrada Union 1,600 m*); Junín (*Huancapistana*). **Bolivia:** Cochabamba (*Chapare, Cristal Mayu, Yungas del Palmar*); La Paz (*Caranavi, Farinas, Río Zongo, San Antonio*).

TAXONOMY – This species, here reinstated, is separated from *Yanguna spatiosa* by differences in the male and female genitalia of the three subspecies. In the male genitalia, the harpe is shorter and thicker (compare Fig. 42 (*Y. spatiosa spatiosa*) and Fig. 45 (*Y. aspilos lara ssp. n.*)), and in the female genitalia the sterigma has a shorter dorsal process and a longer and thicker ventral process, and the ventral lobes of the 8th sternite are shorter (only seen in *Y. aspilos aspilos stat. rest.* (compare Fig. 43 (*Yanguna spatiosa spatiosa*)). The forewings are also more rounded. The DNA barcode between *Yanguna spatiosa spatiosa* (n= 6) and *Yanguna aspilos lara ssp. n.* (n= 4), described below, shows a minimum divergence of 2.75%, one more reason for the separation of both species.

TYPE:

Yanguna aspilos Mabilles & Boulet, 1908. The original description mentions an unspecified number of males without data [South America – on the label of the lectotype]. To ensure the correct identification of *Yanguna aspilos*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Amerique du Sud 1906 Herm. Rolle Coll. Boulet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Yanguna aspilos* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.



Text-Fig. 44. *Yanguna aspilos aspilos stat. rest.* Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (Bolivia, Cochabamba, Chapare, Cristal Mayu, X-1976, Steinbach leg., OM 34.604).

Yanguna aspilos lara* O. Mielke, Brockmann & C. Mielke, *ssp. n.

Pls. 40–41, Figs. 7 (holotype ♂), 8 (allotype ♀) (d, v); Text-Fig. 45 (♂ genitalia)

DISTRIBUTION – Central-eastern to southeastern Peru.

Peru: Cusco (*Quillabamba*); Huánuco (*Tingo Maria*); San Martín.

DESCRIPTION – Male: forewing length 31 mm. Head, body and wings very similar to *Y. spatiosa spatiosa*, differing by the more rounded fore and hindwings, as in the other subspecies of *Y. aspilos stat. rest.*, by the light orange colour of both wings dorsally, and by the presence of a large light orange band (nearly 4 mm wide) at the base of the ventral hindwing, from the humeral area, crossing the discal cell, and until the extreme base of CuA₁–CuA₂ and adjacent area in upper CuA₂–2A; this band is larger and more compact than in *Y. spatiosa julian ssp. n.* The DNA barcode shows a minimum divergence of 2.75% between *Y. spatiosa spatiosa* (n= 6) and *Y. aspilos lara ssp. n.* (n= 4).

Female: the only known female with a forewing length of 36 mm and is similar to the male, but the wings are larger and rounder.

ETYMOLOGY – The specific name is in honour of Lara Mielke Merlin, granddaughter of Olaf H. H. Mielke and niece of Carlos G. C. Mielke.

TYPES:

Holotype male with the following labels: / Holotypus/ X-1973 Tingo Maria, Huánuco, Peru/ gen[itália]. prep[arada]. [O.] Mielke 1990/ OM 21.711/ Holotypus *Yanguna aspilos lara* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det., 2019/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Peru Dept. Huanuco, Umgeb. Tingo Maria II-2017 Chaupiyunca ex coll. M. Büche 1800-2200 m/ Allotypus *Yanguna aspilos lara* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/. MSFR.

Paratypes: 1 ♂, X-1986, San Martín, Peru, OM 26.833 (OM); 2 ♂, 1992, Tingo Maria, Huánuco, Peru, 650 m, ex coll. Rojas Villegas, OM 58.365, OM 58.482 (OM); 2 ♂, Tingo Maria, Huánuco, Peru, Villegas leg., HESP-EB 01 098, HESP-EB 01 100 (EB); 2 ♂, 1992, Tingo Maria, Huánuco, Peru, Villegas leg., HESP-EB 01 097, HESP-EB 01 099 (EB); 4 ♂, VI-2015, Tingo Maria, Huánuco, Peru, 1500-2000 m, ex coll. M. Büche (EB); 1 ♂, XII-2002, 1 ♀, I-2003, 4 ♂, 2003, 2 ♂, IX-2014, 1 ♂, XII-2007, 1 ♂, I-2018, Tingo Maria, Huánuco, Peru (CJU); 1 ♂, Quillabamba, Cusco, Peru, III-2000 (CJU).



Text-Fig. 45. *Yanguna aspilos lara ssp. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) gnathos and uncus, ventral view; c) tegumen, saccus, uncus and gnathos, lateral left view; d) right valva, internal view; e) aedeagus, lateral left view; f) aedeagus, lateral right view; g) aedeagus, dorsal view; h) aedeagus, ventral view (holotype).

Yanguna aspilos mabillei* H. H. Druce, 1909, *comb. n.

Pls. 40–41, Figs. 11 (♂) (d, v)

DISTRIBUTION – Central-eastern Peru.

Peru: Junín (*San Ramón*); Pasco (*Huancabamba*).

TAXONOMY – This subspecies, considered until now a

subspecies of *Yanguna spatiosa*, has the same male genitalia as *Yanguna aspilos* **stat. rest.**, resulting in this the new combination.

TYPE:

Yanguna mabiliei H. H. Druce, 1909. The original description mentions an unspecified number of males from Huancabamba, Peru, 6000-10000 ft., Boettger leg. in collection Druce. To ensure the correct identification of *Yanguna mabiliei*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Huancabamba, [Pasco] E. Peru 6000-10,000 ft. Boettger/ *Yanguna mabiliei* H. H. Druce Type Anns. Mag. 1900/; two other labels will be added: / Lectotypus/ Lectotypus *Yanguna mabiliei* H. H. Druce, 1909, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

APYRROTHRIXINA Grishin, 2019

Seven genera.

TAXONOMY – The subtribe is characterized by the genome; no morphological characters are known (Grishin *in* Zhang *et al.* 2019).

Creonpyge O. Mielke, 2002

One species.

DISTRIBUTION – Honduras, Costa Rica to Colombia.

Creonpyge creon (H. Druce, 1874)

Three subspecies.

DISTRIBUTION – Honduras, Costa Rica to Colombia.

Creonpyge creon creon (H. Druce, 1874)

Pls. 40–41, Figs. 13 (♂), 14 (♀) (d, v)

Syn.: *Pyrrhopyge* [**sic**] *cyclops* Staudinger, 1876

DISTRIBUTION – Honduras, Costa Rica to Colombia.

Honduras. Costa Rica: Alajuela (*Poacito*); Cartago (*Tres Ríos, Turrialba*); Guanacaste (*Área de Conservación Guanacaste, San Jerónimo*); Heredia (*Volcán Barba*); Puntarenas (*Monteverde*); San José (*Canada*). **Panama:** Chiriquí (*Boquete, Cerro Colorado, Finca Suiza, Santa Clara*); Coclé (*El Cope*); Veraguas (*Calobre, Santa Fe*). **Colombia.**

The citation **Venezuela** (Kirby 1877) is probably incorrect (Orellana [2010]).

TYPES:

Pyrrhopyga [**sic**] *creon* H. Druce, 1874. The original description mentions an unspecified number of specimens from Calobre, Veraguas, [Panama], Arcé leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyga creon*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Calobre. [Veraguas] Panama. Arce./ B.C.A.Lep.Rhop. *Pyrrhopyge creon*, Druce./ *P. creon* H. Druce Type/ Sp. figured. Type/ Godman-Salvin Coll.1912.-23./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga creon* H. Druce, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge [**sic**] *cyclops* Staudinger, 1876. The original description mentions an unspecified number of males and females from Chiriquí, Panama, Ribbe leg. in collection Staudinger. To ensure the correct identification of *Pyrrhopyge cyclops*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Chiriqui [Panama]/ Coll. Staudinger/ = type of *P. creon*. Druce/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge cyclops* Staudinger, 1876, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Another syntype female is herein designated as paralectotype; this specimen with the following labels: / Origin/ Chiriqui [Panama] Ribbe/ *Cyclops* Stgr/ *Creon* Druce *Cyclops* Stgr./ Coll. Staudinger/; two

other labels will be added: / Paralectotypus/ Paralectotypus *Pyrrhopyge cyclops* Staudinger, 1876, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Creonpyge creon taylori (Nicolay & Small, 1981)

Pls. 40–41, Figs. 10 (paratype ♂), 12 (♀) (d, v)

DISTRIBUTION – Costa Rica.

Costa Rica: Puntarenas (*San Vito – Finca Las Cruces*).

TYPE:

Pyrrhopyge creon taylori Nicolay & Small, 1981. Holotype male from 23-VI-1976, Finca Las Cruces, San Vito, Puntarenas, Costa Rica, 1150 m, G. B. Small Jr. leg. AME, transferred to MGCL.

Creonpyge creon lilliana (Nicolay & Small, 1969)

Pls. 40–41, Figs. 15 (paratype ♂), 16 (♀) (d, v)

DISTRIBUTION – Panama.

Panama: Coclé (*La Mesa*); Panamá Oeste (*Cerro Campana*).

TYPE:

Pyrrhopyge creon lilliana Nicolay & Small, 1969. Holotype male with the following labels: / Cerro Campana, Panama, Panama, 27 Nov. '64 G. B. Small/ *Pyrrhopyge creon lilliana* Nicolay & Small Holotype ♂/ photo/. USNM.

Jonaspyge O. Mielke, 2002

Two species.

DISTRIBUTION – Central Mexico to Nicaragua.

Jonaspyge jonas (C. Felder & R. Felder, 1859)

Pls. 42–43, Figs. 1, 4 (♂), 2–3 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *cydonia* H. Druce, 1874

DISTRIBUTION – Central Mexico to Nicaragua.

Mexico: Chiapas (*Buenavista, Coapilla, Comitán de Domínguez – Santa Rosa, Lagos Tisox, Ocozingo, San Antonio, Soconusco*); Hidalgo (*Lontla*); Morelos (*Yautepec*); Oaxaca (*Comaltepec, La Esperanza, Metates, Mixe, Sierra de Juárez, Santiago Zacatepec, Sierra de Mazateca, Talea de Castro, Vista Hermosa*); Veracruz (*Banderilla, Catemaco, Córdoba, Cuesta de Misantla, Fortín de las Flores, Los Tuxtlas, San Juan Seco de Valencia*). **Guatemala:** Alta Verapaz (*San Cristóbal*); Baja Verapaz (*San Jerónimo*); Quetzaltenango (*Volcán de Santa María*). **El Salvador? Nicaragua:** Managua (*Bluefields*).

TYPES:

Pyrrhopyga [**sic**] *jonas* C. Felder & R. Felder, 1859. The original description mentions an unspecified number of females from Mexico in collection C. Felder & R. Felder. To ensure the correct identification of *Pyrrhopyga jonas*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Mexico type/ 3/ *Jonas*. n./ Felder Coll^N Rothschild Bequest B.M. 1939-1/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga jonas* C. Felder & R. Felder, 1859, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [**sic**] *cydonia* H. Druce, 1874. The original description mentions an unspecified number of specimens from Oaxaca, Mexico, Fenochio leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyge cydonia*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Oaxaca, S. Mexico. Fenochio/ Compared with Plötz's drawings of *jonas*, Feld./ *P. Cydonia* H. Druce type/ Sp. figured/ B.C.Amer. Lep.Rhop. *Pyrrhopyge jonas*, Feld./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga cydonia* H. Druce, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Jonaspyge elizabethae Gallardo & Grishin, 2021

Pls. 42–43, Figs. 5 (♂) (d, v)

DISTRIBUTION – Honduras.

Honduras: La Paz (*Opatoro*).

TYPE:

Jonaspyge elizabethae Gallardo & Grishin, 2021. Holotype male with the following labels: / HOLOTYPE ♂ *Jonaspyge elizabethae* HONDURAS: La Paz Dpt., Municipality of Opatoro, 11.4 air mi E of Marcala, along road V-493, el. 2000 m, 14°08'35.1"N, 87°51'56.8"W, 29 March 2018, leg. Robert J. Gallardo/ DNA sample ID: NVG-18038D01, c/o Nick V. Grishin/ genitalia NVG200302-01, Nick V. Grishin USNM.

Jonaspyge tzotzili (Freeman, 1969)

Pls. 42–43, Figs. 6 (holotype ♀) (d, v)

DISTRIBUTION – Southern Mexico.

Mexico: Chiapas (*Ocozingo, Comitán de Domínguez – Santa Rosa*); Oaxaca.

TYPE:

Pyrrhopyge tzotzili Freeman, 1969. Holotype female with the following labels: / Ocozingo, Chis. [Chiapas] Mex.[ico] VII-42 T. Escalante/ Holotype ♀ *Pyrrhopyge tzotzili* Freeman/. USNM.

Apyrrothrix Lindsey, 1921

Two species.

DISTRIBUTION – USA to Honduras.

Apyrrothrix arizonae (Godman & Salvin, 1893)

Pls. 42–43, Figs. 7, 9 (♂), 8, 10 (♀) (d, v)

DISTRIBUTION – USA to northern Mexico.

USA: Arizona (*Baboquivari Mountains, Chiricahua Mountains, Cochise Co., Elgin, Graham Co. – Fort Grant, Huachuca Mountains, Paradise, Pima Co., Pinal Co., Prescott, Santa Cruz Co., Santa Rita Mountains, Tucacaori Mountains*); New Mexico (*Hidalgo*); Texas (*Marfa*). **Mexico:** Chihuahua (*Yepachi*); San Luis Potosí (*Huasteca Potosina, Sierra de Alvarez*); Sonora (*Bacanora, Cananea, El Palmito, Yécora*); Tamaulipas (*Mante, Tampico, Victoria*).

TYPE:

Pyrrhopyge arizonae Godman & Salvin, 1893. The original description mentions an unspecified number of males and females from Fort Grant, Arizona, [USA], Morrison leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyge arizonae*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H. T/ Fort Grant, Arizona. [USA] Morrison./ Godman-Salvin Coll. 1912.-23 B.C.A.Lep.Rhop. *Pyrrhopyge arizonae*. G. & S./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge arizonae* Godman & Salvin, 1893, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Apyrrothrix araxes (Hewitson, 1867)

Two subspecies.

DISTRIBUTION – Mexico to Honduras.

Apyrrothrix araxes araxes (Hewitson, 1867)

Pls. 42–43, Figs. 11, 15 (♂), 12, 13–14, 16 (♀) (d, v)

DISTRIBUTION – Central-northern Mexico to Honduras.

Mexico: Colima; Distrito Federal (*Mexico, Tacuba, Valle de Mexico*); Durango (*Milpas, San Dimas*); Guanajuato (*Sierra de Santa*

Rosa); Guerrero (*Rincón, Tierra Colorada, Xucumanatlán*); Hidalgo (*Cuesta Colorado*); Jalisco (*Atemajac de Brizuela, Mixtlán, Nextipac, Tecolotlán, Tizapán*); Mexico (*Malinalco, Tenancingo*); Michoacán (*Uruapan*); Morelos (*Ahuatepec, Cuernavaca, Sierra de Huautla, Tepoztlán*); Nayarit (*Banderas Bay, El Refilión, Jasmine, Rancho La Noria, Santa Isabel, Sierra Del Nayar, Tepic, Venustiano Carranza, Xalisco*); Nuevo León; Oaxaca (*Putla, San Andrés, Teotitlán del Valle, Díaz Ordaz*); Puebla (*Puebla, Valle de Puebla*); Sinaloa (*Concordia, Loberas*); Tamaulipas (*Tampico, Victoria*); Sonora (*Yécora*); Veracruz (*Catemaco, Córdoba, Fortín, Los Tuxtlas*). **Honduras. El Salvador?**

TYPE:

Erycides araxes Hewitson, 1867. The original description mentions an unspecified number of specimens from Mexico in collection Hewitson. To ensure the correct identification of *Erycides araxes*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Mexique/ Mexico Hewitson Coll. 79.-69. *Myscelus araxes*. 1./; two other labels will be added: / Lectotypus/ Lectotypus *Erycides araxes* Hewitson, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Apyrrothrix araxes cyrillus (Plötz, 1879), *stat. rev.*

Pls. 42–43, Figs. 17 (holotype ♂), 18 (♀) (d, v)

DISTRIBUTION – Southwestern Mexico.

Mexico: Oaxaca (*Díaz Ordaz, Putla, San Andrés, Teotitlán del Valle*).

TAXONOMY – The holotype of *Pyrrhopyga* [**sic**] *cyrillus* and another specimen female, both from the same locality (Oaxaca, Mexico) and collector in MfN, have a different phenotype from nominotypical subspecies, but a similar DNA barcode (*A. araxes araxes* (n= 5) and *A. araxes cyrillus* *stat. rev.* (n= 2)). The minimum divergence between a specimen of *A. araxes araxes* from Durango, Mexico and the female of *A. araxes cyrillus* *stat. rev.* is only 0.7%. These specimens present a ventral hindwing with a very thin lighter dark border at the external margin than *A. araxes araxes*, similarly to other specimens from the above mentioned localities in Oaxaca. As this phenotype is only known from Oaxaca, the taxon is revalidated as a subspecies of *A. araxes*: *Apyrrothrix araxes cyrillus* *stat. rev.* Specimens are also figured in the “Butterflies of America” website (Warren *et al.* 2020).

TYPE:

Pyrrhopyga [**sic**] *cyrillus* Plötz, 1879. Holotype male (one specimen, number 4926 is mentioned) with the following labels: / Holotypus/ 4926/ Oaxaca [Mexico] Deppe./ *Cyrillus* Plötz */ *Cyrillus* Plötz. Stett. Ent. Zeitschr. 1879 p. 529/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga cyrillus* Plötz, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Chalypge O. Mielke, 2002

Three species.

DISTRIBUTION – Mexico to central-eastern Bolivia.

Chalypge chalybea (Scudder, 1872)

Two subspecies.

DISTRIBUTION – Central-western to southwestern and central-northern Mexico to Honduras.

Chalypge chalybea chalybea (Scudder, 1872)

Pls. 42–43, Figs. 19, 21 (♂), 20 (♀) (d, v)

DISTRIBUTION – Central-western to southwestern Mexico.

Mexico: Aguascalientes; Guanajuato (*Salamanca*); Jalisco (*Guadalajara, Huentitán, La Venta del Astillero, Tlajomulco*); Guerrero (*Orizaba*); Oaxaca (*Candelaria Loxicha, Sierra de Juárez*); Querétaro.

TYPE:

Pyrrhopyga [**sic**] *chalybea* Scudder, 1872. The original description

mentions an unspecified number of specimens from Mexico. To ensure the correct identification of *Pyrrhopyga chalybea*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / M. C. Z. Type 16587/ Mexico/ *Pyrrhopyga chalybea* Scudd. Type/ 1187/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga chalybea* Scudder, 1872, O. Mielke, Brockmann & C. Mielke det. 2019/. MCZ.

***Chalpyge chalybea chloris* (Evans, 1951)**

Pls. 42–43, Figs. 22, 24 (♀), 23 (♂) (d, v)

DISTRIBUTION – Central-northern Mexico to Honduras.

Mexico: Chiapas (*Comitán de Domínguez – Santa Rosa, Ocuilapa de Juárez, San Cristóbal de la Casas, San Antonio Buenavista, San Jerónimo*); Colima (*Comala, San Jerónimo, Suchitlán*); Distrito Federal (*Mexico*); Durango (*Ventanas*); Guerrero (*Acahuizotla, Mescal*); Jalisco (*Ahuacapán, Ajijic, Barranca de Huentitán, Boca de Tomatlán, La Calera, Mismaloya, Puerto Vallarta, Tuxcacuesco*); Mexico (*Malinalco, Toluca*); Michoacán (*Morelia, Pedernales, Uruapan*); Morelos (*Ayala, Chamilpa, Chiconcuac, Cocoyoc, Cuautla, Cuernavaca, Las Estacas, Progreso, Puente de Ixtla, Rancho Viejo, Sierra de Huatla, Tepoztlán, Xochitepec, Yautepec*); Nayarit (*Bucerías, Compostela, Jumatán, La Bajada, La Yerba, Mecatán*); Oaxaca (*Candelaria Loxicha, Putla, San Miguel del Puerto, Sierra de Juárez*); Puebla (*Atlixco, Huiluco, La Galarza, Matamoros, Puebla*); San Luis Potosí (*San Nicolás Tolentino*); Sinaloa (*Chirimollos, Copala, Culiacán, Mazatlán*); Veracruz (*Los Tuxtlas, Misantla, Orizaba*). **Guatemala. Honduras:** Francisco Morazán (*Zamorano*). **El Salvador?**

TYPE:

Pyrrhopyga chalybea chloris Evans, 1951. Holotype male with the following labels: / Type – *chloris* Evans/ Mexico. Hewitson Coll. 79.-69/ *Pyrrhopyga zereda*. 2/ Original of fig. of *Zereda* in Ex. Butt. IV/. NHMUK.

***Chalpyge zereda* (Hewitson, 1866)**

Five subspecies.

DISTRIBUTION – Western and central Colombia to western Ecuador and eastern Ecuador to southeastern Peru.

***Chalpyge zereda zereda* (Hewitson, 1866)**

Pls. 44–45, Figs. 5 (♂) (d, v)

DISTRIBUTION – Ecuador to southeastern Peru.

Ecuador. Peru: Cusco (*Cosñipata Valley – Quebrada Santa Isabel 1,200 to San Pedro 1,500 m*); Huánuco (*Chinchao, Río Monzón, Tingo Maria*); San Martín.

The citations **Mexico** (Hewitson 1871, Plötz 1879) and **Colombia:** Cundinamarca (*Bogotá*) (Kirby 1871) are probably incorrect.

TYPE:

Pyrrhopyga [sic] zereda Hewitson, 1866. The original description mentions an unspecified number of specimens from Ecuador in collection Hewitson. To ensure the correct identification of *Pyrrhopyga zereda*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Ecuador Hewitson Coll. 79.-69. *Pyrrhopyga hygieia*. 1./ *zereda* Hew Type/ type of original description not of figure/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga zereda* Hewitson, 1866. O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Chalpyge zereda hygieia* (C. Felder & R. Felder, 1867)**

Pls. 44–45, Figs. 1, 3 (♂), 2 (neotype ♂ of *Pyrrhopyge phoebus*), 4 (♀) (d, v)

Syn.: *Pyrrhopyga [sic] bogotana* Reakirt, 1868
Pyrrhopyge phoebus Röber, 1925

DISTRIBUTION – Western and central Colombia to western Ecuador.

Colombia: Boyacá (*Muzo*); Caldas (*Manizales – Quebrada El Águila, Manizales*); Cauca; Cundinamarca (*Bogotá – Llanos de San Martín, Fusagasugá*); Risaralda (*Montezuma, Pereira*); Valle del Cauca (*El Darién – Calima*). **Ecuador:** Esmeraldas (*Chuchuvi*); Imbabura (*Lita, Paramba*).

TYPES:

Tamyris hygieia C. Felder & R. Felder, 1867. The original description mentions an unspecified number of specimens from Bogotá, [Cundinamarca] Nova Granada [Colombia], Lindig leg. in collection Felder. To ensure the correct identification of *Tamyris hygieia*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type; Bogota [Cundinamarca, Colombia] Lindig type/ *Hygieia* n./ Felder Collⁿ/ Rothschild Bequest B.M.1939-1./; two other labels will be added: / Lectotypus/ Lectotypus *Tamyris hygieia* C. Felder & R. Felder, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [sic] bogotana Reakirt, 1868. The original description mentions an unspecified number of specimens from Fusagasugá (as Insagasugá), [Cundinamarca], New Grenada [Colombia] in collection Reakirt. To ensure the correct identification of *Pyrrhopyge bogotana*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / *Pyr. Bogotana* Reak. N. Granada Orig. Coll. Reakirt Type/ *P. bogotana*, Reak. is = *hygieia*, Fldr. according to – Seitz-5,839. [?] A./ 5/ Lepidoptera Type Photograph No. 140 Field Museum/ FMNH-INS 0000 095 202/ Photographed A[?] e Stone KE EM[?] catalog/ DNA sample ID: NVG-15039C01 c/o Nick V. Grishin/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga bogotana* Reakirt, 1868, O. Mielke, Brockmann & C. Mielke det 2014/. FMNH.

Pyrrhopyge phoebus Röber, 1925. Holotype male or female from Cauca Valley, Colombia in collection Röber. The holotype was not found in SMT or ZfB, where the Röber's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Pyrrhopyge phoebus*, mentioned as a synonym of *Pyrrhopyge hygieia hygieia* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Calima, Valle del Cauca, Colombia/ Neotypus *Pyrrhopyge phoebus* Röber, 1925 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 27.219/. OM-DZUP.

***Chalpyge zereda rufipectus* (Godman & Salvin, 1879)**

Pls. 44–45, Figs. 6 (♂) (d, v)

DISTRIBUTION – Western Colombia and western and eastern Ecuador.

Colombia: Cauca (*Popayán*). **Ecuador:** Chimborazo (*Riobamba*); Cotopaxi (*Angamarca*); Guayas (*El Salado, Guayaquil*); Napo (*Río Napo*); Pastaza (*Abitagua, Mera, Río Pastaza, Sarayacu*); Tungurahua (*Ambato, Baños, Baños to Canelos, El Topo, Río Topo*).

TYPE:

Pyrrhopyga [sic] rufipectus Godman & Salvin, 1879. The original description mentions an unspecified number of specimens from Río Topo, Ecuador, Buckley leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyga rufipectus*, one syntype male is **herein designated as the lectotype**, this specimen with the following labels: / Type/ Río Topo, [Tungurahua] Ecuador. C. Buckley. Druce Coll./ *Pyrrhopyge rufipectus*, G. – S. Type/ Godman-Salvin Coll. 1912.-23./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga rufipectus* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK

***Chalpyge zereda zepha* (Evans, 1951)**

Pls. 44–45, Figs. 7 (holotype ♂) (d, v)

DISTRIBUTION – Eastern Ecuador.

Ecuador: Zamora-Chinchi (Zamora).

TYPE:

Pyrrhopyge hygieia zepha Evans, 1951. Holotype male with the following labels: / Type – *zepha* Evans/ Zamora, [Zamora-Chinchipe] Ecuador, 3–4,000 ft. (O. T. Baron)/ Rothschild Bequest B.M.1939-1./ NHMUK.

***Chalypyge zereda insana* (Staudinger, 1876)**

Pls. 44–45, Figs. 8 (lectotype ♂ of *Pyrrhopyge insana*), 9 (♂) (d, v)

Syn.: *Pyrrhopyge fassli* Boullet, 1910

DISTRIBUTION – Eastern Ecuador to central-eastern Peru.

Ecuador: Loja (*Loja*); Morona-Santiago (*Huamboya*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Pedro Ruiz Gallo, Rodríguez de Mendoza, Vista Alegre*); Junín (*Chanchamayo, El Porvenir, La Merced, Río Colorado, Río Perené, San Ramón*); Pasco (*Huancabamba, Oxapampa, Pozuzo*); San Martín (*Juanjui, Rioja – El Afluente 1,300 m to Jorge Chávez 1,450 m, Santa Cruz*).

The citations **Panama:** Chiriquí (Mabille 1903, Mabille & Boullet 1908, Mabille 1912, Draudt 1921), Colombia (Boullet 1910, Mabille 1912, Fassl 1918, Draudt 1921, Apollinar Maria 1928), and **Colombia:** Cauca (Hayward 1947) are probably incorrect.

TYPES:

Pyrrhopyge insana Staudinger, 1876. The original description mentions an unspecified number of males from Peru, Thamm leg. and one specimen without data. To ensure the correct identification of *Pyrrhopyge insana*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Lectotypus/ Origin./ Chanchamayo [Junín] Thamm/ Peru *Insana* Stgr/ Coll. Staudinger/; another label will be added: / Lectotypus *Pyrrhopyge insana* Staudinger, 1876, O. Mielke, Brockmann & C. Mielke det. 2014/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published. Another syntype is herein designated as paralectotype; this specimen with the following labels: / Origin./ Chanchamayo [Junín, Peru] Thamm/ *Insana* Stgr./ Coll. Staudinger/; two other labels will be added: / Paralectotypus/ Paralectotypus *Pyrrhopyge insana* Staudinger, 1876, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Pyrrhopyge fassli Boullet, 1910. The original description mentions an unspecified number of males from Huancabamba, Peru, 1908, Fassl leg., one male, Bolivia, 1904, Staudinger leg., and two males, Colombia in MNHN (collection E. Boullet). To ensure the correct identification of *Pyrrhopyge fassli*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Huancabamba [Pasco] Pérou 1908 A. H. Fassl Coll. Boullet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge fassli* Boullet, 1910, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. Another specimen with similar labels is herein designated as paralectotype; MNHN. The abdomen of the lectotype is undamaged, while that of the paralectotype seems to be dissected. The other specimens were not examined.

***Chalypyge rufinucha* (Godman & Salvin, 1879),**

stat. rest.

Pls. 44–45, Figs. 10 (♂) (d, v)

Syn.: *Pyrrhopyge apollo* Röber, 1925

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco. **Bolivia:** Beni (*Beni*); La Paz (*Caranavi, Chaco, Chulumani, Farinas, Río Suapi, Río Zongo, Yungas*); Santa Cruz (*Estrella*).

TAXONOMY – The male genitalia (valvae) of this species is very different from *Chalypyge zereda*, resulting in the status reinstated. The differences are given by Evans (1951).

TYPES:

Pyrrhopyga [**sic**] *rufinucha* Godman & Salvin, 1879. The original description mentions an unspecified number of specimens from Bolivia,

Buckley leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyga rufinucha*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Godman-Salvin Coll. 1912.–23. *Pyrrhopyge rufinucha*, G. & S./ Bolivia. C. Buckley./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga rufinucha* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyge apollo Röber, 1925. The original description mentions an unspecified number of males from Río Zongo, Bolivia, 750 m, Fassl leg. To ensure the correct identification of *Pyrrhopyge apollo*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Syntypus/ Río Songo [**sic**] [La Paz] Bolivia, 750 m. Coll. Fassl/ Syntype ♂ *Pyrrhopyge apollo* Röber, 1925/ *Pyrrhopyge aesculapus*/ Stauding. & Bang-Hass Dresden, Ankauf 1961/ Staatl. Museum für Tierkunde Dresden/ GART, specimen ID: 02547, Exemplar + Etiketten, documentiert, specimen + label, data documented, 2003/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyge apollo* Röber, 1925, O. Mielke, Brockmann & C. Mielke det. 2019/. SMT.

***Aesculapyge* Grishin, 2019**

One species.

DISTRIBUTION – El Salvador to Ecuador.

***Aesculapyge aesculapus* (Staudinger, 1876)**

Pls. 44–45, Figs. 11 (♂), 12 (lectotype ♂ of *Pyrrhopyge aesculapus*), 13 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *variegaticeps* Godman & Salvin, 1879

DISTRIBUTION – El Salvador to Ecuador.

El Salvador: Santa Ana (*Matapan*). **Costa Rica:** Alajuela (*Monte Poás, Poasito*); Cartago (*Asahar de Cartago, Casa Mata, Orosi, Volcán Turrialba*); Guanacaste (*Área de Conservación Guanacaste, San Jerónimo*); Heredia (*Paso Llano, Río Sucio, Volcán Barva*); Puntarenas (*Osa*); San José (*Canada, Guaitil de Pirris, Palmichal*); San Ramón (*Bajo Rodríguez*). **Panama:** Chiriquí (*Bambito, Cerro Colorado, Santa Clara*); Veraguas (*Santa Fe*). **Colombia. Ecuador.**

TYPES:

Pyrrhopyge aesculapus Staudinger, 1876. The original description mentions an unspecified number of specimens from Chiriquí, Panama, Ribbe leg. To ensure the correct identification of *Pyrrhopyge aesculapus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Lectotypus/ Origin./ Chiriqui [Panama] Ribbe/ *aesculapus*/ *Aesculapus* Stdgr *variegatica* G & S./ Coll. Staudinger/; another label will be added: / Lectotypus *Pyrrhopyge aesculapus* Staudinger, 1876, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published. Another two male syntypes are herein designated as paralectotypes; these specimens have the following labels: (1) / Origin./ Chiriqui [Panama] Ribbe/ abgebildet/ Coll. Staudinger/; and (2) / Origin./ Chiriqui Ribbe/ = type of *P. variegaticeps* G & S./ Coll. Staudinger/; two other labels will be added on each: / Paralectotypus/ Paralectotypus *Pyrrhopyge aesculapus* Staudinger, 1876, O. Mielke, Brockmann & C. Mielke det. 2104/. MfN.

Pyrrhopyga [**sic**] *variegaticeps* Godman & Salvin, 1879. The original description mentions an unspecified number of specimens from Costa Rica, Rogers leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyge variegaticeps*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ R. Sucio, [Heredia] Costa Rica. H. Rogers./ Sp. figured/ *Pyrrhopyge variegaticeps*, G. & S. Type/ B.C.Amer. Lep. Rhop. *Pyrrhopyge Aesculapius* [**sic**], Staud./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga variegaticeps* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Cyanopyge O. Mielke, 2002, *stat. rest.*

One species.

DISTRIBUTION – Northern Colombia.

TAXONOMY – This genus was synonymized with *Melanopyge* by Zhang *et al.* (2019), but revalidated here. Although the genomes include some similarities with the species of this genus, *Cyanopyge sangaris* **comb. rest.** remains well segregated from the closely related species of *Melanopyge*. The estimated time of divergence for *Melanopyge* is very similar to that of *Chalpyge* **stat. rest.**, which is also accepted by Zhang *et al.* (2019). In addition to the genome, there are morphological differences that justify revision of its generic status.

Cyanopyge **stat. rest.** is characterized by the following characters. Mesotibia longer than half length of mesofemur; forewing discal cell length about same size of 2A; hindwing with dcs the same size as dcm+dc; hindwing with 2A longer than Rs for a length longer than dcm+dc; external face of mesotibia with a mixture of fine and divergent scales, wider than the tibia. Male genitalia with asymmetric harpe. Female genitalia with sterigma tubular and with a dorsal lobe, apparently closing the ostium as in the *Jonaspyge* and *Aesculapyge* species. Male genitalia are figured by Nicolay & Small (1981) and female genitalia are illustrated below (Fig. 46).

Melanopyge is characterized by the following characters. Mesotibia half the length of mesofemur; forewing discal cell length longer than 2A; hindwing with dcs shorter than dcm+dc; hindwing with dcs shorter than dcm+dc; hindwing with 2A longer than Rs for a length shorter than dcm+dc; external face of mesotibia with a mixture of fine and divergent scales, narrower than the tibia. Male genitalia with symmetric harpe. Female genitalia: sterigma tubular and with a dorsal bifid expansion, not closing the ostium.

Cyanopyge sangaris (Skinner, 1921), **comb. rest.**

Pls. 44–45, Figs. 14 (holotype ♂) (d, v); Pls. 74–75, Figs. 13, 16 (♀) (d, v); Text-Fig. 46 (♀ genitalia)

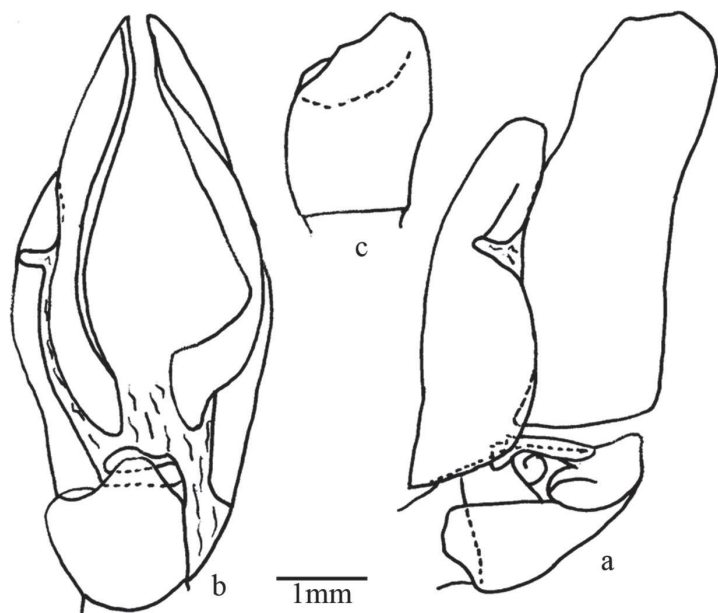
DISTRIBUTION – Northern Colombia.

Colombia: Magdalena (Sierra Nevada de Santa Marta, Cerro Kennedy, Gabali 1,300 m, Onaca, Sierra San Lorenzo – Minca – Hacienda Cincinnati, Valparaiso).

REMARKS – Endemic to the Sierra Nevada de Santa Marta.

TYPE:

Pyrrhopyge sangaris Skinner, 1921. Holotype (one male is mentioned) with the following labels: / Hacienda Cincinnati Sierra San Lorenzo Magdalena, Colombia/ Type No. 7057 *Pyrrhopyge sangaris* Henry Skinner/ 4500 feet (R) VII, 23, 1920/. CMP.



Text-Fig. 46. *Cyanopyge sangaris* **comb. rest.** Female genitalia: a) segment VIII, papilla analis and sterigma, lateral left view; b) segment

VIII, papilla analis and sterigma, posterior view; c) sterigma, ventral view (Colombia, Magdalena, Santa Marta, Onaca, NHMUK).

Melanopyge O. Mielke, 2002

Six species.

DISTRIBUTION – Mexico to western Ecuador and to northwestern Venezuela.

Melanopyge mulleri (Bell, 1934)

Pls. 44–45, Figs. 15 (♂), 16 (♀) (d, v)

Syn.: *Apyrrothrix plancartei* C. Hoffmann, 1941

DISTRIBUTION – Eastern to southern Mexico.

Mexico: Campeche (*Calakmul*); Chiapas; Oaxaca (*Chiltepec*, *Comaltepec*, *Huautla de Jiménez*, *Metates*, *Santa María Yacochi*); Veracruz (*Catemaco*, *Dos Amates*, *Fortín*, *Los Tuxtlas*, *Presidio*).

TYPES:

Apyrrothrix mulleri Bell, 1934. Holotype male with following labels: / *Apyrrothrix mulleri* Bell. Holotype ♂ Slide 4964 U.S.N.M./ Genitalia Slide by E. Bell USNM 90.666 ♂/ R. Muller Collector/. USNM.

Apyrrothrix plancartei C. Hoffmann, 1941. Holotype male with the following labels: / Holotipo/ Ex coll. Toro Estado de Veracruz, [Mexico]/ C. C. Hoffmann det. *Apyrrothrix plancartei* Hffm./ Coll. C. C. Hoffmann Cat. No. 3697 *Apyrrothrix plancartei* C. C. Hoffmann/. AMNH.

Melanopyge hoffmanni (Freeman, 1977)

Pls. 44–45, Figs. 17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Eastern to southern Mexico to Guatemala.

Mexico: Chiapas (*Boca de Chajul*, *Palenque*, *Pichucalco*); Oaxaca (*Chimalapa*, *Comaltepec*, *Sierra de Juárez*, *Tontotepec*); Tabasco (*Tenosique*); Veracruz (*Uxpanapa*). **Belize:** Stann Creek (*Middlesex*); Toledo (*Punta Gorda*). **Guatemala:** Izabal (*Morales – Cayuga*).

TYPE:

Pyrrhopyge hoffmanni Freeman, 1977. Holotype male with the following labels: / Tenosique, Tabasco. Mex[ico]. 3 Sept. 1962 E. C. Welling/ Holotype *Pyrrhopyge hoffmanni* Freeman/. AMNH.

Melanopyge maculosa (Hewitson, 1866)

Pls. 44–45, Figs. 19 (♂), 20 (♀) (d, v)

Syn.: *Tamyris agathon* C. Felder & R. Felder, 1867

DISTRIBUTION – Panama to western Ecuador and to northwestern Venezuela.

Panama: Colón (*Gamboa*, *Gatun*, *Parque Nacional Soberanía*, *Piña*, *Río Chagres*); Panamá (*Madden Forest*, *Summit*); Panamá Oeste (*Cerro Campana*). **Colombia:** Boyacá (*Muzo*, *Otanche*); Caldas (*Victoria*); Cauca; Chocó (San José del Palmar); Cundinamarca (*Bogotá*); Santander (Girón); Valle del Cauca (*Buenaventura – Río Dagua*). **Ecuador:** Esmeraldas (*Chuchuvi*); Imbabura (*Paramba*). **Venezuela:** Zulia (*El Tucuco*).

TYPES:

Pyrrhopyga [*sic*] *maculosa* Hewitson, 1866. The original description mentions an unspecified number of specimens from Bogotá [Colombia] in collection Hewitson. To ensure the correct identification of *Pyrrhopyga maculosa*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H.42/ N. Granada [Colombia] Hewitson Coll. 79.-69. *Pyrrhopyga maculosa*. 2./ 131/ NHMUK(E) #982992/ *Pyrrhopyge maculosa maculosa*; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga maculosa* Hewitson,

1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Tamyris agathon C. Felder & R. Felder, 1867. The original description mentions an unspecified number of males and females from Bogotá, Nova Granada [Colombia], Lindig leg. in collection Felder. To ensure the correct identification of *Tamyris agathon*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type – *agathon* Felder/ Bogota [Cundinamarca, Colombia] Lindig type/ H. 2538. *Tamyris agathon* Felder/ *agathon* Fld. Type/ Rothschild Bequest B.M. 1939-1./; two other labels will be added: / Lectotypus/ Lectotypus *Tamyris agathon* C. Felder & R. Felder, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Melanopyge erythrosticta* (Godman & Salvin, 1879)**

Pls. 44–45, Figs. 21 (♂), 22 (♀) (d, v)

DISTRIBUTION – Southern Mexico to Panama.

Mexico: Oaxaca. **Guatemala:** El Petén (*Sayaché*). **Honduras.** **Nicaragua:** Chontales (*Santo Domingo*). **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste*). **Panama.**

TYPE:

Pyrrhopyga [*sic*] *erythrosticta* Godman & Salvin, 1879. Lectotype male (Burns & Janzen 2001) with the following labels: / Chontales, Nicaragua. T. Belt./ B.C.A.Lep.Rhop. *Pyrrhopyge erythrosticta*, G. & S./ Godman-Salvin Coll. 1912.-123./ Lectotype *Pyrrhopyga erythrosticta* Godman & Salvin ♂ John M. Burns 2001/. NHMUK.

Melanopyge burnsi* O. Mielke, Brockmann & C. Mielke, *sp. n.

Pls. 44–45, Figs. 23 (holotype ♂), 24 (allotype ♀) (d, v); Text-Figs. 47 (♂ genitalia), 48 (♀ genitalia)

DISTRIBUTION – Southern Mexico to Panama.

Mexico: Oaxaca (*Chimalapa, Tuxtepec*). **Belize:** Stann Creek (*Middlesex*); Toledo (*Punta Gorda*). **Honduras:** Colón (*Trujillo*). **Nicaragua.** **Costa Rica:** Cartago (*Turrialba*); Guanacaste (*Área de Conservación Guanacaste*); Puntarenas (*Corcovado*); San José (*Cantón de Mora*). **Panama:** Chiriquí (*Lino*).

These localities follow “Butterflies of America” (BoA) and the type material data.

DESCRIPTION – Male: vertex, frons, palpi, genae, thorax, abdomen dorsally and ventrally, as well as all wing ornamentation, as those of *M. erythrosticta*, *M. hoffmanni* and *M. maculosa*. Forewing length 24 mm (n= 1). Forewing dorsally with the red spot at the base in CuA₂–2A as in *M. erythrosticta* and *M. hoffmanni*; semi-hyaline white spots as in *M. maculosa*, so the spot in the upper half of CuA₂–2A not below the spot in CuA₁–CuA₂ but rather slightly shifted towards outer margin, thus the spots in the disc cell, in CuA₁– CuA₂ and the upper one in CuA₂–2A arranged in a straight line and not as in *M. erythrosticta* and *M. hoffmanni* where the spot in the upper half of CuA₂–2A is below the spot in CuA₁–CuA₂; whitish fringes between M₂–2A and of general colour at the veins. Hindwing with white fringes and ground colour at the veins. Forewing ventrally with basal yellow similar to the three mentioned species and fringes as dorsally. Hindwing ventrally with basal yellow 2/3 of the width of the wing, forming an external margin of the ground colour of about 4 mm; fringes white and of ground colour at the veins as in *M. maculosa*. Genitalia: apophyses of the tegumen almost reaching the distal tip of the uncus, the latter wide, narrowing abruptly and ending as a bifid lobe; valva with approximately straight harpe gradually tapering towards the tip, dorsal margin sinuous and weakly serrated (strongly sinuous and serrated in *M. maculosa*), dorso-proximal projection straight, short, not reaching the dorsal limit of the valva, this projection in the right valva narrowing to the tip while in the left valva slightly dilated at the tip (*M. erythrosticta* and *M. hoffmanni* with projection long, reaching the dorsal limit of the valva); sacculus approximately rectangular and not projected beyond the dorso-proximal projection of the harpe as in *M. maculosa* (*M. erythrosticta* and *M. hoffmanni* with

sacculus projected beyond the dorso-proximal projection of the harpe and almost reaching its distal tip).

Female: very similar to the male. Forewing length 31 mm (n= 1). The hindwing ventral basal yellow narrower than in male, occupying the basal half to 2/3 of the wing. Genitalia: Eighth tergite elongated, ventrally fused to sterigma; the latter tubular, slightly longer on the right side and with a bilobed dorsal process with anterior and posterior projections, while in *M. maculosa* the sterigma similar, but this process only with posterior projection.

TAXONOMY – Species very similar to *M. maculosa* due to the arrangement of the white semi-hyaline spots on the forewing; those in the disc cell, in CuA1–CuA2 and in the upper part of CuA2–2A in a straight line, but distinguished immediately by the presence of a red spot on the forewing dorsally (absent in *M. maculosa*). In addition, it differs by the male genitalia; harpe approximately straight, tapering to the tip with dorsal margin slightly sinuous and slightly serrated and without dorso-central expansion (*M. maculosa* has a wider harpe, slightly sinuous and dorsal margin more sinuous and more strongly serrated and with dorso-central expansion folded inwards).

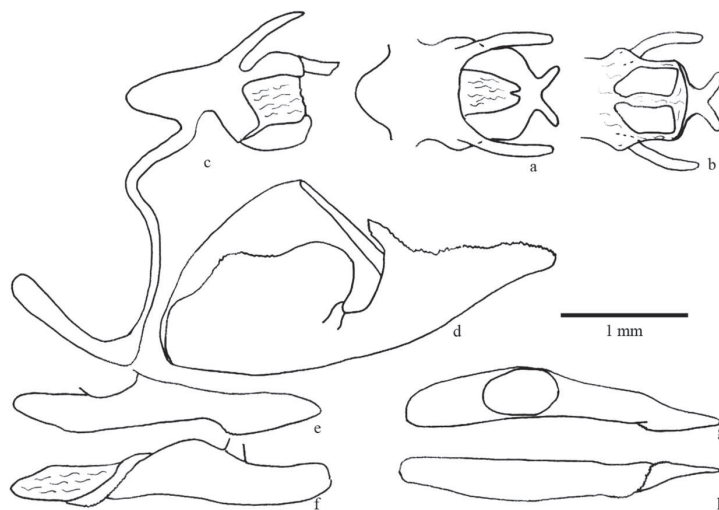
ETYMOLOGY – The species is a dedication to our friend John Burns, United States National Museum, Smithsonian Institution, Washington, DC, USA, for many years of collaboration.

TYPES:

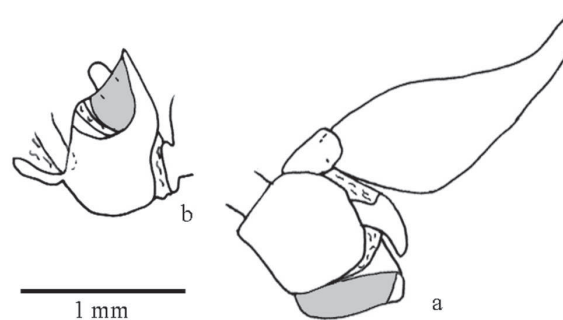
Holotype male with the following labels: / Holotypus/ Chiriqui Panamá/ Gen[itália]. Prep[arada]. [O.] Mielke 2019/ Holotypus *Melanopyge burnsi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/ OM 66.050/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ Chiriqui Panamá/ Gen[itália]. Prep[arada]. [O.] Mielke 2019/ Allotypus *Melanopyge burnsi* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2019/. OM 66.057/. OM-DZUP.

Paratype: 1 ♂, 4-III-2014, Corcovado, Osa, Puntarenas, Costa Rica (EB).



Text-Fig. 47. *Melanopyge burnsi* *sp. n.* Male genitalia: a) tegumen and uncus, dorsal view; b) apophyses of tegumen, uncus and gnathos, ventral view; c) tegumen, saccus, uncus and gnathos, lateral left view; d) right valva, internal view; e) aedeagus, lateral left view; f) aedeagus, lateral right view; g) aedeagus, dorsal view; h) aedeagus, ventral view (holotype).



Text-Fig. 48. *Melanopyge burnsi* *sp. n.* Female genitalia: a) segment VIII and sterigma, lateral left view; b) sterigma, ventral view (allotype).

Melanopyge cossea (H. Druce, 1875)

Pls. 46–47, Figs. 1 (♂) (d, v)

DISTRIBUTION – Central Colombia.

Colombia: Boyacá (*Otanche*); Cundinamarca (*Paima*).

TYPE:

Pyrrhopyga [*sic*] *cossea* H. Druce, 1875. The original description mentions an unspecified number of specimens from Colombia in collection Druce. To ensure the correct identification of *Pyrrhopyga cossea*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type HT/ Colombia Chesterton./ *Pyrrhopyga cossea* H. Druce. type./ 134/ Godman-Salvin Coll. 1912.-23. *Pyrrhopyge cossea*, Druce./ NHMUK(E) #982993/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga cossea* H. Druce, 1875, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

MIMONIADINA Grishin, 2019

Twelve genera.

TAXONOMY – The subtribe is characterized by the genome; no morphological characters are known (Grishin in Zhang *et al.* 2019).

Ardaris Watson, 1893

Two species.

DISTRIBUTION – Northwestern Venezuela.

TAXONOMY – Based on the genome of *Pyrrhopyginae*, Zhang *et al.* (2019) synonymized *Mimardaris* **stat. rev.**, with *Ardaris*. However, they show that within *Ardaris* there are three distinct groups of species well characterized by morphological differences and by chromatic patterns. As such, *Mimardaris* **stat. rev.** is revalidated and *Aerardaris* **gen. n.** is described for *Pyrrhopyga* [*sic*] *aerata*.

Ardaris, a genus with two included species (*A. eximia* and *A. hantra*), is characterized by the following characters. Mesotibia with a few long and thin scales. Forewing projected. Hindwing with dcs longer than the half of dcm+dcu. Male genitalia: harpe with tip turned dorsally and with expansion pointed towards its base; uncus slightly expanded at its proximal half; fultura inferior not dorsally united. Female genitalia: sterigma small, more sclerotized at the ventral base of the ostium, tubular, and not fused to the lateral portion of the sternite VIII, the latter densely sclerotized only at the inferior portion; dorsal portion of sterigma without distal median projection; lamella antevaginalis not pronounced; without tiny bags (function unknown) at the anterior margins of the lateral portions of the sternite VIII. Forewing with white hyaline to yellowish semi-hyaline spots in discal cell and from the apex to the centre of anal margin. The male and female genitalia are shown in Orellana ([2010]). Characters that distinguish it from *Mimardaris* **stat. rev.** are listed under the latter.

Ardaris eximia (Hewitson, 1871)

Pls. 46–47, Figs. 2 (♂) (d, v); Pls. 74–75, Figs. 12 (♀) (d, v)

DISTRIBUTION – Northwestern Venezuela.

Venezuela: Barinas (*Barinitas*); Lara (*Los Nepes*, *Sierra de Portuguesa*, *Via Sanare* – *Caspo*); Mérida (*Bailiadores*, *Chachopo*, *Cordillera de Mérida*, *La Carbonera*, *Páramo La Culata*, *San Esteban*, *Valle Grande*, *via La Azulita*); Táchira (*Jáurgui*, *Páramo La Negra*); Trujillo (*Road Boconó* – *La Negría*).

TYPE:

Pyrrhopyga [*sic*] *eximia* Hewitson, 1871. The original description mentions an unspecified number of specimens from Venezuela, Göring leg. in collection Hewitson. To ensure the correct identification of *Pyrrhopyga eximia*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H.78/ Venezuela Hewitson Coll. 79.-69 *Erycides eximia*, Hew/ 224/ *Ardaris eximia eximia* Hewitson, 1867/ NHMUK(E) #982963/; two other

labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga eximia* Hewitson, 1871, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Ardaris hantra Evans, 1951

Pls. 46–47, Figs. 3 (♂), 4 (♀) (d, v)

DISTRIBUTION – Northwestern Venezuela.

Venezuela: Mérida (*Alto Albarregas*, *Las Piñuelas*, *Mérida* – *Páramo Santo Domingo* and *Los Frailes*, *Mucubají*, *Mucuchíes*, *Páramo Campanario*, *Páramo La Gavidia*, *Páramo La Culata*, *Parque Nacional Sierra Nevada*, *Sierra de la Culata* – *Río Albarregas* and *Páramo Los Conejos*, *Piñango*, *Valle del Domingo*, *via Piñango* – *Pico El Águila*).

TYPE:

Ardaris eximia hantra Evans, 1951. Holotype male with the following labels: / Type – *hantra* Evans/ 227/ Venezuela E[stado]. Merida. Paramos de Campanario 13,500ft. 18.IX.1938/ J. Hanbury Tracy BM. 1938-117/ Paramos de Campanario Est. Merida Venezuela 12,500ft 13,500 118.9.38 J. H. T 36/ *Ardaris eximia hantra* Evans, 1951/ NHMUK(E) #982962/. NHMUK.

Evans (1951) mentioned Paramos de Companeiro [*sic*], as the type locality.

Aerardaris O. Mielke, Brockmann & C. Mielke, **gen. n.**

DESCRIPTION – *Aerardaris* **gen. n.** is characterized by the following characters. Mesotibia with short scales only. Forewing not projected. Hindwing with vein dcs longer than the half of dcm+dcu. Male genitalia: harpe straight with dorsal expansion bilobed; uncus well expanded at the beginning of its distal half; fultura inferior not dorsally united. Female genitalia: sterigma developed, uniformly sclerotized, tubular, and fused to the lateral portions of the sternite VIII; dorsal portion of sterigma without distal median projection; lamella antevaginalis pronounced, rectangular, connected to the ostium by three fine sclerotizations, two lateral and one median, and with tiny bags (function unknown) at the anterior margins of the lateral portions of the sternite VIII, the latter uniformly sclerotized. Forewing without semi-hyaline or hyaline spots, dorsally with pronounced green reflex. The male genitalia are shown in Nicolay & Small (1981) and the female genitalia are figured here (Fig. 49).

Type species: *Pyrrhopyga* [*sic*] *aerata* Godman & Salvin, 1879, herein designated. The only included species.

DISTRIBUTION – Northern Colombia.

ETYMOLOGY – The generic name is a combination of *aerata* and *Ardaris*.

Aerardaris aerata (Godman & Salvin, 1879), **comb. n.**

Pls. 46–47, Figs. 5 (♂), 6 (♀) (d, v); Pls. 74–75, Figs. 15 (♂) (d, v); Text-Fig. 49 (♀ genitalia)

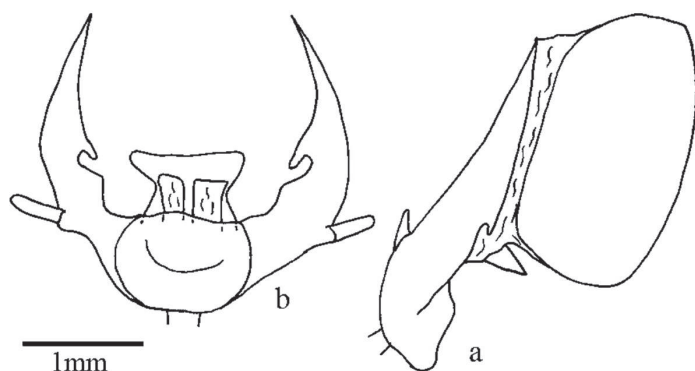
DISTRIBUTION – Northern Colombia.

Colombia: Magdalena (*Caverna*, *Cerro Kennedy* 2,600 m, *El Libano*, *El Mamón*, *Pueblo Viejo*, *San Lorenzo*, *San Pedro de la Sierra*, *San Sebastian de Buenavista*, *Tromba*, *Vista Nieve*). All localities are in the Sierra Nevada de Santa Marta above 2,000 m.

REMARKS – Endemic to the Sierra Nevada de Santa Marta.

TYPE:

Pyrrhopyga [*sic*] *aerata* Godman & Salvin, 1879. Holotype (one specimen is mentioned) male with the following labels: / Type H T/ Pueblo Viejo, [Sierra Nevada de Santa Marta, Magdalena], Colombia. Simons./ *Pyrrhopyga aerata* gen.prep. [O.] Mielke 1976/ Godman-Salvin Coll. 1912.-23. *Pyrrhopyge aerata* G. & S./ B.M.(N.H.) Rhopalocera Slide No. 13013/. B.M.(N.H.) Rhopalocera Slide No. 13013/. NHMUK.



Text-Fig. 49. *Aerardaris aerata* **comb. n.** Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (Colombia, Magdalena, Sierra Nevada de Santa Marta, OM 11.939).

Mimardaris O. Mielke, 2002, **stat. rev.**

Seven species.

DISTRIBUTION – Colombia to central-eastern Bolivia.

TAXONOMY – *Mimardaris* **stat. rev.** was synonymized with *Ardaris* by Zhang *et al.* (2019), but is here revalidated. Although the genomes of the species of *Mimardaris* **stat. rev.** show some similarities with the species of *Ardaris*, the species of *Mimardaris* **stat. rev.** remain well segregated, suggesting the validity of *Mimardaris* **stat. rev.** The estimated time of divergence is similar to that of *Chalypge*, another genus accepted by Zhang *et al.* (2019). In addition to the genome, there are many differences in the morphology of the body and wing patterns between *Ardaris* and *Aerardaris* **gen. n.**

Mimardaris **stat. rev.** is characterized by the following characters (see also the comparative descriptions of *Ardaris* in *Aerardaris* **gen. n.**). Mesotibia with some thin and long scales. Forewing projected. Hindwing with vein dcs shorter than the half of dcm+dcu. Male genitalia: harpe straight with one or two dorsal expansions; uncus well expanded at its proximal half; fultura inferior dorsally united. Female genitalia: sterigma developed, uniformly sclerotized, tubular, and fused to the lateral portion of the sternite VIII, the later uniformly sclerotized; dorsal portion of the sterigma with distal median projection; lamella antevaginalis not pronounced and with tiny bags (function unknown) at the anterior margins of the lateral portions of the sternite VIII. Forewings with three sets of yellowish semi-hyalines spots: one median band from the centre of discal cell to the distal third of CuA₂–2A, two spots overlapping in M₂–M₃ and M₃–CuA₁ and four to five apical spots. Male genitalia of some species shown by Evans (1951) and the female genitalia of *M. periphema* **stat. rest., comb. rev.** figured here (Fig. 50).

Mimardaris pityusa (Hewitson, 1857), **comb. rev.**

Pls. 46–47, Figs. 7 (♂), 8 (♀) (d, v)

Syn.: *Mimoniades aemulus* Skinner, 1920

Mimoniades hemitaenia Röber, 1925

DISTRIBUTION – Western, central and eastern Colombia to central-eastern Bolivia.

Colombia: Antioquia (*Jardín*); Boyacá (*Muzo*); Caquetá (*Florencia*); Cauca; Cordillera Occidental; Cundinamarca (*Bogotá – La Aguadita, Pacho, Zipacón*); Huila (*San José de Isnos – Slopes of Central Cordillera*); Magdalena Valley; Putumayo; Tolima (*Canon de Tolima*); Valle del Cauca. **Ecuador:** Loja (*San Francisco*); Morona-Santiago (*Huamboya, Macas*); Napo (*Baeza*); Pastaza (*Sarayacu*); Tungurahua (*Ambato, Baños, El Rosario, Río Negro, Río San Francisco*); Zamora-Chinchipe (*Zamora*). **Peru:** Cusco (*Cosñipata Valley, Machu Picchu*); Pasco (*Huancabamba, Oxapampa*); Pastaza (*Río Pastaza*); San Martín (*Tocache*). **Bolivia:** Cochabamba (*Cocapata, Locotal, San Jacinto*).

TYPES:

Pyrrhopyga [**sic**] *pityusa* Hewitson, 1857. The original description mentions an unspecified number of females (**recte** males) from New Granada [Colombia] [**recte** Ecuador – on the label of the lectotype]. To ensure the correct identification of *Pyrrhopyga pityusa*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H. 27/ Ecuador Hewitson Coll. 79.–69. *Pyrrhopyga pityusa*. 5./ 217/ NHMUK(E) #982965/ *Mimoniades pityusa* Hewitson, 1870/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga pityusa* Hewitson, 1857, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Mimoniades aemulus Skinner, 1920. Holotype (one male is mentioned) with the following labels: / Ambato [Tungurahua] Ecuador/ Type No. 7054 *Mimoniades aemulus* Henry Skinner/. ANSP, transferred to CMP.

Mimoniades hemitaenia Röber, 1925. Lectotype male (O. Mielke 1989a) with the following labels: / W. Colomb[ia]. Magdal[ena]./ *hemitaenia* Lectotypus *Mimoniades hemitaenia* Röber, 1925 O. Mielke det. 1988/ Staudinger. u. Bang-Haas Dresden Ankauf 1961/ Staatl. Museum für Tierkunde/ GART specimen ID: 02564 Exemplar + Etiketten dokumentiert specimen + label data documented/. SMT.

Mimardaris minthe (Godman & Salvin, 1879), **comb. rev.**

Pls. 46–47, Figs. 9 (♂) (d, v)

Syn.: *Mimoniades angustifascia* Röber, 1925

DISTRIBUTION – Western Colombia to western Ecuador, central and eastern Colombia to central-eastern Peru.

Colombia: Cauca (*Popayán*); Cordillera Oriental; Cundinamarca (*Guayabetal – Susumuco, Medina*); Meta (*Villavicencio*). **Ecuador:** Bolívar (*Balzapamba*); Loja (*Loja, San Francisco*); Morona-Santiago (*Huamboya, Macas*); Napo (*Misahualli, Tena*); Pastaza (*Abitagua, Puyo, Río Pastaza, Sarayacu*); Pichincha (*Quito*); Santo Domingo de los Tsáchilas (*Santo Domingo de los Colorados*); Tungurahua (*Ambato, Baños, Colonia Azuaya, El Topo, Hacienda Santa Inés, Río Negro, Río San Francisco, Río Topo, Río Verde, Río Zuñac*); Zamora-Chinchipe (*Charapi, Zamora*). **Peru:** Amazonas (*Cordillera del Condor – Falso Paquishá*); Junín (*Shimá*); San Martín (*Rioja – Puente Aguas Verdes*).

TYPES:

Pyrrhopyga [**sic**] *minthe* Godman & Salvin, 1879. The original description mentions an unspecified number of specimens from Río Topo, Ecuador, Buckley leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyga minthe*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H. 73/ Río Topo, [Tungurahua] Ecuador. C. Buckley leg. Druce Coll./ Godman-Salvin Coll. 1912.–23. *Mimoniades minthe* G. & S./ 222/ *Mimoniades minthe* Godman & Salvin, 1879/ NHMUK(E) #982964/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga minthe* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Mimoniades angustifascia Röber, 1925. Holotype male with the following labels: / Macas [Morona-Santiago] Ecuador/ *angustifascia* Lectotypus *Mimoniades angustifascia* Röber, 1925 O. Mielke det. 1988/ Staudinger. & Bang Haas Dresden. Ankauf 1967/ Staatl. Museum für Tierkunde Dresden/ GART specimen ID: 02546 Exemplar + Etiketten dokumentiert specimen + label data documented 2003/. SMT.

Mimardaris montra (Evans, 1951), **comb. rev.**

Pls. 46–47, Figs. 10 (♂) (d, v)

DISTRIBUTION – Ecuador to central-eastern Bolivia.

Ecuador. Peru: Cusco (*Aguas Calientes, Callanga, Machu Picchu*); Huánuco (*Tingo Maria*); Junín (*Andamarca, Chanchamayo, Río Colorado, San Ramón – Mina Pichita*); Pasco (*Huancabamba, Pozuzo*). **Bolivia:** Cochabamba (*Cristal Mayu*).

TYPE:

Mimoniades montra Evans, 1951. Holotype male with the

following labels: / Type – *montra* Evans/ Huancabamba, [Pasco] N. E. Peru/ 215/ R. Oberthür Coll. Brit. Mus. 1931-136/ NHMUK(E) #982966/ *Mimoniades montra* Evans, 1951/. NHMUK.

***Mimardaris porus* (Plötz, 1879), comb. rev.**

Two subspecies.

DISTRIBUTION – Northwestern, western, central and eastern Colombia to eastern Peru.

***Mimardaris porus porus* (Plötz, 1879), comb. rev.**

Pls. 46–47, Figs. 11 (♂) (d, v)

Syn.: *Mimoniades punctiger* Mabilles & Boulet, 1908
Mimoniades minthe var. *egena* Mabilles & Boulet, 1908

DISTRIBUTION – Northwestern, western, central and eastern Colombia to eastern Ecuador.

Colombia: Antioquia (*El Retiro, Frontino, Mesopotamia*); Boyacá (*Otanche*); Caldas (*Aguadas, Manizales*); Cauca (*Corinto, El Tombo, Popayán*); Chocó (*San Jose del Palmar*); Risaralda (*Pueblo Rico*); Valle del Cauca (*Aguatal, Cali, Calima, Río Bravo – Buga, Turne*). **Ecuador:** Napo (*El Chaco*).

The citations **Peru** (Mabilles & Boulet 1908) and **Bolivia** (Draudt 1921, Lamas 1969, Lewis 1973, 1975) are incorrect.

TYPES:

Pyrrhopyga [**sic**] *porus* Plötz, 1879. The original description mentions an unspecified number of specimens from Colombia. To ensure the correct identification of *Pyrrhopyga porus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ aff. *Pityusa* Hew. Columbia 76 Däme (Weymer/ Coll. Weymer/ *Porus* Wmr. i. l. Plötz Columbia/ 12: 7./ *Porus* Wmr i l no 246 best. Plötz/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga porus* Plötz, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. MN.

Mimoniades punctiger Mabilles & Boulet, 1908. The original description mentions two males, without data, in MNHN (collection Boulet). To ensure the correct identification of *Mimoniades punctiger*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Pérou 1904 O. Staudinger Coll. Boulet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Mimoniades punctiger* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. The other syntype is herein designated as paralectotype; this specimen with the following labels: / Type/ [illegible] /; two other labels will be added: / Paralectotypus/ Paralectotypus *Mimoniades punctiger* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Mimoniades minthe var. *egena* Mabilles & Boulet, 1908 was mentioned as a synonym of *Mimardaris porus porus* by Evans (1951). The original description mentions two males from Peru in MNHN. These two syntypes were not found in MNHN, or in any museum or collection mentioned above (see “Abbreviations of the collections”). Perhaps a neotype should be designated to ensure the correct identification of the taxon. So far, there is no available specimen from Peru for a neotype designation. This variation doesn’t include the apical spots and the submarginal spots in M_2 – M_3 and M_3 – CuA_1 , and could be a senior synonym of *Mimardaris porus mortis* described from Peru, while *Mimardaris porus porus* is described from Colombia.

***Mimardaris porus mortis* (Evans, 1951), comb. rev.**

Pls. 46–47, Figs. 12 (♂) (d, v); Pls. 74–75, Figs. 17 (♀) (d, v)

DISTRIBUTION – Eastern Peru.

Peru: Amazonas (*Chachapoyas – Yamborasbamba, Molinopampa, Oso Perdido 2,200 m, Rodríguez de Mendoza, Vista Alegre*); Cusco (*Machu Picchu*); Huánuco (*Tingo Maria*); San Martín (*Huayabamba, Puente Aguas Verdes, Rioja – El Afluente 1,200 m to Jorge Chávez 1,450 m, Santa Cruz 1,200 m*).

TYPE:

Mimoniades porus mortis Evans, 1951. The holotype male with the following labels: / Type – *mortis* Evans/ Pérou, Dép. Amazonas (Chachapoyas) M. de Mathan 1889/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Mimardaris sela* (Hewitson, 1866), comb. rev.**

Three subspecies.

DISTRIBUTION – Central and eastern Colombia to central-eastern Peru.

***Mimardaris sela sela* (Hewitson, 1866), comb. rev.**

Pls. 46–47, Figs. 17 (♂), 21 (♀) (d, v)

DISTRIBUTION – Central and eastern Colombia.

Colombia: Boyacá (*Santa María*); Cundinamarca (*Bogotá, Chirajara, Guayabetal – Susumuco*); Meta (*Upper Río Negro, Villavicencio – Pipiral*); Tolima (*Cajamarca*).

The citations **Peru** (Spitz 1932) and **Bolivia** (Mabilles & Boulet 1908) are incorrect.

TYPE:

Pyrrhopyga [**sic**] *sela* Hewitson, 1866. The original description mentions an unspecified number of males from Nova Granada [Colombia]. To ensure the correct identification of *Pyrrhopyga sela*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ N. Granada Hewitson Coll. 79.-69. *Pyrrhopyga sela*. 1./ 210/ NHMUK(E) #982969/ *Mimoniades sela sela* Hewitson, 1866/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga sela* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Mimardaris sela aequatorea* (Röber, 1925), comb. rev.**

Pls. 46–47, Figs. 13 (♂) (d, v); Pls. 74–75, Figs. 14 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Peru.

Ecuador: Loja (*Loja*); Morona-Santiago (*Macas, Sucúa*); Napo (*Puerto Misahuallí*); Pastaza (*El Partidero – Upper Río Anzú, Mera, Río Pastaza, Sarayacu*); Tungurahua (*Hacienda La Mascota, Topo*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Molinopampa, Pongo del Rentema, Rodríguez de Mendoza, Vista Alegre*); Cajamarca; San Martín (*Huayabamba, Rioja, Rioja – San Agustín 1,000 m to Jorge Chávez 1,400 m, Santa Cruz 1,200 m*).

TYPE:

Mimoniades aequatorea Röber, 1925. Holotype (one male is mentioned) with the following labels: / Ecuad[or]. Macas [Morona-Santiago]/ *aequatorea* Holotypus *Mimoniades aequatorea* Röber, 1925 O. Mielke det. 1988/ Stauding. u. Bang-Hass Dresden. Ankauf 1961/ Staatl. Museum für Tierkunde Dresden/ GART specimen ID: 02542 Exemplar + Etiketten dokumentiert specimen + label data documented/. SMT.

***Mimardaris sela peruviana* (Draudt, 1921), comb. rev.**

Pls. 46–47, Figs. 14–15, 18–19 (♂), 22 (♀) (d, v)

Syn.: *Mimoniades pityusa* ab. *chanchamayonis* Strand, 1921, **nom. nud.** (ICZN, Art. 45.6.2)

Mimoniades pityusa form *chanchamayonis* Draudt, 1924, validated (ICZN, Art. 45.6.4), **syn. n.**

DISTRIBUTION – Eastern Peru.

Peru: Amazonas (*Chachapoyas, Huambo*); Huánuco (*Tingo Maria*); Junín (*Andamarca, Chanchamayo, La Merced, Pampa Hermosa, Perené, Río Colorado, San Ramón, Satipo, Shimá*); Pasco (*Pozuzo*); San Martín (*Juanjuí, Mashoyacu, Moyobamba, Rioja – El Afluente 1,300 m to Jorge Chávez 1,450 m, Río Mishollo, Roque*).

TAXONOMY – *Mimoniades pityusa* form *chanchamayonis* was accepted as another subspecies of *Mimardaris sela* **comb. rev.**, but the characters given by Evans (1951) don't distinguish these taxa: "uph [upper hindwing] the discal tawny discal band extends into space 7, but the spot in space 7 is not larger than the spot in space 6 – *peruviana*; uph discal tawny band broadens apically and the spot in space 7 is much larger than the spot in space 6 – *chanchamayonis*." Our large series from Tingo Maria, Huánuco, shows both phenotypes sympatric in this locality. So, *Mimoniades pityusa* form *chanchamayonis* **syn. n.** is here considered as a new synonymy of *Mimoniades peruviana* Draudt, 1921.

The citations **Colombia** (Bell 1934b, Hayward 1947, Bryk 1953) are probably incorrect.

The last locality (*Yungas*) given by Evans (1951) may be from a mislabelled specimen.

TYPES:

Mimoniades pityusa ab. *chanchamayonis* Strand, 1921. Holotype male (one specimen is mentioned) with the following labels: / Typus/ Chanchamayo, [Junín] Peru A. Heyne, Berlin – Wilm./ *Mimoniades pityusa* Hew. ab. *chanchamayonis* m. Strand det./ K. 4/ *pityusa* Hew. ab. *Chanchamayones* Strand/. DEL. The name is a **nomen nudum** (ICZN, Art. 45.6.2).

Mimoniades peruviana Draudt, 1921. Lectotype male (O. Mielke 1993) with the following labels: / Lectotypus/ *peruviana* U/ Lectotypus *Mimoniades peruviana* Draudt, 1921, O. Mielke det. 1979/ Coll. A. Seitz/ 450/ Süd Peru/. MSFR.

Mimoniades sela form *chanchamayonis* Draudt, 1924. As the taxon was validated by Draudt (1924) (ICZN, Art. 45.6.4), the holotype of this taxon is the same holotype of *Mimoniades pityusa* ab. *chanchamayonis* Strand, 1921. **Syn. n.**

Mimardaris lomax (Evans, 1951), **comb. rev.**

Pls. 46–47, Figs. 16 (♂) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley – Pilcopata 500 m to Wayquecha 2,950 m, Marcapata*); Puno (*Oroya – Río Inambari*). **Bolivia:** Cochabamba (*Alto Palmar*).

TYPE:

Mimoniades lomax Evans, 1951. Holotype male with the following labels: / Type - *lomax* Evans/ Marcapata [Cusco] E. Peru, 4500 ft./ Rothschild Bequest B.M.1939-1./ 213/ NHMUK(E) # 982967/ *Mimoniades lomax* Evans, 1951/. NHMUK.

Mimardaris periphema (Hewitson, [1875]), **stat rest., comb. rev.**

Pls. 46–47, Figs. 20 (♂), 23 (♀) (d, v); Text-Fig. 50 (♀ genitalia)

Syn.: *Mimoniades inaequalis* Röber, 1925

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (*Chaupichaca, Cosñipata Valley – Chontachaca 950 m to Rocotal 2,000 m, Marcapata*); Pasco (*Pozuzo*); Puno (*Río Inambari, Sandia, San Gaban, Uruhasi*). **Bolivia:** Beni; Cochabamba (*Alto Palmar, Yungas del Palmar*); La Paz (*Caranavi, Chaco, Chulumani, Coroico, Farinas, Río Zongo, Taipiplaya*).

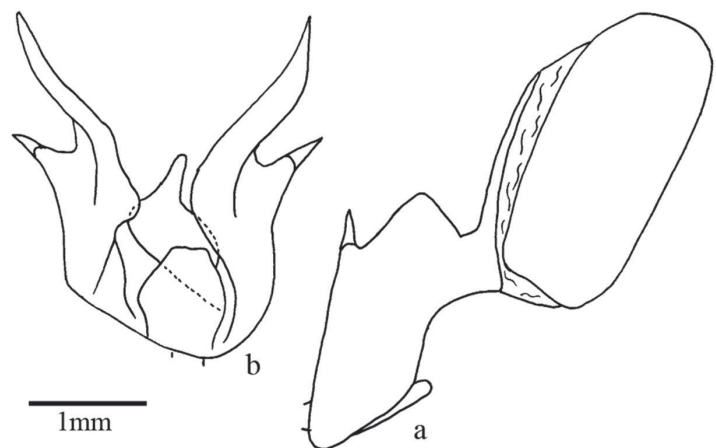
The citations **Colombia** (Hayward 1947), **Ecuador** and **Peru** (Mabille & Boulet 1908), **Peru** (Mabille 1912, Draudt 1921, Spitz 1932, Lamas 1969) and **Peru:** Junín (*Chanchamayo*) (Lindsey 1925) are probably incorrect.

TAXONOMY – *Mimardaris periphema* **stat rest., comb. rev.**, currently a subspecies of *Mimardaris sela* **comb. rev.**, has here its status revalidated. This taxon is very different on the ventral hindwing (see Pl. 47, fig. 20 and 23) is sympatric with *M. sela peruviana* **comb. rev.**, at least, in Pozuzo, Pasco, Peru and the DNA barcode (n= 3) has the minimum divergence to this taxon (n= 4) of 2.1%. There are no significant differences in the male genitalia.

TYPES:

Pyrrhopyga [**sic**] *periphema* Hewitson, [1875]. The original description mentions an unspecified number of specimens from Bolivia, Buckley leg. in collection Hewitson. To ensure the correct identification of *Pyrrhopyga periphema*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – *periphema* Hew./ Type – H. 71/ Bolivia Hewitson Coll. 79.-69. *Pyrrhopyga periphema* .1./ NHMUK(E) #982968/ *Mimoniades sela periphema* Hewitson, 1873/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga periphema* Hewitson, [1875], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Mimoniades inaequalis Röber, 1925. Lectotype male (O. Mielke 1989a) with the following labels: / Bolivia/ *inaequalis*/ Lectotypus *Mimoniades inaequalis* Röber, 1925. O. Mielke det. 1988/ Stauding. & Bang-Hass Dresden Ankauf 1961/ Staatl. Museum für Tierkunde Dresden/ GART specimen ID: 02566 Exemplar + Etiketten dokumentiert specimen + label data documented/. SMT.



Text-Fig. 50. *Mimardaris periphema* **stat rest., comb. rev.** Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (Chaco, La Paz, Bolivia, OM 11.989).

Sarbia Watson, 1893

Syn.: *Metardaris* Mabille, 1903

Seven species.

DISTRIBUTION – Southeastern Peru to northwestern Argentina, central to southern Brazil, northeastern Argentina and Uruguay.

Sarbia cosinga (Hewitson, 1874)

Four subspecies.

DISTRIBUTION – Southeastern Peru to northwestern Argentina.

TAXONOMY – The geographical distribution of the four subspecies is not clear. Some subspecies may occur in nearby area. The species occurs between the altitudes of 1,400 and 3,700 m.

Sarbia cosinga cosinga (Hewitson, 1874)

Pls. 48–49, Figs. 8, 12–13 (♂), 9–10 (♀) (d, v)

Syn.: *Metardaris consinga* [**sic**] var. *obscura* Mabille, 1903

Metardaris consinga [**sic**] var. *sanguinea* Mabille, 1903

Metardaris stygiana Mabille & Boulet, 1908, **nom. nud.** (ICZN, Art. 11.6)

Metardaris territa Mabille & Boulet, 1908, **nom. nud.** (ICZN, Art. 11.6)

DISTRIBUTION – Southeastern Peru to northwestern Argentina.

Peru: Cusco (Challanga, Marcapata, Paucartambo – Quebrada Chusa 3150 m, Vilcanota); Department? (Acampo). **Bolivia:** Chuquisaca (Huata); La Paz (Achumani, Cercado, Illimani, La Paz, Mapiri, Río Tamampaya); Potosí (Ancoma); Department? (Quatiguaya). **Argentina:** Salta (Santa Victoria 1,400 m).

REMARKS – This subspecies flight very close to *S. cosinga catana* near Paucartambo, Cusco, Peru, in a very small altitude difference. While *S. cosinga catana* occur at 3,500 m in Acjanaco, approximately 25 km from Paucartambo, *S. cosinga cosinga* occurs at 3,150 m in the Quebrada Chusa, about 10 km from Paucartambo on the same road to Pilcopata.

TYPES:

Pyrrhopyga [**sic**] *cosinga* Hewitson, 1874. The original description mentions an unspecified number of specimens from Bolivia, Buckley leg. in collection Buckley. To ensure the correct identification of *Pyrrhopyga cosinga*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – *cosinga* Hew./ Type H T/ Bolivia Hewitson Coll. 79.-69. *Pyrrhopyga cosinga*. 2./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga cosinga* Hewitson, 1874, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Metardaris consinga [**sic**] var. *obscura* Mabilles, 1903. The original description mentions an unspecified number of specimens from Bolivia. To ensure the correct identification of *Metardaris consinga* var. *obscura*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Bolivie 1904 O. Staudinger Coll. Boulet Museum Paris/ *M. cosinga* var. *obscura* Mab. G. H. p.1 *stygiaria* & *territa* Stgr. in litt./; two other labels will be added: / Lectotypus/ Lectotypus *Metardaris consinga* var. *obscura* Mabilles, 1903, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Metardaris consinga [**sic**] var. *sanguinea* Mabilles, 1903. The original description mentions an unspecified number of specimens from Bolivia. To ensure the correct identification of *Metardaris consinga* var. *sanguinea*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H.80/ var. *sanguinea* Ex musaeo P. Mabilles/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Metardaris consinga* var. *sanguinea* Mabilles, 1903, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Metardaris stygiaria Mabilles & Boulet, 1908. The name was published as a synonym of *Metardaris cosinga* var. *obscura*, a **nomen nudum** (ICZN, Art. 11.6).

Metardaris territa Mabilles & Boulet, 1908. The name was published as a synonym of *Metardaris cosinga* var. *obscura*, a **nomen nudum** (ICZN, Art. 11.6).

***Sarbia cosinga catana* (Evans, 1951)**

Pls. 48–49, Figs. 1, 7 (♂), 2, 11 (♀) (d, v)

DISTRIBUTION – Southern Peru to central-eastern Bolivia.

Peru: Apurímac (Chincheros, Chuquibambilla); Cusco (Abra Acjanaco 3,500 m, Abra Malaga, between Quillabamba and Machu Picchu, Cajon, Calca, Cordillera de Vilcabamba, Cosñipata Valley – Wayquecha 2,950 m), Machu Picchu, Quebrada Tambillo); Huánuco (Caina, Huacar); Junín (Maraynioc, Yauli); Pasco (Río Palcazu); Puno (Acopampa, Carabaya – Santo Domingo, Oconeque). **Bolivia:** Cochabamba (El Limbo); La Paz (Acero Marka, Farinas, Incachaca, Parque Nacional Madidi 3,000–3,600 m, Pongo); Potosí (Ancoma).

TYPE:

Metardaris cosinga catana Evans, 1951. Holotype male with the following labels: / Type – *catana* Evans/ Limbani, Carabaya, [Puno, Peru], dry s., 9500 ft., May 04. G. Ockenden)/ Rothschild Bequest B.M. 1939-1/. NHMUK.

***Sarbia cosinga cedra* (Evans, 1951)**

Pls. 48–49, Figs. 3 (♂), 4 (♀) (d, v); Pls. 74–75, Figs. 9 (♂) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (Cajon). **Bolivia:** Cochabamba (Abra de Colomi,

Chapare, Corani Pampa); La Paz (Parque Nacional Madidi 1,500–1,800 m, San Antonio).

TYPE:

Metardaris cosinga cedra Evans, 1951. Holotype male the following labels: / Type – *cedra* Evans/ Abra de Colomi 3600 Meters Andes Mts. [Cochabamba] Bolivia IX-12/ Brit. Mus. 1934-187/. NHMUK.

***Sarbia cosinga circe* (Evans, 1951)**

Pls. 48–49, Figs. 5 (holotype ♂), 6 (♀) (d, v)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: Cochabamba (100 km NO); La Paz (Cuticucho – Río Zongo, Yani, Yungas).

TYPE:

Metardaris cosinga circe Evans, 1951. Holotype male with the following labels: / Type – *circe* Evans/ Bolivia Yungas La Paz 1000 m ex H. Rolle 1902/ 265/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK. The altitude of 1,000 m is probably wrong.

***Sarbia damippe* Mabilles & Boulet, 1908**

Pls. 48–49, Figs. 21 (♂), 22 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil, northeastern Argentina and Uruguay.

Brazil: Minas Gerais (Camanducaia – Monte Verde, Catas Altas – Caraça, Datas, Diamantina, Passa Quatro, Poços de Caldas, Virgínia); Rio de Janeiro (Itatiaia); São Paulo (Bananal – Bocaina, Barueri, Campos do Jordão, Castro, Jundiá, Piquete – Barreira de Piquete, São Paulo); Paraná (Curitiba, Fernandes Pinheiro, Guarapuava, Inácio Martins, Lapa, Mangueirinha, Palmas, Quitandinha, Rio Negro, São José dos Pinhais, Tibagi, União da Vitória); Santa Catarina (Bom Jardim da Serra, Curitiba, Joinville – Vila Nova, Lages, Mafra, Massaranduba, Ponte Alta do Norte, Rio Negrinho, Santa Cecília, São Bento do Sul – Rio Vermelho, São Joaquim, Urubici – Morro da Igreja); Rio Grande do Sul (Bom Jesus, Cambará do Sul, Caxias do Sul, Pelotas, Porto Alegre, Santo Augusto, São Francisco de Paula, Vacaria). **Uruguay:** Cerro Largo (S. del Quebracho). **Argentina:** Misiones.

The citation **Venezuela** (Evans 1951) is incorrect (Orellana [2010]).

TYPE:

Sarbia damippe Mabilles & Boulet, 1908. The original description mentions an unspecified number of males and females from Rio Grande do Sul, [Brazil] and Brazil. To ensure the correct identification of *Sarbia damippe*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Brésil méridional 1904 O. Staudinger Coll. Boulet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Sarbia damippe* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. This specimen has the club of both antennae very well curved. There are five other syntypes; these have the following labels: (1 – female) / Type/ Sr. Catarina Brésil 1906 Herm. Rolle Coll. Boulet Museum Paris/; (2 – female) / Type/ Porto Alegre Brésil 1908 Herm. Rolle Coll. Boulet Museum Paris/; (3 – male) / Type Brésil 1840 Coll. Lacordaire Museum de Paris/; (4 and 5 – male and female) / Type/ Brésil méridional 1904 O. Staudinger Coll. Boulet Museum de Paris/; two other labels will be added on each: / Paralectotypus/ Paralectotypus *Sarbia damippe* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

***Sarbia oneka* (Hewitson, 1866)**

Pls. 48–49, Figs. 23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Brazil

Brazil.

The citations **Ecuador** (Mabilles 1903, 1912, Mabilles & Boulet 1908, Draudt 1921), **Venezuela** (Hewitson 1866, Bell 1934b, 1946, Williams & Hayward 1944, O. Mielke & Casagrande 2002, O. Mielke 2004) and **Brazil:** Amazonas (Mabilles & Boulet 1908) are probably

incorrect (Orellana [2010]). There are no exact data about this species.

TYPE:

Pyrrhopyga [**sic**] *oneka* Hewitson, 1866. Lectotype female (O. Mielke & Casagrande 2002) with the following labels: / Type/ Type H T/ 44 124/ Venezuela Becker 44-124/ Lectotypus/ Lectotype *Pyrrhopyga oneka* Hewitson, 1866, O. Mielke & Casagrande det./ NHMUK.

***Sarbia pertyi* (Plötz, 1879)**

Pls. 50–51, Figs. 1 (♂), 2 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Minas Gerais (*Passa Quatro*); Rio de Janeiro (*Itatiaia*); São Paulo; Paraná (*Campo Largo*, *Castro*, *Curitiba*, *Guarapuava*, *Lapa*, *Palmas*, *Pirai do Sul*, *Porto Amazonas*, *Quitandinha*, *Rio Negro*, *São José dos Pinhais*, *Teixeira Soares – Valinhos do Sul*, *União da Vitória*); Santa Catarina (*Corupá*, *Curitibanos*, *Itaiópolis*, *Joinville – Vila Nova*, *Lages*, *Mafra*, *Massaranduba*, *Monte Castelo*, *Otacilio Costa*, *Ponte Alta do Norte*, *Rio Negrinho*, *Santa Cecília*, *São Bento do Sul – Rio Vermelho*, *São Joaquim*, *Urubici – Morro da Igreja*); Rio Grande do Sul (*Bom Jesus*, *Cambará do Sul*, *São Francisco de Paula*, *Vacaria*).

The citation **Guyana** (Mabille 1903) is incorrect.

TYPE:

Pyrrhopyga [**sic**] *pertyi* Plötz, 1879. The original description mentions an unspecified number of specimens from Brazil. No syntypes were found in MfN or ZSM, where nearly all Plötz's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Pyrrhopyga pertyi*, mentioned as *Sarbia pertyi* by Evans (1951), a male is **herein designated as a neotype**; the specimen with the following labels: / Neotypus/ 24-II-1983 Paine, Lajes, SC [Santa Catarina, Brazil]. 900 m. [O.] Mielke & Casagrande leg./ Neotypus *Pyrrhopyga pertyi* Plötz, 1879 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.275/. DZUP.

***Sarbia soza* Evans, 1951**

Pls. 50–51, Figs. 3 (♂), 4 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Minas Gerais (*Camanducaia – Monte Verde*, *Delfim Moreira*, *Passa Quatro*); Rio de Janeiro (*Itatiaia*); São Paulo (*Bananal – Serra da Bocaina*, *Campos do Jordão*); Rio de Janeiro (*Itatiaia*); Paraná (*Castro*); Santa Catarina (*Lages*, *Santa Cecília*).

TYPE:

Sarbia oneka soza Evans, 1951. Holotype male with the following labels: / Type – *soza* Evans/ Castro, Parana. [Brazil]/ E.D.Jones Coll. Brit. Mus. 1919-295/. NHMUK.

***Sarbia xanthippe* (Latreille, [1824])**

Two subspecies.

DISTRIBUTION – Central to southern Brazil and northeastern Argentina.

***Sarbia xanthippe xanthippe* (Latreille, [1824])**

Pls. 50–51, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Central to southeastern Brazil.

Brazil: Goiás (*Alto Paraíso*); Bahia (*Rio de Contas – Pico das Almas* 1,600 m); Minas Gerais (*Catas Altas – Caraça*, *Diamantina*, *Santana do Riacho – Serra do Cipó*, *São Roque de Minas*).

The citations **Brazil:** Amazonas (*Manaus*) (Bell 1934b), **Argentina:** Misiones (Köhler 1923, Hayward 1973), **Argentina:** Buenos Aires (Hayward 1939, 1941, 1948, 1973) and **Paraguay** (Hayward 1948) are incorrect.

TYPE:

Hesperia xanthippe Latreille, [1824]. The original description

mentions an unspecified number of specimens from Brazil, Langsdorff leg. To ensure the correct identification of *Hesperia xanthippe*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *H. Xanthippe* Lat Brésil/ Ex musaeo P. Mabille 1923/ *xanthippe* R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Hesperia xanthippe* Latreille, [1824], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Sarbia xanthippe spixii* (Plötz, 1879)**

Pls. 50–51, Figs. 7 (♂), 8 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil and northeastern Argentina.

Brazil: Minas Gerais (*Barbacena*, *Camanducaia – Monte Verde*, *Conceição dos Ouros*, *Itamonte*, *Passa Quatro*, *Poços de Caldas*, *Ponto dos Volantes*, *Virgínia*); Rio de Janeiro (*Itatiaia*, *Resende – Garganta do Registro*); São Paulo (*Bananal – Parque Nacional da Bocaina*, *Campos do Jordão*, *Delfim Moreira*, *Morro Agudo*, *Piquete – Barreira de Piquete*); Paraná (*Araucária*, *Balsa Nova*, *Bituruna*, *Campo Largo*, *Castro*, *Curitiba*, *Guarapuava*, *Guaratuba – Pontal do Itararé* 1,400 m, *Inácio Martins*, *Lapa*, *Palmas*, *Quatro Barras*, *Rio Negro*, *São José dos Pinhais*, *Tibagi*, *Tijucas do Sul*, *Turvo*); Santa Catarina (*Bom Jardim da Serra*, *Blumenau*, *Corupá*, *Curitibanos*, *Lages*, *Mafra*, *Massaranduba*, *Monte Castelo*, *Ponte Alta do Norte*, *Rio dos Cedros*, *Santa Cecília*, *São Bento do Sul – Rio Vermelho*, *São Joaquim*, *Urubici*; *Morro da Igreja*); Rio Grande do Sul (*Ana Rech*, *Bom Jesus*, *Cambará do Sul*, *Canela*, *Caxias do Sul*, *Gramado*, *Pelotas*, *São José dos Ausentes*). **Argentina:** Buenos Aires; Misiones.

The citations **Paraguay** (Hayward 1948) is probably incorrect.

TYPE:

Pyrrhopyga [**sic**] *spixii* Plötz, 1879. The original description mentions an unspecified number of specimens from Brazil in collection Weymer. To ensure the correct identification of *Pyrrhopyga spixii*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ *spixi*, Plötz./ *atys* Mab./ *Atys* Mab. *Spixi* Plötz/ Blumenau [Santa Catarina, Brazil]/ Coll. Staudinger/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga spixi* Plötz, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

***Sarbia curitiba* O. Mielke & Casagrande, 2002**

Pls. 50–51, Figs. 9 (holotype ♂), 10 (allotype ♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Minas Gerais (*Poços de Caldas*); São Paulo (*Campos do Jordão*); Paraná (*Castro*, *Curitiba*, *General Carneiro*, *Guarapuava*, *Palmas*, *Prudentópolis*, *São José dos Pinhais*, *Tibagi*); Santa Catarina (*Bom Jardim da Serra*, *Curitibanos*, *Lages*, *Monte Castelo*, *Ponte Alta do Norte*, *Santa Cecília*, *São Bento do Sul – Rio Vermelho*, *São Joaquim*, *Urubici – Alto Rio Canoas*; *Morro da Igreja*; *Santa Teresinha*); Rio Grande do Sul (*Bom Jesus*, *Cambará do Sul*, *Canela*, *São Francisco de Paula*, *São José dos Ausentes*, *Vacaria*).

TYPE:

Sarbia curitiba O. Mielke & Casagrande, 2002. Holotype male with the following labels: / Holotipo/ [Cascatinha], Curitiba, Paraná, 900 metros, Brasil, 22-I-1977, O. Mielke leg./ DZ 5.055/ Holotipo *Sarbia curitiba* [O.] Mielke & Casagrande, 2002/. DZUP.

***Sarbiena* Grishin, 2019**

One species.

DISTRIBUTION – Central-eastern to southeastern Brazil.

***Sarbiena catomelaena* (Mabille & Boulet, 1908)**

Two subspecies.

DISTRIBUTION – Central-eastern to southeastern Brazil.

***Sarbiena catomelaena catomelaena* (Mabille & Boulet, 1908)**

Pls. 48–49, Figs. 14 (♂), 15 (♀), 16 (neotype ♂ of *Sarbia mello-leitaoi*) (d, v)

Syn.: *Sarbia mello-leitaoi* Hayward, 1942, incorr. orig. spell. (ICZN, Art. 32.5)

Sarbia melloleitaoi; Bridges, 1983; correct spelling

DISTRIBUTION – Southeastern Brazil.

Brazil: Minas Gerais (*Aiuruoca – Serra dos Garcias, Catas Altas – Caraça, Diamantina, Itabirito, Nova Lima, Passa Quatro, Poços de Caldas, Santana do Riacho – Serra do Cipó, Virgínia*); Rio de Janeiro (*Itatiaia*); São Paulo (*Campos do Jordão, Delfim Moreira*).

The citation **Argentina:** Misiones (Canals 2003) is likely incorrect.

TYPES:

Sarbia catomelaena Mabille & Boulet, 1908. The original description mentions one male from Minas Gerais [Brazil] in MNHN (Coll. Boulet) and one male without data in collection Mabille. To ensure the correct identification of *Sarbia catomelaena*, one syntype female (not male as stated and figured in the original description) is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Minas Geraes Brésil 1885 E. Gounelle/; two other labels will be added: / Lectotypus/ Lectotypus *Sarbia catomelaena* Mabille & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Sarbia mello-leitaoi Hayward, 1942. Holotype male with the following labels: / Tipus/ 23-2.1920/ *Sarbia melloleitaoi* Hayward. ♂ Holotipo./ No 60015 Proc. Itatiaia [Rio de Janeiro State, Brazil] Zikán [leg.]/. MNRJ. This specimen was lost by fire (2-IX-2018). To ensure the correct identification of *Sarbia mello-leitaoi*, mentioned as a synonym of *Sarbia catomelaena* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Virginia S[ul]. Minas Ger [rais, Brazil]. Faz[enda dos]. Campos 1500 m J. Zikán [leg.] 22-II-1918/ Neotypus *Sarbia mello-leitaoi* Hayward, 1942 O. Mielke, Brockmann & C. Mielke det. 2019/ OM 36.420/. DZUP. The locality Fazenda dos Campos, Virgínia, Minas Gerais is very close to Itatiaia, Rio de Janeiro; only 50 km in a straight line.

Sarbiena catomelaena pico* O. Mielke & Casagrande, *ssp. n.

Pls. 48–49, Figs. 19 (holotype ♂), 20 (allotype ♀) (d, v)

DISTRIBUTION – Central-eastern Brazil (Bahia).

Brazil: Bahia (*Rio de Contas – Pico das Almas 1,600 m*).

DESCRIPTION – Male: forewing length 22 mm. Very similar to *S. catomelaena catomelaena*, but the yellow bands on both wings, dorsally and ventrally, are narrower (nearly 1 mm); the ventral yellow spots at the thorax are absent in the forewing and reduced in hindwing, and the ventral ventro-basal yellow spot at the base of the hindwing is longer and curved, reaching the yellow portions of the abdomen. The genitalia don't differ from those of *S. catomelaena catomelaena*.

Female: forewing length 26 mm. Similar to male, but the yellow of the abdomen under the forewing is present, but reduced.

ETYMOLOGY – The name refers to the name of the “Pico das Almas”, the type locality.

REMARKS – Both known specimens were collected nearly 12:00 h on light-rose Asteraceae flowers at the margin of the “Campo do Queiroz” (Queiroz's field), just on the trail to the “Pico das Almas”.

TYPES:

Holotype male with the following labels: / Holotypus/ 27-I-2007, Pico das Almas, Rio de Contas, Bahia, Brasil, 1.600 m, O. Mielke & Casagrande leg./ gen[itália]. prep[arada]. [O.] Mielke 2020/ Holotypus *Sarbiena catomelaena pico* O. Mielke & Casagrande. O Mielke & Casagrande det. 2020/ DZ 16.094/. DZUP.

Allotype female with the following labels: / Allotypus/ 27-I-2007, Pico das Almas, Rio de Contas, Bahia, Brasil, 1.600 m, O. Mielke & Casagrande leg./ Allotypus *Sarbiena catomelaena pico* O. Mielke & Casagrande. O Mielke & Casagrande det. 2020/ DZ 51.482/. DZUP.

***Santea* Grishin, 2019**

One species.

DISTRIBUTION – Southeastern to southern Brazil and northeastern Argentina.

***Santea antias* (C. Felder & R. Felder, 1859)**

Pls. 48–49, Figs. 17 (♂), 18 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil and northeastern Argentina.

Brazil: Minas Gerais (*Passa Quatro, Poços de Caldas, Virgínia*); São Paulo (*Casabranca, Iperó, Teodoro Sampaio – Porto Cabral*); Paraná (*Castro, Curitiba, Foz do Iguaçu, Guarapuava, Inácio Martins, Londrina, Palmas, Palmeira, Pirai do Sul, Ponta Grossa, Reserva, Tibagi – Estiva, União da Vitória, Ventania*); Santa Catarina (*Bom Jardim da Serra, Canoinhas, Corupá, Massaranduba, Monte Castelo, Seara – Nova Teutônia, Urubici*); Rio Grande do Sul (*Canela*). **Argentina:** Misiones (*Puerto Iguazú*).

The citation **Colombia:** Cundinamarca (*Bogotá*) (Mabille & Boulet 1908) is incorrect.

TYPE:

Pyrrhopyga [*sic*] *antias* C. Felder & R. Felder, 1859. The original description mentions two males and one female from Brazil in collection C. Felder & R. Felder. To ensure the correct identification of *Pyrrhopyga antias*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Brasil merid-/ *Antias*. n./ 244/ Rothschild Bequest B.M.1939-1/ NHMUK(E) #982957/ *Sarbia antias* Felder, 1859/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga antias* C. Felder & R. Felder, 1859, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Mysarbia* O. Mielke, 2002**

One species.

DISTRIBUTION – Costa Rica to southern Brazil.

***Mysarbia sejanus* (Hopffer, 1874)**

Three subspecies.

DISTRIBUTION – Costa Rica to western Ecuador, central and eastern Colombia to central-eastern Bolivia, Venezuela to French Guiana and Brazil.

***Mysarbia sejanus sejanus* (Hopffer, 1874)**

Pls. 50–51, Figs. 11 (♂) (d, v)

DISTRIBUTION – Western and eastern Ecuador to southeastern Peru.

Ecuador: Chimborazo (*Riobamba*); Loja (*Loja, San Francisco*); Morona-Santiago (*Macas*); Pastaza (*Mera, Puyo*); Santo Domingo de los Tsáchilas (*Alluriquín*); Tungurahua (*Sarayacu*); Zamora-Chinchipe (*Zamora*). **Peru:** Huánuco (*Cucharas, Tingo Maria*); Junín (*Chanchamayo, La Merced, Perené, Río Colorado, Satipo, Shima*); Pasco (*Palcazu*); Puno (*Sandia*); San Martín (*Juanjuí*).

The citation **Costa Rica** (Butler & H. Druce 1874) may be incorrect.

TYPE:

Pyrrhopyge sejanus Hopffer, 1874. The original description

mentions an unspecified number of males from Peru/ Bolivia. To ensure the correct identification of *Pyrrhopyge sejanus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Lectotypus/ Perú Schmeltz/ *Sejanus* Hopff.* / 18259/; another label will be added: / Lectotypus *Pyrrhopyge sejanus* Hopffer, 1874, O. Mielke, Brockmann & C. Mielke det. 2014/. MfN. Although the lectotype label was attached in 1979 by O. Mielke, the designation was never published.

***Mysarbia sejanus erythrostigma* (Röber, 1925)**

Pls. 50–51, Figs. 12 (♂) (d, v)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: Beni (*Río Benicito, Río Yata*); Cochabamba (*Cristal Mayu*); La Paz (*Caranavi, Farinas, Mapiri, Río Zongo, San Ernesto, Sarampiuni*); Santa Cruz (*Buenavista*).

TYPE:

Mysoria erythrostigma Röber, 1925. Lectotype male (O. Mielke 1989a) with the following labels: / Río Songo [*sic*] [La Paz] Bolivia, 750 m Coll. Fassl/ *thasus* Cr. *erythrostigma* Röb. ? Bolivia/ Stauding. & Bang-Hass Dresden Ankauf 1961/ Staatl. Museum für Tierkunde Dresden/ Lectotypus *Mysoria erythrostigma* Röber, 1925 O. Mielke det. 1988/ GART specimen ID: 02557 Exemplar + Etiketten dokumentiert specimen label data documented 2003/. SMT.

***Mysarbia sejanus stollii* O. Mielke, 2002**

Pls. 50–51, Figs. 13–14 (♂), 15 (♀) (d, v)

Syn.: *Papilio thasus* Stoll, 1782, *praeocc.* (Stoll, 1780)

DISTRIBUTION – Costa Rica to western Ecuador, central and eastern Colombia to southern Peru, Venezuela to French Guiana and Brazil.

Costa Rica: Cartago (*Hacienda Moravia de Chirripo*); Guanacaste (*Área de Conservación Guanacaste*). **Panama:** Darién (*Caná*). **Colombia:** Amazonas (*Leticia, San Martín*); Boyacá (*Muzo, Otanche*); Caquetá (*Ortegaúza*); Chocó (*Istmina*); Cundinamarca (*Bogotá, La Aguadita*); Putumayo (*Mocoa*). **Ecuador:** Esmeraldas (*Río Chuchuvi*). **Peru:** Amazonas (*Huambo, Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Pilcopata 500 m*); Huánuco (*Tingo Maria*); Loreto (*Contamana, Curinama, Iquitos, Pebas, Quebrada Polis, Río Putumayo, San Antonio, Ucayali*); Madre de Dios (*Parque Nacional Soberanía Bahuaia Sonene, Parque Nacional del Manu, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco), Puerto Maldonado, Tambopata Reserve*); Pasco (*Villa Rica*); San Martín (*Juanjuí, Moyobamba*); Tacna; Ucayali (*Pucallpa*). **Venezuela:** Amazonas (*Camani, La Esmeralda, Puerto Ayacucho, San Pedro de Cataniapo, Upper Río Mavaca*); Aragua (*Parque Nacional Henry Pittier*); Bolívar (*Gran Sabana, Kanaracuni – Alto Caura, Maripa, Río Caura, Snapure, Wariwantey*); Táchira (*Las Cuevas – Represa Uribante Caparo, Río Doradas, Río Frío, Río Negro, San Joaquín de Navay*); Trujillo (*Betijoque, La Gira*); Zulia (*Misión El Rosário*). **Suriname:** Paramaribo. **French Guiana:** Cayenne (*Piste de la Crique Trésor, Sinnamary – Pont Malmanoury*). **Brazil:** Amazonas (*Manicoré, Río Negro, Río Uaupés, São Gabriel da Cachoeira – Iauareté, São Paulo de Olivença, Uaupés*); Pará (*Óbidos*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor, Xapuri*); Rondônia (*Cacaulândia, Campos dos Urupás, Candeias do Jamari, Cautário, Ji-paraná, Pimenta Bueno, Porto Velho, Vilhena*); Mato Grosso (*Alta Floresta, Diamantino, Juína, Pantanal, Verá*); Mato Grosso do Sul (*Dourados*); Maranhão (*Aldeia Aracu – Igarapé Gurupi-Una*); Distrito Federal (*Brasília*); Minas Gerais (*Brumadinho, Carmo do Rio Claro, Diamantina, Divinópolis, Ouro Branco, Paracatu*); Rio de Janeiro (*Casimiro de Abreu – Reserva Biológica Poço das Antas, Duque de Caxias, Petrópolis*); São Paulo; Paraná (*Rolândia*).

TYPES:

Papilio thasus Stoll, 1782. The original description mentions an unspecified number of specimens from Suriname. To ensure the correct identification of *Papilio thasus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Coll. Calkoen Suriname/ *Pyrrhopyga thasus* Cr./ / Museum Leiden/

Museum Leiden *Mysoria thasus* Cr. Det: J. Mabilles/ *Mysoria thasus* Cr./; two other labels will be added: / Lectotypus/ Lectotypus *Papilio thasus* Stoll, 1782, O. Mielke, Brockmann & C. Mielke det. 2019/. NBC. There are two more syntypes, one male and one female, herein designated as paralectotypes; these specimens have the following labels: / Calkoen Suriname/ *Pyrrhopyga thasus* Cr./ *Mysoria thasus* Cr./ Museum Leiden/ ♂ / Museum Leiden *Mysoria thasus* Cr. Det : J. Mabilles/; and / Col. Calkoen Suriname/; two other labels will be added on each: / Paralectotypus/ Paralectotypus *Papilio thasus* Stoll, 1782, O. Mielke, Brockmann & C. Mielke det. 2019/. NBC.

Mysarbia sejanus stollii O. Mielke, 2002 is a **nomen novum** for *Papilio thasus* Stoll, 1782; so the name bearing type specimen is the lectotype of *Papilio thasus* Stoll, 1782.

***Mysoria* Watson, 1893**

Four species.

DISTRIBUTION – Mexico to Uruguay.

***Mysoria amra* (Hewitson, 1871)**

Pls. 50–51, Figs. 16 (neotype ♂ of *Amenis brasiliensis*), 19 (♂), 20 (♀) (d, v)

Syn.: *Amenis brasiliensis* Mabilles, 1903

DISTRIBUTION – Western and eastern Mexico to Guatemala.

Mexico: Chiapas (*Motozintla, Ocozacoautla, Rizo del Oro*); Colima (*Água Dulce, Colima, Comala, El Salto, Manzanillo, Punta de Agua de Camotlán*); Guerrero (*Acapulco, El Treinte, Rincón, Tierra Colorada, Villa de Álvarez – Agua Dulce*); Jalisco (*Boca de Tomatlán, Chamela, Cihuatlán, El Agostadero, Guadalajara, La Calera, La Cuesta, Mismaloya, Puerto Vallarta, Sierra de Talpa*); Michoacán (*Acalpican, Coahuayana*); Morelos (*Sierra de Huautla*); Nayarit (*Compostela, El Guamúchil, El Refilión, Jumatán, Laguna Santa Maria del Oro, Las Piedras, Mecatán, Paso de Mesillas, Playas Novillero, San Blas, Singayta*); Oaxaca (*Candelaria Loxicha, Putla, San Miguel del Puerto, San Pedro*); Veracruz (*Misantla*). **Guatemala:** San Marcos (*Río Naranjo*). **El Salvador:**

The citations **Brazil** (Mabilles 1903) and **Guyana** (Hall 1939, 1940) are incorrect.

TYPES:

Pyrrhopyga [*sic*] *amra* Hewitson, 1871. The original description mentions an unspecified number of specimens from Mexico in collection Saunders & Hewitson. To ensure the correct identification of *Pyrrhopyga amra*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H. 37/ Mexico Hewitson Coll. 79.-69. *Pyrrhopyga amra*. 3./ 270/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga amra* Hewitson, 1871, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Amenis brasiliensis Mabilles, 1903. The original description mentions an unspecified number of specimens from Brazil, certainly an incorrect locality. This species occurs only between Mexico and El Salvador (?). No syntypes were found in any museum or collection mentioned above (see “Abbreviations of the collections”). To ensure the correct identification of *Amenis brasiliensis*, mentioned as a synonym of *Mysoria amra* by Mabilles & Boulet (1908) and Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Candelaria Loxicha Oaxaca, Mexico, 500 m 29-VIII-1978 Welling leg./ Neotypus *Amenis brasiliensis* Mabilles, 1903 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.245/. DZUP.

***Mysoria affinis* (Herrich-Schäffer, 1869)**

Pls. 50–51, Figs. 17 (♂), 18 (♀) (d, v)

Syn.: *Mysoria wilsoni* Freeman, 1969

DISTRIBUTION – Western and eastern Mexico to Guatemala.

Mexico: Chiapas (*Bombana, Comitán de Domínguez, Jiquipilla – Parque El Aguacero, La Trinitaria – Las Delicias, Mapastepec*); Colima (*Colima, Comala, Comitán, La Salada, Madrid, Manzanillo – El Salto, Minatitlán – Platanarillos, Ojo de Agua, Pedro Nuñez, Punta de Agua de Camotlán, Río Comala, Tecumán – La Salada, Villa de Álvarez – Agua Dulce*); Guanajuato (*Paracuaro*); Guerrero (*Acahizotla, Acapulco, Atoyac – Rincón de las Parotas, Chapultepec, Colotlipa, El Treinte, Iguala, La Venta, Mezcala, Taxco, Tepecoacuilco de Trujano – Río Mezcala, Zihuatanejo*); Jalisco (*Autlán – Ahuacapán, Cihuatlán, El Agostadero, Ejutla, La Huerta – Chamela Bay, Mazatán, Mismaloya, P. H. Cupatitzio, Puerto Vallarta – Banderas Bay, San José del Rincón, San Miguel de Hidalgo, Tuxcacuesco*); Mexico (*Toluca – San Nicolás Tolentino*); Michoacán (*Arteaga – Arteaga, Coahuayana – San Telmo, Gabriel Zamora – Barranca de Cóbano, La Huacana – Presa Zicuirán, Las Juntas, Rancho El Zorrillo*); Morelos (*Ayala, Cuernavaca, Puente de Ixtla, Rancho Viejo, Sierra de Huautla, Tepoztlán*), Nayarit (*Compostela, Sayulita, Tepic*); Oaxaca (*Nejapa de Madero, Salina, San Bartolo Yautepec, San Juan Lajarcia, San Pedro Pochutla – Puerto Ángel, Santa Maria Tlahuitoltepec – Nejapa, Sierra de Juárez*); Puebla (*Matamoros – Jaulillas*); San Luis Potosí (*San Nicolás Tolentino*); Sinaloa (*Culiacán, Mazatlán, Vanadie*); Veracruz (*Misantla, Presidio*). **Guatemala:** Santa Rosa (*Guazacapán*). **El Salvador:**

The citation **Brazil:** Amazonas or Pará (Fischer-Sigwart 1923) is incorrect.

TYPES:

Pyrrhopyga [*sic*] *affinis* Herrich-Schäffer, 1869. The original description mentions an unspecified number of specimens from Tropical America. To ensure the correct identification of *Pyrrhopyga affinis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Coll. H.-Sch./ Coll. Staudinger/ *affinis*, HS./ 2: 3/ *affinis* m./ 91./ *Affinis* H.-Sch./; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga affinis* Herrich-Schäffer, 1869, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.

Mysoria wilsoni Freeman, 1969. Holotype male with the following labels: / Mexcala Gro. [Guerrero] Mex. VII-22-56 Kent Wilson/ Holotype ♂ *Mysoria wilsoni* Freeman/. AMNH.

***Mysoria ambigua* (Mabille & Boulet, 1908)**

Pls. 50–51, Figs. 21 (lectotype ♂ of *Amenis proxima*), 22–23 (♂), 24 (♀) (d, v)

Syn.: *Amenis proxima* Mabille & Boulet, 1908

DISTRIBUTION – Western and eastern Mexico to western Colombia.

Mexico: Chiapas (*Boca de Chajul, Ocosingo – San Quentin*); Jalisco (*Guadalajara*); Tabasco; Veracruz (*Córdoba, Los Tuxtlas*). **Guatemala:** Alta Verapaz (*Polochic Valley*). **Honduras:** Atlántida (*La Ceiba*); Cortés (*San Pedro Sula*); Gracias a Dios (*Brus Laguna*). **El Salvador?** **Nicaragua:** Carazo (*Jinotepe*); Chontales; Granada (*Granada*); Managua; Matagalpa. **Costa Rica:** Alajuela (*San Mateo*); Cartago (*Cerro Azahar*); Guanacaste (*Área de Conservación Guanacaste, Las Cañas, Río Grande*); Puntarenas (*Barranca – La Guaria, Buenos Aires, Reserva Osa, San Vito*); San José (*Colón*). **Colombia:** Valle del Cauca (*Cali*).

TYPES:

Amenis ambigua Mabille & Boulet, 1908. Holotype male (one male is mentioned without data [Honduras, San Pedro Sula – on the label of the holotype]) with the following labels: / Type/ Honduras [Cortés] San Pedro Sula/ *amenis ambigua* Ex Oberthür Coll. Brit. Mus. 1927-3./ NHMUK.

Amenis proxima Mabille & Boulet, 1908. The original description mentions an unspecified number of males and females from Mexico, Costa Rica, Colombia and Paraguay in MNHN (collection Boulet). To ensure the correct identification of *Amenis proxima*, mentioned by Evans (1951) as a synonym of *M. barcastus ambigua*, one syntype male

is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Env. de Guadalajara Etat de Jalisco, Mexique. 1897 L. Digue/; two other labels will be added: / Lectotypus/ Lectotypus *Amenis proxima* Mabille & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. There are six more syntype males, here designated paralectotypes, one from Ozahar de Cartago, Costa Rica, 1,500 m, 1907, P. Biolley leg., one from Cauca, Colombia, 1904, C. Ribbe leg., two from Colombia, 1896, P. Vuillot leg. and two from Sapucay, Paraguay, 1905, Percy I. Lathy leg.; two other labels will be added: / Paralectotypus/ Paralectotypus *Amenis proxima* Mabille & Boulet, 1908. O. Mielke, Brockmann & C. Mielke det. 2019/. The two specimens from Sapucaí, Paraguari, Paraguay are *Mysoria barcastus barta*. MNHN.

***Mysoria barcastus* (Sepp, [1851])**

Six subspecies.

DISTRIBUTION – Panama to western and central Colombia and to French Guiana, Trinidad and Tobago, central-eastern Bolivia to north-northeastern Argentina and northern and northeastern Brazil to Uruguay.

***Mysoria barcastus barcastus* (Sepp, [1851])**

Pls. 52–53, Figs. 7 (♂), 8 (♀) (d, v)

Syn.: *Pyrrhopyga* [*sic*] *verbena* Butler, 1869

DISTRIBUTION – Guyana and Suriname.

Guyana: Mahaica-Berbice (*Blairmont*). **Suriname:** Brokopondo (*Brownsberg, Saramacca River*); Commewijne (*De Nieuves Grond, Mariënborg, Nieuw Amsterdam*); Marowijne (*Cottica River, Moengotapoe-Wawia*); Paramaribo (*Saint Barbara Plantation, Zorg en Hoop*); Sipaliwini (*Aona at Maroni*); Wanica (*Lelydorp*).

TYPES:

Papilio barcastus Sepp, [1851]. The original description mentions an unspecified number of specimens from Suriname. No syntypes were found in any museum or collection mentioned above (see “Abbreviations of the collections”) and according to Horn & Kahle (1936) the collection is lost. To ensure the correct identification of *Papilio barcastus*, mentioned as *Mysoria barcastus* by Evans (1951), a female is **herein designated as the neotype**; the specimen with the following label: / Surinam [Brokopondo] Brownsberg 28.VII.1996 leg. E. Jongerling/; two other labels will be added: / Neotypus/ Neotypus *Papilio barcastus* Sepp, [1851] O. Mielke, Brockmann & C. Mielke det. 2015/. SMT.

Pyrrhopyga [*sic*] *verbena* Butler, 1869. The original description mentions an unspecified number of females from South America in collection Mile. To ensure the correct identification of *Pyrrhopyga verbena*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type – H. 36/ Type – *verbena* Butler/ S. America 1503a/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga verbena* Butler, 1869, O. Mielke, Brockmann & C. Mielke det./ NHMUK.

***Mysoria barcastus venezuelae* (Scudder, 1872)**

Pls. 52–53, Figs. 1, 4, 12 (♀), 2, 11 (♂) (d, v)

Syn.: *Papilio acastus* Cramer, 1775, *praeocc.* (Linnaeus, 1758)
Mysoria decolor Mabille & Boulet, 1908
Mysoria cayennae Mabille & Boulet, 1908
Mysoria caucana Röber, 1925

DISTRIBUTION – Panama to western and central Colombia and to French Guiana, Trinidad and Tobago (Tobago) and northern Brazil (Amapá and Amazonas).

Panama: Chiriquí (*Bugaba, Lino, Potrerillos, Río Tolé, Santa Cruz*); Colón (*Gamboá*); Herrera (*Las Minas*); Isthmus of Panamá; Los Santos (*Río Oria*); Panamá (*Ancon, Chame, Chiva Chiva Trail, Cocoli,*

Corozal, Frijoles, Madden Forest, Panamá, Paraiso, Parque Nacional Soberanía, Pedro Miguel, Summit, Taboga Island); Panamá Oeste (Cerro Campana); Veraguas (Calobre). **Colombia:** Antioquia (La Pintada, Medellín, Santa Fe de Antioquia); Arauca; Boyacá (Muzo, Otanche); Caldas (Victoria); Cauca; Cesar (Manauare); Cundinamarca (Bogotá?, El Boquerón); Magdalena (Jordan – Sierra Nevada de Santa Marta); Meta (Remolino, San Martín – Llanos of Río Meta; Upper Río Negro, Villavicencio); Tolima (Carmen de Apicalá, Cunday, Honda, Payande); Valle del Cauca. **Venezuela:** Amazonas (Cacuri, Puerto Ayacucho, Río Ocano); Azoátegui (Boca del Guasey); Aragua (Choroní, Colonia Tovar, El Limón, Guamitas, La Isleta, La Victoria, Maracay, Parque Nacional Henri Pittier, San Sebastián, Turiamo, Turmero); Barinas (El Real); Bolívar (El Dorado, Quebrada de Jaspe, Río Caura, Río Maniapure, Suapure); Carabobo (El Palito, Las Trincheras, Güigüe, Quebrada Agua Clara, San Esteban); Cojedes (Río San Carlos, Río Santo Domingo); Delta Amacuro (Tucupita); Distrito Capital (Naiguatá, Río Mazare); Guárico (Calabozo, Changuango, El Socorro, Parque Nacional Aguaro – Guaripito, Río Tiznado); Isla Margarita (La Sierra); Lara (Barquisimeto, between Puente Tabla y Buria, Simón Planas, via Hacienda Montevideo – San Pedro, Yeritagua); Mérida; Monagas (Barrancas, Josepin, Maturín); Nueva Esparta (Guatamare); Portuguesa (Aparición); Sucre (Carúpano); Táchira (El Piñal, La Morita, Río Chucuri, Río Doradas, Río Frío, Río Negro, San Cristóbal); Trujillo (La Haciendita, Monay, Rafael Rangel, Valera); Yaracuy (Río Guama, Minas de Aroa, Yaritagua); Zulia (Bachaquero, road La Fría – Catacumbo, El Tucucu, La Kasmera, Lagunillas, Las Tres Bocas, Mara, Maracaibo, Misión El Rosario, Perijá, Tucuco). **Trinidad and Tobago:** Island of Tobago (Cocoa Wattie). **Guyana:** Demerara (Parika); East Berbice-Corentyne (Mt. Roraima); Essequibo Islands-West; Upper Demerara-Berbice (Omai). **Suriname:** Paramaribo. **French Guiana:** Cayenne (Camp Patawa, Caussade, Cayenne, East of Kourou, Macouria – Pointe Macouria and Delanglade, Matoury); Saint-Laurent-du-Maroni (Saint-Laurent-du-Maroni). **Brazil:** Amapá (Mazagão); Amazonas (Barcelos).

TYPES:

Papilio acastus Cramer, 1775. The original description mentions an unspecified number of specimens from Suriname. No syntypes were found in NBC or NHMUK, where many of Cramer's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Papilio acastus*, mentioned as a synonym of *Mysoria barcastus venezuelae* by Evans (1951), a female is **herein designated as the neotype**; the specimen with the following labels: / E H Jonkers Paramaribo Suriname 16-6-1971/ Museum Leiden Coll. E. H. Jonkers Acc. 1980-03/; two other labels will be added: / Neotypus/ Neotypus *Papilio acastus* Cramer, 1775 O. Mielke, Brockmann & C. Mielke det. 2015/. NBC.

Pyrrhopyga [sic] *venezuelae* Scudder, 1872. The original description mentions an unspecified number of specimens from Venezuela. To ensure the correct identification of *Pyrrhopyga venezuelae*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *venezuelae* Scudd. type/ M. C. Z. Type 16.588/ Ven[ezeu]a/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga venezuelae* Scudder, 1872, O. Mielke, Brockmann & C. Mielke det. 2019/. MCZ. Another syntype is herein designated as paralectotype; this specimen with the following labels: / Paratype *Pyrrhopyga venezuelae* Scud. det. Bates/ Ven[ezeu]a/ *venezuelae* Scudd. type/ M. C. Z. Type 16.588/; two other labels will be added: / Paralectotypus/ Paralectotypus *Pyrrhopyga venezuelae* Scudder, 1872, O. Mielke, Brockmann & C. Mielke det. 2019/. MCZ.

Mysoria decolor Mabilles & Boulet, 1908. The original description mentions two males from Costa Rica and Panama in MNHN (collection Boulet) [1862, Colombia (Bogotá – on the label of the lectotype), M. v. Lindig on the label of one syntype] and in collection Mabilles [Cauca, Colombia on the label of another syntype]. To ensure the correct identification of *Mysoria decolor*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Bogota [Cundinamarca] Colombie 1862 M. v. Lindig Museum de Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Mysoria decolor* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. Another syntype is herein designated

as paralectotype; this specimen with the following labels: / Type/ Cauca Colomb[ia]. Mabilles/ *Mys. decolor* Mab./ Ex Oberthür Coll. Brit. Mus. 1927-31/; two other labels will be added: / Paralectotypus/ Paralectotypus *Mysoria decolor* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Mysoria cayennae Mabilles & Boulet, 1908. The original description mentions ten males and five females from Honduras, Amazonas [Brazil] and French Guiana in MNHN (collection Boulet) and collection Mabilles. To ensure the correct identification of *Mysoria cayennae*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ S'. Laurent du Maroni Guyana Francaise 1905 E. Le Moutt Coll. Boulet Museum Paris/; two other labels will be added: / Lectotypus/ Lectotypus *Mysoria cayennae* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. There are eight other syntypes in MNHN with the same labels; all are herein designated as paralectotypes and two other labels will be added: / Paralectotypus/ Paralectotypus *Mysoria cayennae* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. Another syntype female in NHMUK is here also designated as paralectotype; this specimen with the following labels: / Type/ *Mysoria cayennae* M. & B. Cayenne [French Guyane] / Ex Oberthür Coll. Brit. Mus. 1927-3./; two similar labels will be added. The lectotype is illustrated in the website "Butterflies of America" (<http://butterfliesofamerica.com>).

Mysoria caucana Röber, 1925. Holotype (one male is mentioned) from Cauca Valley, Colombia. The holotype was not found in SMT or ZfB, where Röber's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Mysoria caucana*, mentioned as a synonym of *Mysoria barcastus venezuelae* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Type [of *Amenis proxima* Mabilles & Boulet, 1908]/ [Valle del] Cauca Colombie 1904 C. Ribbe Coll. Boulet Museum Paris/; two other labels will be added: / Neotypus/ Neotypus *Mysoria caucana* Röber, 1925 O. Mielke, Brockmann & C. Mielke det. 2015/. MNHN.

Mysoria barcastus pallens (Mabilles, 1891)

Pls. 52–53, Figs. 3, 13 (♀) (d, v)

DISTRIBUTION – Central-northern Colombia to southern Venezuela and northern Brazil (Roraima).

Colombia: Cundinamarca (Bogotá, El Boquerón); Santander (Barichara). **Venezuela:** Amazonas (Cacuri, Río Ocamo); Bolívar (Quebrada de Jaspe). **Brazil:** Roraima (Alto Alegre – Ilha de Maracá, Maloca da Serra do Sol, Paulo).

TYPE:

Pyrrhopyga [sic] *pallens* Mabilles 1891. The original description mentions an unspecified number of males from Brazil (**recte** Colombia – on the label of the lectotype). To ensure the correct identification of *Pyrrhopyga pallens*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ N. Granada [Colombia] v. N/ *acastus*. true!/ *Acastus* Cr. Phidias ♀ L. *Barcastus* Sepp./ *pallens* Mab./ 3: 3 * type/ Coll. Staudinger/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga pallens* Mabilles, 1891, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Mysoria barcastus alta Evans, 1951

Pls. 52–53, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Trinidad and Tobago (Trinidad) and Guyana.

Trinidad and Tobago: Arima (Arima); Rio Claro-Mayaro (Xeres Field); Port of Spain (St. Ann's Valley); San Juan-Laventille (Santa Cruz Valley); Point Fortin. **Guyana:** Upper Demerara-Berbice (Essequibo River).

TYPE:

Mysoria barcastus alta Evans, 1951. Holotype male with the following labels: / Type – *alta* Evans/ St. Anns Valley [Port of Spain] Trinidad/ Adams Bequest. B.M. 1912-399/. NHMUK.

Mysoria barcastus antila Evans, 1951

Pls. 52–53, Figs. 9 (♂), 10 (♀) (d, v)

DISTRIBUTION – Northern Peru to northern and southeastern Brazil.

Peru: Loreto (*Chasuta – Iquitos, Iquitos, Pebas, Quebrada Polis, Requena, Río Ucayali, Yurimaguas*). **Brazil:** Amazonas (Lower *Rio Madeira, Manicoré, Maués, Rio Negro, Rio Papori, São Paulo de Olivença, Tefé*); Pará (*Itaituba, Juruti, Óbidos, Rio Arapiuns, Santarém*); Maranhão (*Penalva, Peri Mirim*); Minas Gerais (*Jaíba*).

TYPE:

Mysoria barcastus antila Evans, 1951. Holotype male with the following labels: / Type – *antila* Evans/ Amazonas Santarem [Pará, Brazil] M. de Mathan I.1878/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Mysoria barcastus barta Evans, 1951

Pls. 52–53, Figs. 14, 16 (♀), 15, 17–18 (♂) (d, v)

DISTRIBUTION – Central-eastern Bolivia to north-northeastern Argentina and northeastern Brazil to Uruguay.

Bolivia: Beni (*Bueyes*); Cochabamba (*Alto Palmar, Cristal Mayu, Todos Santos*); Beni (*Santa Rosa – Río Yacuna*); La Paz (*Sarampiuni*); Santa Cruz (*Bermejo, Chiquitos, Guarayos, Roboré – Valle de Tucavaca, Warnes*). **Brazil:** Mato Grosso (*Barra do Bugres, Cáceres, Campo Novo do Parecis, Chapada dos Guimarães, Diamantino, Jaurú, Pantanal, Salobra, Urucum*); Mato Grosso do Sul (*Água Clara, Guaicurus*); Maranhão (*Imperatriz*); Goiás (*Anápolis, Chapada dos Veadeiros, Rio Araguaia, Vianópolis*); Distrito Federal (*Brasília, Planaltina*); Bahia (*Caetité – Maniaçu, Lençóis*); Minas Gerais (*Belo Horizonte, Carmo do Rio Claro, Corinto, Lassance, Paracatu, Passa Quatro, São Roque de Minas*); São Paulo (*Araçatuba, Cosmópolis, Ilha Seca, Morro Agudo, Presidente Venceslau, Rio Claro, Santana do Parnaíba, São Paulo, Tatui, Teodoro Sampaio – Parque Estadual do Morro do Diabo and Porto Cabral, Varpa*); Paraná (*Alto Paraíso, Campo Mourão, Chopinzinho, Diamante do Norte, Foz do Iguaçu, Francisco Beltrão, Ivaiporã, Jacarezinho, Jaguariaíva, Lapa, São Jorge do Ivaí, São Jorge do Patrocínio, Sulina, Toledo, Três Barras do Paraná, Vitorino*); Santa Catarina (*Joaçaba, Seara – Nova Teutônia*); Rio Grande do Sul (*Alegrete, Bagé – Palmas, Barra do Quaraí, Couto, Feliz, Frederico Westphalen, Ijuí, Iraí, Panambi, Pareci Novo, Pelotas, Porto Alegre, Porto Mauá, Salvador do Sul, Santa Rosa, Santo Augusto, São Francisco de Paula, São Luiz Gonzaga, Viamão*). **Paraguay:** Boquerón (*Filadelfia*); Caaguazú (*Coronel Oviedo*); Guairá (*Colonia Carlos Pfannl, Villarica – Mbubuevo*); Paraguari (*Sapucaí*). **Uruguay:** Artigas (*Isla de Zapallo*). **Argentina:** Chaco; Corrientes (*Santo Tome*); Entre Ríos (*Concordia*); Formosa; Misiones (*Puerto Iguazú, Uruguay – Provincial Park*).

TYPE:

Mysoria barcastus barta Evans, 1951. Holotype male with the following labels: / Type – *barta* Evans/ Paraguay Central P. Germain 1888/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Amysoria O. Mielke, 2002

One species.

DISTRIBUTION – Southern Mexico to central-northern and central Colombia and to northern Venezuela.

Amysoria galgala (Hewitson, 1866)

Pls. 52–53, Figs. 21–22 (♂), 23 (♀) (d, v)

Syn.: *Tamyris strigifera* C. Felder & R. Felder, 1867

DISTRIBUTION – Southern Mexico to central-northern and central Colombia and to northern Venezuela.

Mexico: Chiapas (*Mopastepec*). **Panama:** Chiriquí (*David*); Coclé (*Valle de Anton*); Panamá (*Isla San José*). **Colombia:** Arauca; Boyacá (*Otanche*); Caldas; Cundinamarca (*Bogotá – Llanos de San Martín*); Huila; La Guajira; Magdalena (*Santa Marta*); Meta (*Remolino, Villavicencio*); Tolima. **Venezuela:** Aragua (*Choroní, Colonia Tovar, El Limón, La Victoria, Maracay, Parque Nacional Henry Pittier – Rancho Grande, Pozo del Diablo, Villa de Cura*); Carabobo (*Dist. Bejuma, Puerto Cabello, San Esteban, Valencia*); Cojedes (*El Baúl, Río San Carlos, Río Santo Domingo*); Delta Amacuro (*Tucupita*); Distrito Capital (*Caracas – Río Mazare*); Falcón (*La Vela de Coro*); Lara (*Barquisimeto, Iribarren, Tarabana near Cabudare, via Hacienda Montevideo – San Pedro, via Barquisimeto – Duaca*); Miranda (*Valles del Tuy, Suapure*); Nueva Esparta (*Isla Margarita, La Sierra*); Sucre (*Carúpano*); Trujillo (*Valera*); Yaracuy (*Aroa*); Zulia (*Maracaibo*).

TYPES:

Pyrrhopyga [*sic*] *galgala* Hewitson, 1866. The original description mentions an unspecified number of specimens from Venezuela in collection Hewitson. To ensure the correct identification of *Pyrrhopyga galgala*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H 38/ Venezuela Hewitson Coll. 79.-69. *Pyrrhopyga galgala*. 1. – Venez[uela]/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga galgala* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Tamyris strigifera C. Felder & R. Felder, 1867. The original description mentions an unspecified number of males from Venezuela, Moritz leg., and Bogotá, Nova Granada [Colombia], Lindig leg. in collection Hewitson. To ensure the correct identification of *Tamyris strigifera*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type; Bogota [Cundinamarca, Colombia] Lindig type/ Felder Collⁿ/ *strigifera*. n./; two other labels will be added: / Lectotypus/ Lectotypus *Tamyris strigifera* C. Felder & R. Felder, 1867, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Amenis Watson, 1893

Four species.

DISTRIBUTION – Panama to northwestern Argentina and to northern Venezuela, Suriname, northeastern Brazil and Paraguay.

Amenis ponina (Herrich-Schäffer, 1869)

Pls. 52–53, Figs. 19 (♂), 20 (♀), 24 (neotype ♂ of *Pyrrhopyga ponina*) (d, v)

DISTRIBUTION – Panama to eastern Ecuador, northern Venezuela and Suriname.

Panama: Chiriquí. **Colombia.** **Venezuela:** Aragua (*Choroní, Choroní Bajo, Parque Nacional Henry Pittier – Rancho Grande*); Carabobo (*Puerto Cabello, San Esteban, Valencia*); Distrito Capital (*La Guaira*); Táchira (*Minas de Carbón*). **Ecuador:** Loja (*Loja*). **Suriname.**

TAXONOMY – A female labelled as “Origin.” (= Type) of *Pyrrhopyga* [*sic*] *ponina* in the Museum für Naturkunde (MfN) is a specimen of the “*Pyrrhopyge papius* species group” (Evans 1951) and the original description differs from this specimen. Probably the labels were swapped.

TYPE:

Pyrrhopyga [*sic*] *ponina* Herrich-Schäffer, 1869. The original description mentions an unspecified number of specimens from Tropical America. No syntypes were found in MfN or ANSP, where the Herrich-Schäffer’s types are deposited, or in any museum or collection mentioned above (see “Abbreviations of the collections”). To ensure the correct identification of *Pyrrhopyga ponina*, mentioned as *Amenis ponia ponina* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ IV-1973 Choroní bajo, Aragua, Venezuela. Romero leg./ Neotypus *Pyrrhopyga ponina* Herrich-Schäffer, 1869 O. Mielke, Brockmann & C. Mielke det. 2015/ OM 36.851/. OM-DZUP.

***Amenis picia* Evans, 1951, stat. n.**

Pls. 54–55, Figs. 1–2 (♂), 3 (♀) (d, v); Text-Figs. 51 (♂ genitalia), 52 (♀ genitalia)

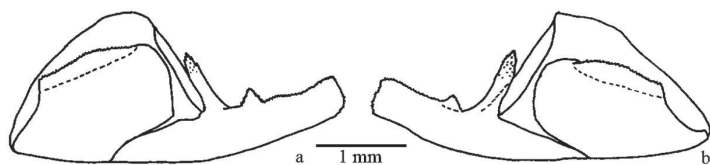
DISTRIBUTION – Western, central and eastern Colombia.

Colombia: Boyacá (*Muzo, Otanche*); Cundinamarca (*Bogotá, Silvania*); Huila (*Altamira*); Meta (*Puerto López*); Risaralda (*Belén de Umbria*); Tolima (*Aguadita, Cunday, El Santario, Giradot, Payandé, Río Chili, Villarica*).

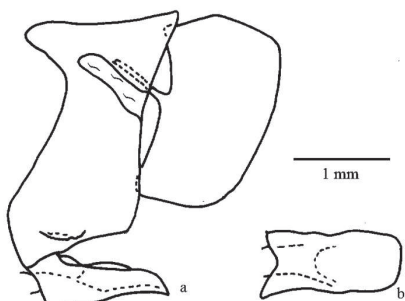
TAXONOMY – *Amenis picia* stat. n., described as a subspecies of *Amenis pionia*, is herein assigned to full species status, considering that aside from the differences in wing colouration mentioned in the original description, there are additional morphological differences in the male and female genitalia. In the male genitalia, the valvae are practically symmetrical, with the harpe of the right valva being much shorter and distally thicker (in *A. p. pionia* the valvae are clearly asymmetrical and the right valva is long and thin); the uncus is not longitudinally divided by a membranous area (in *A. p. pionia* it is divided). In the female genitalia the sterigma is distally curved ventrally (Fig. 52) (in *A. pionia pionia* it is straight – Fig. 55). The species are allopatric.

TYPE:

Amenis pionia picia Evans, 1951. Holotype male with the following labels: / Type – *picia* Evans/ Nouvelle-Grenade [Colombia] Région de Bogota reçu en décembre 1917 du frère Apollinaire-Marie/ R. Oberthür Coll. Brit. Mus. 1931-136/ NHMUK(E) #982961/ *Amenis pionia picia* Evans, 1951/. NHMUK.



Text-Fig. 51. *Amenis picia* stat. n. Male genitalia: a) right valva, internal view, b) left valva, internal view (Colombia, Tolima, Giradot, 6-I-1957, J. Foerster leg., OM 36.117).



Text-Fig. 52. *Amenis picia* stat. n. Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (Colombia, Tolima, Cunday, 14-IX-1980, C. Callaghan leg., OM 36.672).

***Amenis rogeri* Orellana, [2010]**

Pls. 54–55, Figs. 4 (♂) (d, v)

DISTRIBUTION – Northern Colombia to northwestern Venezuela.

Colombia: Norte de Santander (*Cúcuta*). **Venezuela:** Carabobo (*Valencia*); Falcón (*Estación Experimental El Cují*); Lara (*Sanare*); Mérida (*Aldea La Aguada*); Trujillo (*Pampán*); Zulia (*Mara – Represa Tule*).

TYPE:

Amenis rogeri Orellana, [2010]. Holotype male from Venezuela, Mérida, Aldea La Aguada, Pueblo Nuevo. MIZA.

***Amenis pionia* (Hewitson, 1857)**

Three subspecies.

DISTRIBUTION – Western, central and eastern Colombia,

central-eastern Bolivia, northern Venezuela, northeastern Brazil, Paraguay and northwestern Argentina.

TAXONOMY – The identification of the species is based on the original figure by Hewitson (1857) and the comments by Evans (1951). This species was described based on a least a couple, as the author described the male and female. Kirby (1879) mentioned six specimens in the Hewitson collection from Venezuela and New Granada [Colombia] and Evans (1951) accepted the original figure of a female from Venezuela as the type and, as such, characterized the species in his review of the Pyrrhopyginae. Orellana ([2010]) accepted this concept, providing figures of the male and its genitalia. This species shows a disjunct distribution, from Venezuela and Colombia and from southern Bolivia to Argentina.

The populations from the aforementioned countries Venezuela and Colombia, Bolivia, Paraguay and Argentina (*Amenis pionia pionia*), and from northeastern Brazil (*Amenis pionia pedro ssp. n.*) are isolated by more than 2,000 km from each other and the DNA barcode minimum divergence between the populations of Paraguay and Argentina (no Bolivian specimens were barcoded) (n= 2) and northeastern Brazil (n= 3) is 1.7% to 1.8%. The figures of the male genitalia of *A. pionia* in Godman & Salvin (1893 – somewhat resembling *A. rogeri*), in Bell (1933; quite different from the known taxa of *Amenis*) and in Evans (1951) are uninformative.

***Amenis pionia pionia* (Hewitson, 1857)**

Pls. 54–55, Figs. 5 (lectotype ♀ of *Amenis similis*), 6, 9–10 (♂), 11 (♀) (d, v); Text-Figs. 53–54 (♂ genitalia), 55 (♀ genitalia)

Syn.: *Amenis similis* Röber, 1925

DISTRIBUTION – Western, central and eastern Colombia to northern Venezuela, central-eastern Bolivia, Paraguay and northwestern Argentina.

Colombia: Cordillera Oriental; Cundinamarca (*Bogotá, Fusagasugá, Pandi, Viotá*); Meta; Norte de Santander (*Cúcuta*); Risaralda (*Belén de Umbria*). **Venezuela:** Aragua (*La Victoria – Quebrada Maletero, Parque Nacional Henry Pittier – Rancho Grande*); Distrito Capital (*Caracas, El Valle, Florida, Hacienda La Vega, Los Venados, Macarao*); Lara (*Sanare*); Mérida; Miranda (*Cortada del Guayabo, Los Teques*). **Bolivia:** Chuquisaca (*Carandaiti, Luis Calvo – Tigüipa*); La Paz; Santa Cruz (*Mataral, San José, Valle Grande*); Tarija (*Tarija*); Department? (*Villa Montes*). **Paraguay:** Alto Paraguay (*Palmar de las Islas*). **Argentina:** Jujuy (*Gran Brete*); Salta (*El Aybal, Embarcación, Las Higuierillas, Pocitos, Rivadavia – Santa Maria, San Lorenzo, Urundel, Yuchan*); Tucumán (*Choromoro, San Pedro Calalao, Tucumán*).

TAXONOMY – It was not possible to find differences between the specimens of the populations of Colombia and Venezuela, and Bolivia and Argentina, although they are separated by about 2,500 km. The left valva of male genitalia is very variable (see figures below). The differences between the species of this genus are in the format of the valvae, with the right valva being very long and characteristic in *A. pionia*, while the left valva shows a wide variation and no differences were found between the closely related species (i.e. those having white alar fringes). The female genitalia do not offer good characters, at least this is the impression from the few available specimens of the different *Amenis* taxa studied. The ostium is not curved ventrally in its distal portion, as in *Amenis picia* stat. n., while the specimen from Venezuela (Figs. 55a–b) is proportionally smaller than the specimen from Argentina (Figs. 55c–d), which, in turn, is even somewhat bilobed distally.

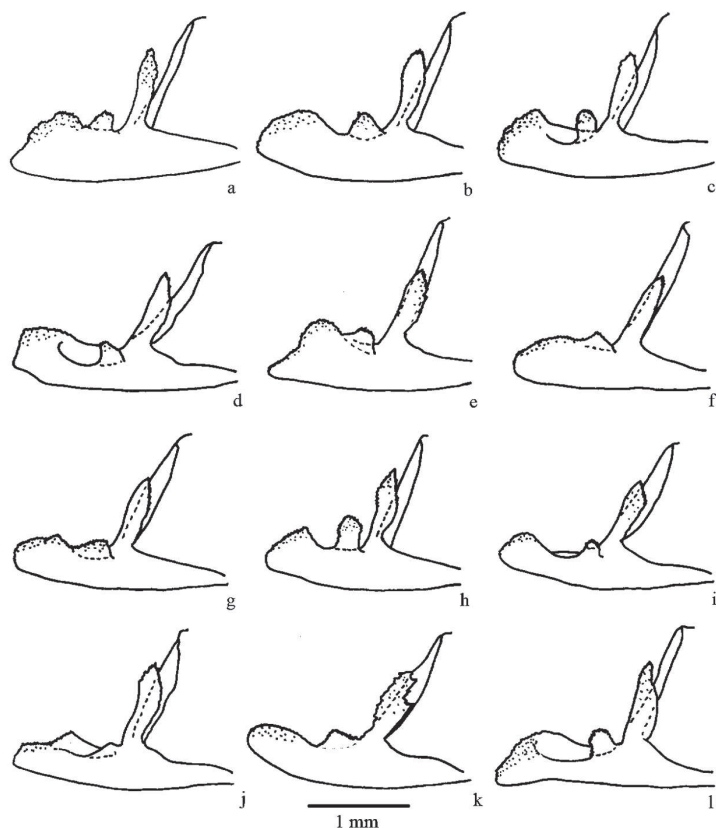
Amenis similis, for which a lectotype was designated by O. Mielke (1989a), also a female, is very similar in its colour pattern, suggesting synonymy with *A. pionia pionia*. Its genitalia were not examined.

TYPES:

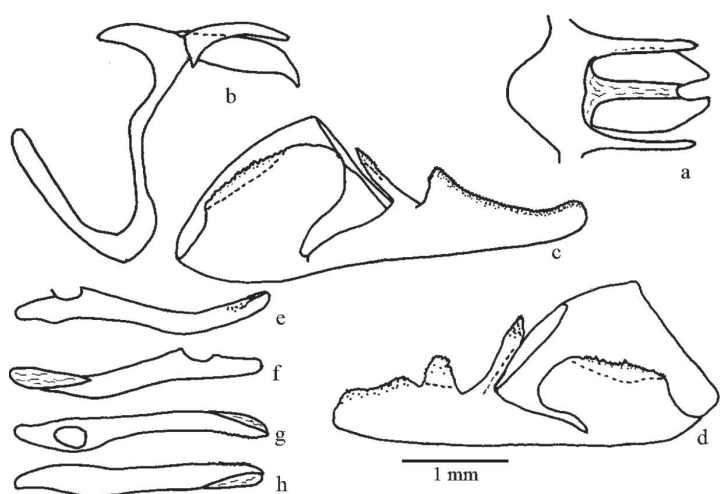
Pyrrhopyga [sic] pionia Hewitson, 1857. The original description mentions an unspecified number of specimens of males and females from New Grenada [Colombia] in collections Saunders and Hewitson. To ensure the correct identification of *Pyrrhopyga pionia*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Hewitson Coll. 79-69. *Pyrrhopyga pionia*

6/; two other labels will be added: / Lectotypus/ *Pyrrhopyga pionia* Hewitson, 1857. O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK. This specimen is figured in the original description and was designated as the "type" by Evans (1951).

Amenis similis Röber, 1925. Lectotype female (O. Mielke 1989a) with the following labels: / *Amenis similis* Röb. Venezuela/ Venezuela/ Stauding. & Bang-Haas Dresden. Ankauf 1961/ Lectotypus *Amenis similis* Roeber, 1925 O. Mielke det. 1988/ Staatl. Museum für Tierkunde Dresden/ GART specimen ID: 02584 Exemplar + Etiketten dokumentiert specimen + label data documented 2003/. SMT.

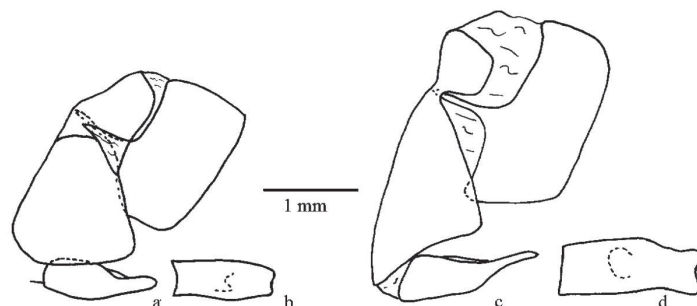


Text-Fig. 53. *Amenis pionia pionia*. Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus and uncus, lateral view; c) right valva, internal view; d) left valva, internal view; e) aedeagus, left view; f) aedeagus, right view; g) aedeagus, dorsal view; h) aedeagus, ventral view (Argentina, Salta, Pocitos, XII-1971, Fritz leg., OM 36.691).



Text-Fig. 54. *Amenis pionia pionia*. Male genitalia, variations in the left valva, apical area: a) Argentina, Salta, Pocitos, XII-1971, Fritz leg., OM 36.675; b) Bolivia, Chuquisaca, Luis Calvo, Tigüipa, I-1971, Fritz leg., OM 39.777; c) Bolivia, Chuquisaca, Luis Calvo, Tigüipa, I-1971, Fritz leg., OM 36.759; d) Bolivia, Yungas de La Paz, Steinbach leg., OM 36.739; e) Bolivia, Department ?, R. Pilcomayo to R. Grande, XII-1903, J. Steinbach leg., ex coll. NHMUK, OM 36.808; f) Bolivia, Santa Cruz, Valle Grande, Km 335 de Cochabamba, 8-II-1971, M. Fritz leg., OM 36.886; g) Argentina, Salta, El Aybal, 880

m, 22-II-2000, A. Varga leg., OM 51.450; h) Argentina, Tucumán, San Pedro de Calalao, 7-30-III-1941, OM 36.715; i) Argentina, Salta, Yuchan, 19-IX-1977, Eisele leg., OM 36.735; j) Paraguay, Alto Paraguay, Palmar de las Islas, Estancia San José, 18-IX-1994, B. Barrios leg., OM 41.053; k) Argentina, Tucumán, Choromoro, Las Huigueras, 100 m, 24-I-1970, O. Mielke leg., DZUP 1.976; l) Venezuela, Distrito Federal, Caracas, Florida, ex. coll. Gagarin, OM 10.529.



Text-Fig. 55. *Amenis pionia pionia*. Female genitalia: a) segment VIII, sterigma and papilla analis, lateral left view; b) sterigma, ventral view (Venezuela, Distrito Federal, Caracas, Río Chacaito, 980 m, 19-XII-1935, Lichy leg., OM 45.461); c) segment VIII, sterigma and papilla analis, lateral left view; d) sterigma, ventral view (Argentina, OM 36.696).

Amenis pionia pedro O. Mielke & Casagrande, *ssp.*

n.

Pls. 54–55, Figs. 7 (holotype ♂), 8 (allotype ♀) (d, v); Text-Figs. 56–57 (♂ genitalia), 58 (♀ genitalia)

DISTRIBUTION – Northeastern Brazil (Ceará and Bahia).

Brazil: Ceará (*Igatu*, *Santa Quitéria*); Paraíba (*Gurinhém* – *Chaves*, *Juazeirinho*); Bahia (*Caetité* – *Maniaçu*).

The citation *Igatu*, Ceará, Brazil is from a figure in Palo Jr. (2017), mentioned as *Amenis pionia pionia*.

DESCRIPTION – Male: forewing length 19–21 mm. Very similar to *Amenis pionia pionia*, distinguished by the blue (sometimes greenish) colour on the ventral surface of the hindwing, where the veins weakly marked by the general colouration, with the discocellular and M_2 normally unmarked, unlike *A. p. pionia* where the veins always well marked; on the ventral surface of the forewing, the spot on CuA_2-2A distinctly yellow, apparently more pronounced than in *A. p. pionia*; the outer margin of the hindwing distinctly concave between M_3-CuA_2 , unlike *A. p. pionia* with rounded wings. It also differs from *A. pionia sandra* where the blue on the dorsal and ventral faces of the forewing is greatly reduced, as well as by the very distinct veins on this surface. The left valva is very variable (Fig. 57).

Female: forewing length 22 mm. Very similar to the male, but with more elongated forewings and a less pronounced concavity of the hindwings. Genitalia with a more elongated sterigma, distally pointed in lateral view and asymmetric in ventral view.

ETYMOLOGY – The specific name is in honour of Pedro Luis dos Santos Dias, Coordinator of the Radiological Protection, Indústrias Nucleares do Brasil, Maniaçu, Caetité, Bahia, Brazil, who issued a permit to collect in this area.

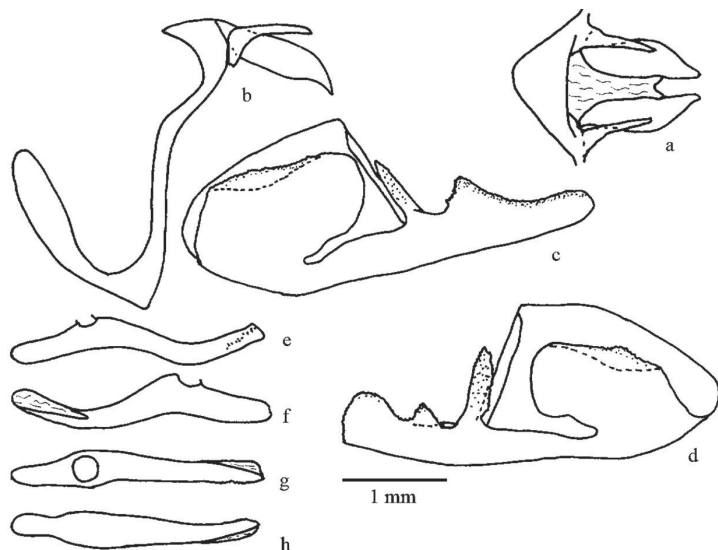
TYPES:

Holotype male with the following labels: / Holotypus/ 5-6-II-2005, 13 Km SE Maniaçu, Caetité, Bahia, [Brazil], 850 m, [O.] Mielke & Casagrande leg./ Holotypus *Amenis pionia pedro* O. Mielke & Casagrande. O. Mielke & Casagrande det. 2017/ DZ 24.624/ DZUP.

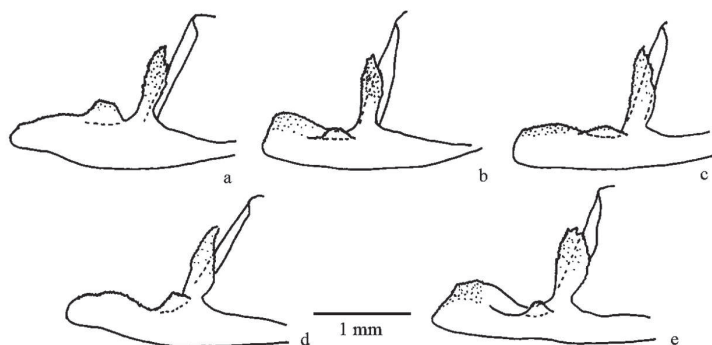
Allotype female with the following labels: / Allotypus/ 10-VII-1960, [Fazenda] Chaves, Gurinhém, Paraíba, [Brazil], Kesselring leg./ Allotypus *Amenis pionia pedro* O. Mielke & Casagrande. O. Mielke & Casagrande det. 2017/ OM 44.389/. OM-DZUP.

Paratypes: 7 ♂, same data as the holotype, DZ 8.704, DZ 8.720, DZ 22.823, DZ 24.776, DZ 24.786, DZ 24.796, DZ 24.806

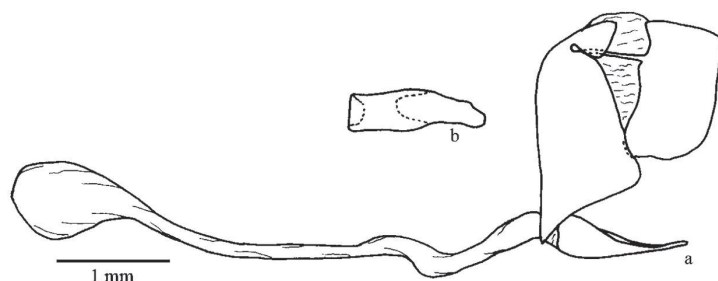
(DZUP); 1 ♂, 4-8-II-2007, 13 Km SE Maniaçu, Caetitê, Bahia, Brazil, 850 m, [O.] Mielke & Casagrande leg., DZ 24.816 (DZUP); 1 ♂, 15-VII-1959, Fazenda Chaves, Gurinhém, Paraíba, Brazil, Ebert leg., DZ 22.813 (DZUP); 1 ♂, 1-IX-1965, 1 ♂, 22-V-1967, Fazenda Chaves, Gurinhém, Paraíba, Brazil, Kesselring leg., OM 10.824, OM 36.822 (OM); 1 ♀, 25-II-1969 Juazeirinho, Paraíba, Brazil, 550 m, Ebert leg., DZ 22.933 (DZUP); 3 ♂, 24-IV-2004, Floresta, Santa Quitéria, Ceará, Brazil, Pessoa leg., OM 67.160, OM 67.167, OM 67.174 (OM); 2 ♂, 10-V-2005, 11-VI-2005, Fazenda Lambedor, Santa Quitéria, Ceará, Brazil, Pessoa leg., OM 67.505, OM 67.482 (OM).



Text-Fig. 56. *Amenis pionia pedro* **ssp. n.** Male genitalia: a) tegumen and uncus, dorsal view; b) tegumen, saccus and uncus, lateral view; c) right valva, internal view; d) left valva, internal view; e) aedeagus, left view; f) aedeagus, right view; g) aedeagus, dorsal view; h) aedeagus, ventral view (Brazil, Bahia, Caetitê, 13 Km SE Maniaçu, 5-6-II-2005, O. Mielke & Casagrande leg., paratype DZ 8.720).



Text-Fig. 57. *Amenis pionia pedro* **ssp. n.** Male genitalia, left valva, apical area: a) Brazil, Ceará, Santa Quitéria, Fazenda Lambedor, 11-VI-2005, Pessoa leg., paratype, OM 67.482; b) Brazil, Paraíba, Gurinhém, Fazenda Chaves, 15-VII-1959, Ebert leg., paratype DZ 22.813; c) Brazil, Paraíba, Gurinhém, Fazenda Chaves, 25-V-1967, Kesselring leg., paratype, OM 36.822; d) Brazil, Bahia, Caetitê, 13 km SE Maniaçu, 5-6-II-2005, [O.] Mielke & Casagrande leg., paratype DZ 8.704; e) Brazil, Bahia, Caetitê, 13 km SE Maniaçu, 850 m, 5-6-II-2005, [O.] Mielke & Casagrande leg., paratype, DZ 22.823.



Text-Fig. 58. *Amenis pionia pedro* **ssp. n.** Female genitalia: a) segment VIII, sterigma, bursa copulatrix and papilla analis, lateral left

view; b) sterigma, ventral view (Brazil, Paraíba, Juazeirinho, 550 m, 25-II-1969, Ebert leg., paratype, DZ 22.933).

Amenis pionia sandra Orellana, [2010]

Pls. 54–55, Figs. 12 (♂) (d, v); Pls. 74–75, Figs. 18 (♀) (d, v)

DISTRIBUTION – Colombia to northern Venezuela.

Colombia. Venezuela: Barinas (*Altamira de Cáceres*); Distrito Capital (*Caracas – Río Chacaito*); Táchira (*Fundación Río Negro, Río Doradas, via San Pablo*).

TYPE:

Amenis pionia sandra Orellana, [2010]. Holotype male from Venezuela, Táchira, Río Doradas. MIZA.

Mimoniades Hübner, 1823

One species.

DISTRIBUTION – Southeastern to southern Brazil.

Mimoniades ocyalus Hübner, 1823

Pls. 54–55, Figs. 22–23 (♂), 24 (♀) (d, v); Pls. 74–75, Figs. 21 (neotype ♂) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Minas Gerais (*Delfim Moreira, Passa Quatro*); Espírito Santo (*Santa Teresa*); Rio de Janeiro (*Campos, Casimiro de Abreu, Itatiaia, Nova Friburgo, Mendes, Petrópolis, Resende, Rio de Janeiro, Teresópolis*); São Paulo (*Campos do Jordão, Engenheiro Lefevre, Piquete – Barreira de Piquete, Moji das Cruzes, São Bento do Sapucaí*); Paraná (*Castro, Londrina*); Santa Catarina (*Águas Mornas – Teresópolis, Alto Rio dos Cedros, Ibirama – Barra do Prata, Joinville – Rio do Júlio, São Bento do Sul – Rio Vermelho*).

The citation **Brazil:** Amazonas (Bell 1934b) is incorrect.

TYPE:

Mimoniades ocyalus Hübner, 1823. The original description mentions an unspecified number of males from Brazil. No syntypes were found in MfN, where a few Hübner's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Mimoniades ocyalus*, mentioned as *Mimoniades ocyalus* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ 22-23-I-2004, 15 KM SE Delfim Moreira, MG [Minas Gerais, Brazil], 1500-1700 m, [O.] Mielke & Casagrande leg./ Neotypus *Mimoniades ocyalus* Hübner, 1823 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.225/. DZUP.

Mahotis Watson, 1893

Five species.

DISTRIBUTION – Colombia to central-western Argentina, central to southern Brazil and Paraguay.

Mahotis nурсcia (Swainson, 1821)

Three subspecies.

DISTRIBUTION – Western and eastern Ecuador to central-eastern Peru.

Mahotis nурсcia nурсcia (Swainson, 1821)

Pls. 54–55, Figs. 16 (neotype ♂ of *Tamyris nурсcia*), 17–18, 20 (♂), 19 (♀) (d, v)

DISTRIBUTION – Western, central and eastern Colombia to western and eastern Ecuador and to southern Peru.

Ecuador: Bolívar (*Balzapamba*); Cotopaxi (*Angamarca*); Loja (*Loja*); Manabí (*Cojimies*); Morona-Santiago (*Huamboya, Macas, Río Abanico, Zuñac*); Napo (*Baeza, Cotundo, Misahualli, Río Jondachi*); Pastaza (*Mera, Río Pastaza, Sarayacu*); Pichincha (*Pacto*); Tungurahua (*Ambato, Baños, El Topo, Hacienda la Mascota, Hacienda Santa Inés, La Palmera, Mapoto, Río Blanco, Río Margaritas, San Francisco, Río Topo, Río Ulva, Río Verde, Yunguilla, Río Zuñac*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Alto Nieva 1,930 m, Chachapoyas – Yambrasbamba, Huambo, Pomacochas, Pongo del Rentema, Rodríguez de Mendoza*); Cajamarca (*Charapi*); Huánuco (*Tingo Maria*); San Martín (*Juanjuí, Moyobamba, Rioja – San Agostin 1,000 m to Jorge Chávez 1,450 m, Roque, Santa Cruz 1,200 m, Santa Rosa del Mirador*).

The citation **Colombia:** Antioquia (*Frontino*, given as *Frontina*) (Evans 1951) seems to be incorrect; it is the type locality of *Mimoniades nurscia malis*. The citation **Venezuela** (Bell 1946) is probably incorrect (Orellana [2010]).

TYPE:

Tamyris nurscia Swainson, 1821. The original description mentions an unspecified number of specimens from South America. No syntypes were found in any museum or collection mentioned above (see “Abbreviations of the collections”). To ensure the correct identification of *Tamyris nurscia*, mentioned as *Mimoniades nurscia nurscia* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Rondos, Tingo Maria, [Huánuco] Peru, 850 m, 16-20-X-1971, Villegas leg./ Neotypus *Tamyris nurscia* Swainson, 1821 O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.205/. DZUP.

***Mahotis nurscia malis* (Godman & Salvin, 1879)**

Pls. 54–55, Figs. 13, 15 (♂), 14 (♀) (d, v)

DISTRIBUTION – Western, central and eastern Colombia to eastern Ecuador.

Colombia: Amazonian area; Antioquia (*Frontino, Medellín – La Romera*); Caquetá (*road Altamira to Florencia*); Cauca; Caldas (*Manizales, Neira, Samaná, San Juan Villalobos*); Cesar (*Serranía del Perijá*); Huila (*Pitalito*); Putumayo (*Mocoa*); Risaralda (*Pereira – Parque Ucumari, Río Otun*); Tolima (*La Marina*); Valle del Cauca (*Anchicayá, Cali, Calima, Dagua, Darién – Lago Calima, Lomitas, Villa Carmelo*). **Ecuador:** Napo (*El Reventador*); Orellana (*Road Loreto – Coca, km 15*).

The citation **Brazil:** Rio Grande do Sul (Weymer 1895) is incorrect.

TYPE:

Pyrrhopyga [sic] malis Godman & Salvin, 1879. The original description mentions an unspecified number of specimens from Frontino, [Antioquia] Colombia in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyga malis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H. 67/ Frontino. Antioquia. [Colombia], T. K. Salmon./ Godman-Salvin Coll. 1912.-23. *Mahotis malis*, G. & S./ NHMUK(E) #982972/ *Mimoniades nurscia malis* Godman & Salvin, 1879/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga malis* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Mahotis nurscia amans* (Skinner, 1920)**

Pls. 54–55, Figs. 21 (♂) (d, v)

DISTRIBUTION – Central-eastern to southern Peru.

Peru: Huancavelica (*Huancamayo*); Huánuco (*Tournavista*); Junín (*Andamarca, Chanchamayo, La Merced, Río Colorado, Río Perené, San Ramón*); La Libertad (*El Porvenir*); Madre de Dios (*Río Manu*); Pasco (*Huancabamba, Oxapampa – Parque Nacional Yanachaga Chemillén, Pozuzo, Río Palcazu*).

TAXONOMY – The original description is very clear about the orange colour of the median band of the forewing, only known in this subspecies. No such specimens were found from Colombia, only from Peru. Probably the type locality **Colombia:** [Huila] (*Neiva*) is incorrect. A similar problem was found in the same paper (Skinner

1920) in the original description of *Pyrrhopyge viriditas* from San Pedro Sula, Honduras, the true locality almost certainly being an unknown locality in Paraguay or northern Argentina.

TYPE:

Mimoniades amans Skinner, 1920. Holotype (one specimen is mentioned) male with the following labels: / Type No. 7056/ Neiva, [Huila] U. S. Col[ombia]./ *Mimoniades amans* Henry Skinner/. ANSP, transferred do CMP.

***Mahotis montana* (J. Zikán, 1938)**

Pls. 56–57, Figs. 1, 3 (♂), 2, 4 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: Minas Gerais (*Camanducaia – Monte Verde, Caparaó, Delfim Moreira, Itamonte, Passa Quatro*); Espírito Santo (*Serra do Caparaó*); Rio de Janeiro (*Itatiaia – Parque Nacional do Itatiaia, Petrópolis – Pedra Açú, Nova Friburgo*); São Paulo (*Campos do Jordão, Piquete – Barreira de Piquete*); Santa Catarina (*Bom Jardim da Serra, Lages, São Joaquim, Urubici – Morro da Igreja*); Rio Grande do Sul (*Bom Jesus, São Francisco de Paula, São José dos Ausentes*).

TYPE:

Mimoniades versicolor montana J. Zikán, 1938. Lectotype male (O. Mielke 1971) with the following labels: / Lectotipo/ Typus/ ♂ / Brazil. Espírito Santo, Campo do Caparaó, 19-II-1915, J. F. Zikán [leg.]/ Coleção J. F. Zikan/ *M. versicolor* f. *montanus* [sic], Zik[án]./ *Mimoniades montana* Zikán, 1938 [O.] Mielke det./ I.O.C., Lepidop. 33.003/. IOC.

***Mahotis versicolor* (Latreille, [1824])**

Pls. 56–57, Figs. 5 (neotype ♂ of *Mimoniades multicifer*), 6, 8 (♂), 7 (♀) (d, v)

Syn.: *Mimoniades multicifer* Hübner, [1831]

DISTRIBUTION – Central to southern Brazil, Paraguay and northern Argentina.

Brazil: Mato Grosso do Sul (*Bataguassú – Rio Guaraguatá*); Distrito Federal (*Brasília*); Minas Gerais (*Barbacena, Belo Horizonte, Cambuquira, Carmo do Rio Claro, Catas Altas – Caraça, Conceição dos Ouros, Curvelo, Delfim Moreira, Espera Feliz, Gonçalves, Itabirito, Itamonte, Juiz de Fora – Chapéu d’Uvas, Mar de Espanha, Nova Lima, Passa Quatro, Poços de Caldas, Pouso Alegre, São Roque de Minas, Três Corações, Vargem Bonita, Virgínia*); Espírito Santo (*Santa Teresa*); Rio de Janeiro (*Bocaina, Campos, Itatiaia, Macaé, Nova Friburgo, Petrópolis, Resende, Rio de Janeiro, Teresópolis*); São Paulo (*Anhembi, Araçatuba, Bananal – Bocaina, Braúna, Campinas, Campos do Jordão, Duartina, Engenheiro Lefèvre, Franco da Rocha, Interlagos, Itanhaém, Itaperi, Itararé, Jacareí, Jundiaí, Monte Alegre do Sul, Presidente Venceslau, Rio Claro, São José do Barreiro – Bocaina, São Paulo, Taubaté, Teodoro Sampaio – Parque Estadual do Morro do Diabo and Porto Cabral*); Paraná (*Arapoti, Cândido de Abreu, Castro, Cianorte, Jaguariaíva, Londrina, Pirai do Sul, Rolândia, São Jorge do Ivaí, Terra Boa, Tibagi, Tunas do Paraná, Tuneiras do Oeste, Ventania*); Santa Catarina (*Águas Mornas – Teresópolis, Dalbérgia, Joinville, Massaranduba*); Rio Grande do Sul (*Pelotas, Zona Missioneira*). **Paraguay. Argentina:** Misiones (*Puerto Iguazú*); La Rioja; Salta.

The citation **Brazil:** Pará (Evans 1951) is incorrect.

TYPES:

Hesperia versicolor Latreille, [1824]. The original description mentions an unspecified number of specimens from Brazil, Langsdorff leg. To ensure the correct identification of *Hesperia versicolor*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *H. Versicolor* Lat. Bresil/ Ex musaeo P. Mabille 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/ NHMUK(E) #982970/ *Mimoniades versicolor versicolor* Latreille, 1879 [**sic**]/; two other labels will be added: / Lectotypus/ Lectotypus *Hesperia versicolor* Latreille, [1824], O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Mimoniades mulcifer Hübner, [1831]. The original description mentions an unspecified number of specimens from Brazil. No syntypes were found in MfN, where a few Hübner's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Mimoniades mulcifer*, mentioned as a synonym of *Mimoniades versicolor versicolor* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ 13-15-II-2010 Fazenda dos Campos, Virgínia, Minas Gerais [Brazil] 1500 m, [O.] Mielke & Casagrande leg./ Neotypus *Mimoniades mulcifer* Hübner, [1831] O. Mielke, Brockmann & C. Mielke det. 2015/ DZ 33.235/. DZUP.

Mahotis eupheme* (Godman & Salvin, 1879), *stat. rest.

Pls. 56–57, Figs. 9–10 (♂) (d, v); Pls. 74–75, Figs. 19 (♀) (d, v)

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia and central-western Argentina.

Ecuador. Peru: Cusco (*Cosñipata Valley – Chontachaca 950 m to Rocotal 2,000 m, Marcapata – Carados, Quincemil*); Puno (*Río Inambari*); San Martín (*Puente Aspuzana*). **Bolivia:** Beni (*Bueyes*); Cochabamba (*Cristal Mayu*); La Paz (*Apolobamba, Caranavi, Chaco, Chulumani, Río Tamampaya, Río Zongo*); Santa Cruz (*Achira, Florida*); Tarija (*Emburazu*). **Argentina:** La Rioja; Salta (*Parque Nacional Baritú, Sala de las Pavas, Serranía de las Pavas*); Santa Fe (*San Lorenzo*).

TAXONOMY – This taxon is currently considered a subspecies of *Mimoniades versicolor*, but the male genitalia of these taxa differ largely in the shape of the valva (see Evans 1951, pl. 6) and by a DNA barcode minimum divergence of 4.3%, *Mimoniades eupheme* *stat. rest.* (n= 4) (*Mimoniades versicolor* (n= 3)) has its species status reinstated.

TYPE:

Pyrrhopyga [*sic*] *eupheme* Godman & Salvin, 1879. The original description mentions an unspecified number of specimens from Cosnipata, Peru, Whitely leg. and Apolobamba, Bolivia, Pearce leg. in collection Godman & Salvin. To ensure the correct identification of *Pyrrhopyga eupheme*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H. 69/ Cosnipata Vall., [Cusco] E. Peru. H. Whitely./ Godman-Salvin Coll. 1912.-23. *Mimoniades eupheme* G. & S./ NHMUK(E) #982971/ *Mimoniades versicolor eupheme* Godman & Salvin, 1879/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga eupheme* Godman & Salvin, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Mahotis baroni* (Godman & Salvin, 1895)**

Pls. 56–57, Figs. 11 (♂), 12 (♀) (d, v); Pls. 74–75, Figs. 20 (♂) (d, v)

DISTRIBUTION – Northwestern Peru.

Peru: Cajamarca (*Llacanora*).

REMARKS – Only known from one locality.

TYPE:

Amenis baroni Godman & Salvin, 1895. Lectotype male (Lamas & Vila 2004) with the following labels: / Type – H. 45/ Cajamarca, N. Peru. 10,000 ft. O. T. Baron./ Godman-Salvin Coll. 1912.-23. *Microceris baroni*, Schaus./ 232/ NHMUK(E) #982959/ *Amenis baroni* Godman & Salvin, 1895/. NHMUK.

***Mimadia* Grishin, 2019**

One species.

DISTRIBUTION – Colombia to central-eastern Bolivia and to French Guyana and northern to southeastern Brazil.

***Mimadia fallax* (Mabille, 1878)**

Four subspecies.

DISTRIBUTION – Western, central and eastern Colombia to central-eastern Bolivia, Venezuela to French Guiana and northern to southeastern Brazil.

TAXONOMY – *Mimadia fallax fallax* with a very narrow, or even absent, blue outer marginal band on both faces of the hindwing; *M. fallax fiska* which has a 3.5 mm wide central white band on the forewing; *M. fallax fida* is characterized by the blue outer marginal band on both faces of the hindwing being very wide at the apex, where it becomes almost white; whereas, in *M. fallax solaris* this band remains pale blue in this region. However, the exact limits of the subspecies distribution are unknown.

***Mimadia fallax fallax* (Mabille, 1878)**

Pls. 56–57, Figs. 14 (♂), 15–16 (♀) (d, v)

DISTRIBUTION – Guyana to French Guiana and northern to central Brazil.

Guyana: East Berbice-Corentyne (*New River*); Essequibo Islands – West Demerara (*River Sipu*); Upper Tukutu – Upper Essequibo (*Kamoa River, Nappi creek*). **Suriname:** Brokopondo (*Brownsberg*); Sipaliwini (*Coeroeni, Coppename, Grauboridorp*); Paramaribo (*Zanderij*); Wanica (*Lelydorp*). **French Guiana:** Cayenne (*Macouria, Montagne de Kaw, Roura*); Saint-Laurent-du-Maroni (*Ile Portal*). **Brazil:** Roraima (*Amajari – Tepequén*); Amazonas (*Barcelos, Jutai, Santa Isabel*); Pará (*Acará, Belém, Novo Progresso – Rio Azul, Rio Cumindá*); Maranhão (*Centro do Guilherme – Montes Áureos, São Luís*); Goiás.

The citations **Argentina:** Misiones (Hayward 1934 – as *Jemadia zonara*, Hayward 1948, 1973, Canals 2003) and Misiones (*Puerto Iguazú*) (Núñez B. 2009) are probably incorrect.

TYPE:

Pyrrhopyga [*sic*] *fallax* Mabille, 1878. The original description, on page 17, mentions that there is one “détérioré” specimen [male] from Brazil in the Musée Royal d’Histoire Naturelle de Bruxelles and on page 22, there is the description of the [male] and of the female, without any locality or collection information. One syntype male is in the Musée Royal d’Histoire Naturelle de Bruxelles, now Institut Royal des Sciences Naturelles de Belgique (IRSC), and another syntype female from Mabille's collection, is now in the NHMUK (Evans 1951). The syntype male is a specimen of *Jemadia gnetus* (Fabricius, 1781) and the syntype female is the species here studied. To ensure the correct identification of *Pyrrhopyga fallax*, the syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Am. Mer./ Stood in Coll. Mabille as *P. fallax*/ *Fallax* P. Mabille/ Ex musaeo P. Mabille 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/ R. Oberthür Coll. Brit. Mus. 1931-136/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga fallax* Mabille, 1878. O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Mimadia fallax fiska* (Evans, 1951)**

Pls. 56–57, Figs. 13 (♂) (d, v)

DISTRIBUTION – Western, central and eastern Colombia.

Colombia: Cauca (*Río Micay*); Cundinamarca (*Bogotá, Medina*); Meta (*Remolino, Upper Río Negro*).

TYPE:

Jemadia fallax fiska Evans, 1951. Holotype male with the following labels: / Holo-type/ Colombic [Cundinamarca] Env. Bogotá Frère Apollinaire Marie 1918/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Mimadia fallax solaris* (Hayward, 1942)**

Pls. 56–57, Figs. 17–18, 22 (♂), 19–20 (♀), 21 (neotype ♂ of *Jemadia solaris*) (d, v)

Syn.: *Jemadia fallax filta* Evans, 1951

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia, eastern Venezuela to Guyana and northern to southeastern Brazil.

Colombia: Amazonas (*Leticia*); Cundinamarca (*Bogotá*); Meta (*Ocoa*, *Upper Río Negro*, *Villavicencio*); Putumayo (*Mocoa*, *Puerto Asís*, *Putumayo*, *Río Putumayo*); Risaralda (*San Belén de Umbria*); Vaupés (*Buenos Aires*, *Taraira* – *Estación Caparus*). **Ecuador:** Pastaza (*Mera*); Sucumbíos. **Peru:** Cusco (*Cosñipata Valley* – *Chontachaca 900 m*); Loreto (*Contamana*, *Iquitos*, *Pebas*, *Picuroyacu*, *Quebrada Polis*, *San Antonio*, *Ucayali*). **Bolivia:** Cochabamba (*Chapare*); Santa Cruz (*Buenavista*). **Venezuela:** Amazonas (*Duida* – *Parque Nacional Marahuaka*, *Gavilán*, *La Esmeralda*, *Mawari-Anejidi*, *Yutajé*); Bolívar (*Cerro Guaiquinima*, *between Santa Elena de Uairén and Ikabariú*, *Ikabariú*, *Kukurital*, *La Gran Sabana*, *Las Nieves*, *Quebrada de Jaspe*, *San Juan de Manapiare*, *Santa Elena de Uairén*, *Río Surukúm*, *via San Francisco de Yuruaní to Paraitepuey*, *Wariwantey*); Sucre (*Guayana*). **Guyana.** **Brazil:** Roraima (*Amajari*); Amazonas (*Manacapuru*, *Manicoré*, *Río Papori*, *São Paulo de Olivença*, *Río Uaupés*); Pará (*Óbidos*, *Santo Antonio do Tauá*); Acre (*Upper Río Jurua*); Rondônia (*Ariquemes*, *Cacaulândia*, *Pimenta Bueno*, *Porto Velho*); Mato Grosso (*Alta Floresta*, *Brasnorte*); Maranhão (*São Luis*); São Paulo (*São José do Rio Preto*).

TYPES:

Jemadia solaris Hayward, 1942. Holotype male with the following labels: / Tipus/ São Paulo de Olivença E. do Amazonas [Brazil] Coll. E. May-Rio/ N° 60010 Proc./ *Jemadia solaris* Hayw. Holotipo ♂/. MNRJ. This specimen was lost by fire (2-IX-2018). To ensure the correct identification of *Jemadia solaris*, mentioned erroneously as a synonym of *Nosphistia zonara* (Hewitson, 1866) by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ São Paulo de Olivença, Amazonas, [Brazil]/ Neotypus *Jemadia solaris* Hayward, 1942 O. Mielke, Brockmann & C. Mielke det. 2019/ OM 37.300/. OM-DZUP.

Jemadia fallax filta Evans, 1951. Holotype male with the following labels: / Pebas Amazonas [Peru] M. de Mathan fin^{X^{brc}} & 1^{er}. Tr. 1880/ R. Oberthür Coll. Brit. Mus. 1931-136/. There is no type label. NHMUK.

Mimadia fallax fida (Evans, 1951)

Pls. 56–57, Figs. 23–24 (♂) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia.

Ecuador: Napo (*Misahualli*, *Río Anzú*, *Río Hollin*, *Río Jondachi*, *Tena*, *Tondachi*); Orellana (*road Loreto* – *Coca*); Pastaza (*Mera*, *Río Pastaza*, *San Jorge*, *Sarayacu*); Tungurahua (*Road to Palora*). **Peru:** Amazonas (*Pongo del Rentema* – *as Rentema Falls*); Cusco (*Cosñipata Valley* – *Pilcopata 550 m to San Pedro 1,400 m*, *Quincemil*); Huánuco (*Carpish*, *Cucharas*, *Tingo Maria*); Junín (*Chanchamayo*, *Perené*, *Satipo*, *Shimá*); Loreto (*Upper Río Negro* – *Arcadia and Castaña*); Madre de Dios (*Upper Río Madre de Dios* – *Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Pichis*, *Río Palcazu*); Puno (*Carabaya* – *Santo Domingo*, *Río Inambari*, *Sandia*, *Chirimayo*, *Yahuarmayo*); San Martín (*Yumbatos*, *Moyobamba*). **Bolivia:** Beni; La Paz (*Caranavi*, *Madidi*, *Parque Nacional Madidi*, *Taipiplaya*).

The citation **Ecuador:** Napo (*Río Hollin*) is from a specimen illustrated as *Phocides thermus* by Piñas & Manzano (1997).

TYPE:

Jemadia fallax fida Evans, 1951. Holotype male with the following labels: / S. Domingo, Carabaya, [Puno, Peru], 4500ft. V. VI. 01. Dry seas. (Ockenden)/ Rothschild Bequest B.M.1939-1/. There is no type label. NHMUK.

Jember Grishin, 2019

Two species.

DISTRIBUTION – Colombia to central-eastern Bolivia, northern Venezuela to Suriname, northern to southeastern Brazil and northern Argentina.

Jember scomber (H. H. Druce, 1908)

Pls. 58–59, Figs. 1 (♂) (d, v)

DISTRIBUTION – Eastern Peru to Bolivia.

Peru: Amazonas (*Pongo del Rentema* – *as Rentema Falls*); Cusco; Huánuco (*Tingo Maria*); Junín (*Satipo*); Pasco (*Pozuzo*); San Martín (*Río Mishollo* – *as Mixillo River*, *Tocache*). **Bolivia.**

The citation **Brazil** (Draudt 1921) is probably incorrect.

REMARKS –The unknown female is likely one of the largest skippers.

TYPE:

Jemadia scomber H. H. Druce, 1908. The original description mentions an unspecified number of females (**recte** males) from Pozuzo, Peru, 5000-6000ft. collected by native collectors. To ensure the correct identification of *Jemadia scomber*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H.53/ Pozuzo, [Pasco] Peru, 5000 to 6000ft Native Collector./ *Jemadia scomber* H. H. Druce Type T.E.S. 1908/ ex coll. Hamilton Druce 1919./ J.J.Joicey Coll. B.M.1925-451./ NHMUK(E) #982979/ *Jemadia scomber* Druce, 1908/; two other labels will be added: / Lectotypus/ Lectotypus *Jemadia scomber* H. H. Druce, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Jember menechmus (Mabille, 1878)

Two subspecies.

DISTRIBUTION – Western, central and eastern Colombia to central-eastern Bolivia, Venezuela to Suriname, northern to southeastern Brazil and northern Argentina.

Jember menechmus menechmus (Mabille, 1878)

Pls. 58–59, Figs. 2 (♂), 3 (♀) (d, v)

DISTRIBUTION – Western, central and eastern Colombia to central-eastern Bolivia, northern Venezuela to Suriname, northern to southeastern Brazil and northern Argentina.

Colombia: Amazonas (*Puerto Nariño*); Arauca; between Boyacá and Casanare; Cauca (*El Tambo*); Cundinamarca (*Bogotá*); Meta (*Upper Río Negro*, *Villavicencio*); Putumayo (*Mocoa*); Tolima (*El Boquerón*); Vaupés (*Mitú*). **Ecuador:** Morona-Santiago (*Macas*); Napo (*Cotundo*, *Jondachi*, *Misahualli*, *Río Anzú*, *Río Coca*, *Yanahurco*); Pastaza (*Sarayacu*); Tungurahua (*Baños*, *Río Negro*, *Río Topo*). **Peru:** Amazonas (*Cordillera del Condor* – *Falso Paquisha*, *Rodríguez de Mendoza*); Cusco (*Chontachaca 950 m*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo*, *Río Perené*, *San Ramón*, *Satipo*, *Shimá*); Loreto (*Boquerón Abad*, *Iquitos*, *Río Ampiacu*, *Río Marañon*, *Upper Río Napo*); Madre de Dios (*Upper Río Madre de Dios* – *Amazonia Lodge 500 m (near Atalaya, Cusco)*); San Martín (*Juanjuí*, *Sapasoa*, *Tocache*). **Bolivia:** Beni (*Río Beni*, *Río Yata*); Cochabamba (*Alto Palmar*); La Paz (*Caranavi*, *Río Zongo*). **Venezuela:** Barinas (*Barinitas*); Bolívar (*Kanarakuni*). **Suriname.** **Brazil:** Amazonas (*Río Uaupés*, *São Paulo de Olivença*, *Tefé*); Rondônia (*Cacaulândia*, *Cachoeira do Samuel*, *Candeias do Jamari*, *Pimenta Bueno*, *Porto Velho*); Mato Grosso (*Diamantino*, *Pantanal*); Goiás (*Chapada dos Veadeiros*); Distrito Federal (*Brasília*); Minas Gerais (*Carmo do Rio Claro*); Alagoas (*Maceió*). **Argentina:** Corrientes (*Corrientes*); Misiones (*Puerto Iguazú*).

TYPE:

Pyrrhopyga [**sic**] *menechmus* Mabille, 1878. The original description mentions an unspecified number of males in Musée Royal d'Histoire Naturelle de Bruxelles, now Institut Royal des Sciences Naturelles de Belgique (IRSC), but no syntype was found there. To ensure the correct identification of *Pyrrhopyga menechmus*, one syntype male in NHMUK is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H.52/ *menechmus*/ 190/ R. Oberthür Coll. Brit. Mus. 1931-136/ R. Oberthür Coll. Brit. Mus. 1931-136/ Stood in Coll. Mabille as *J. menechmus*/ NHMUK(E) #982980/ *Jemadia menechmus* Mabille, 1878/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga menechmus* Mabille, 1878, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

***Jember menechmus desousai* (Orellana, [2010])**

Pls. 58–59, Figs. 5 (paratype ♂), 6 (♂) (d, v)

DISTRIBUTION – Southern Venezuela.

Venezuela: Amazonas (*Maroa, San Pedro de Cataniapo, San Pedro del Orinoco, Yavita*).

REMARKS – The female is unknown.

TYPE:

Jemadia menechmus desousai Orellana, [2010]. Holotype male from Yavita, Amazonas, Venezuela, 9-VII-1953, J. A. Guzman leg. MIZA.

***Jemadia* Watson, 1893**

Four species.

DISTRIBUTION – Mexico to western Ecuador, Colombia to central-eastern Bolivia and to French Guyana to southeastern Brazil.

***Jemadia pseudognetus* (Mabille, 1878)**

Pls. 58–59, Figs. 4, 7 (♂), 8 (♀) (d, v)

Syn.: *Pyrrhopyga* [sic] *dorylas* Plötz, 1879

DISTRIBUTION – Eastern to southern Mexico to western Ecuador, northern and central Colombia, northern Venezuela to northern Brazil.

Mexico: Chiapas (*Boca de Chajul, Comitán de Domínguez, Ocosingo – San Quentin, Verapaz*); Oaxaca (*Chimalapa, Comaltepec, Metates, Palomares, Río Sarabia, Sierra de Juárez, Tuxtepec*); Tabasco (*Tenosique*); Veracruz (*Los Tuxtlas, Santa Rosa*). **Honduras:** Atlántida (*La Ceiba*); Copán (*Santa Rosa de Copán*); Cortés. **Belize:** Cayo (*Camp Sibun, San Luiz*); Stann Creek (*Middlesex*). **Guatemala:** Alta Verapaz (*Chixoy, Polochic Valley*); El Petén (*Parque Nacional Tikal*); Izabal (*Cayuga*). **Nicaragua:** Chontales. **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste*); Limón (*Puerto Viejo*); Puntarenas (*San Vito*). **Panama:** Chiriquí (*Bugaba, David, Lino*); Darién (*Caná*); Herrera (*Chepo de las Minas*); Panamá Oeste (*Cerro Campana*); Veraguas (*Calobre, Santa Fe*). **Colombia:** Boyacá (*Muzo, Otanche*); Cauca (*Popayán*); Cundinamarca (*Bogotá*); Meta (*Upper Río Negro, Villavicencio*); Santander (*Río Carare*); Tolima (*Río Chilí*); Valle del Cauca (*Río Anchicayá, Upper Cauca Valley*). **Venezuela:** Aragua (*Parque Nacional Henry Pittier – Rancho Grande*); Carabobo (*Porto Cabello*); Táchira (*Coloncito, Río Doradas, Río Negro, Río Frío, via Macanillo*); Zulia (*Perijá*). **Ecuador:** Imbabura (*Paramba*); Santo Domingo de los Tsáchilas (*Santo Domingo de los Colorados*). **Peru.** **Brazil:** Amazonas (*São Paulo de Olivença, Tefê*); Pará; Maranhão (*Centro do Guilherme – Montes Áureos*).

The citations **Brazil:** [Minas Gerais] (*Iguatama* (as *Porto Real*)) (Evans 1951) and **Argentina:** Misiones (Hayward 1973, Canals 2003) are probably incorrect.

TAXONOMY – This species is very similar to *Jemadia hospita imitator*, differing only by the dorsal outer blue band of the hindwing not crossing Sc+R₁–Rs. In Colombia, both are probably sympatric.

TYPES:

Pyrrhopyga [sic] *pseudognetus* Mabille, 1878. The original description mentions three specimens (male and female) from Peru in Musée Royal d'Histoire Naturelle de Bruxelles, now Institut Royal des Sciences Naturelles de Belgique (IRSC). Later Mabille (1908, 1912) mentioned the type locality as being Colombia; probably the first locality was a mistake. No syntypes were found in IRSC, so a specimen from Colombia found in NHMUK and identified by Mabille as *P. pseudognetus* is believed to be one syntype. To ensure the correct identification of *Pyrrhopyga pseudognetus*, this syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T/ Colombia/ *pseudognetus* Mab./ R. Oberthür Coll. Brit. Mus. 1931-136/ R. Oberthür Coll. Brit. Mus. 1931-136/ Ex musaeo P. Mabille 1923/ Stood in Coll. Mabille as *J. pseudognetus*/ NHMUK(E) #982987/ *Jemadia hospita pseudognetus* Mabille, 1878/;

two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga pseudognetus* Mabille, 1878, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [sic] *dorylas* Plötz, 1879. Holotype (one number (= one specimen) is mentioned) male with the following labels: / Holotypus/ 4930/ Porto Cabello [Carabobo, Venezuela] Edmüller/ *Dorylas* Plötz */ *Dorylas* Plötz. Stett. Ent. Zeit. 1879. p. 522/ Genitaluntersuchung No. 145 teste M. Hering. 1931/. MfN.

***Jemadia hospita* (Butler, 1877)**

Two subspecies.

DISTRIBUTION – Western and central Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, Suriname to southeastern Brazil.

***Jemadia hospita hospita* (Butler, 1877)**

Pls. 58–59, Figs. 12, 14–18 (♂), 13, 19 (♀), 20 (lectotype ♀ of *Pyrrhopyga hephaestos*) (d, v)

Syn.: *Papilio aethiops* Gmelin, [1790], **praeocc.** (Esper, 1777)

Pyrrhopyga [sic] *ulixes* Plötz, 1879

Pyrrhopyga [sic] *hephaestos* Plötz, 1879, **syn. n.**

Jemadia zamorae Mabille & Boulet, 1908, **nom. nud.** (ICZN, Art. 11.6)

DISTRIBUTION – Western Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, Suriname to southeastern Brazil.

Colombia: Caldas (*Victoria*); Guaviare (*San José del Guaviare*); Putumayo; Tolima (*El Boquerón*). **Ecuador:** Bolívar (*Balzapamba*); Chimborazo (*Dos Puentes, Riobamba*); Esmeraldas (*Chuchuví*); Loja (*Loja*); Morona-Santiago (*Huamboya, Macas, Santiago de Méndez*); Napo (*Archidona, Cotundo, Río Anzú, Río Jondachi, Tena*); Pichincha (*Los Cedros – Chontal*); Pastaza (*Sarayacu*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Quitacalzón 1,100 m*); Huánuco (*Tingo Maria, Río Cucharas*); Junín (*Chanchamayo, Perené, San Ramón, Satipo, Shimá*); Loreto (*Contamana, Iquitos, Pebas, Ucayali, Upper Putumayo*); Madre de Dios (*Parque Nacional del Manu – Pakitza, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Oxapampa, Pichis Road, Pozuzo*); Puno (*Yahuar Mayo*); San Martín (*Juanjuí, Rioja – El Afluente 1,200 m, Moyobamba, Santa Rosa del Mirador, Tarapoto, Tocache*). **Bolivia:** Cochabamba (*Cristal Mayu, Tupari*); La Paz (*Caranavi, Farinas, Mapiri, Río Zongo, Sarampiuni*); Santa Cruz (*Buenavista, Velasco*). **Suriname:** Paramaribo. **Brazil:** Roraima (*Amajari – Rio Surubai*); Amazonas (*Manaus, Rio Madeira, São Paulo de Olivença*); Pará (*Carajás, Óbidos, Santarém*); Acre (*Cruzeiro do Sul, Iquiri, Mâncio Lima – Parque Nacional da Serra do Divisor, Porto Walter, Xapuri*); Rondônia (*Ariquemes, Cacauplândia, Candeias do Jamari, Ji-Paraná, Vilhena*); Mato Grosso (*Alta Floresta, Pantanal*); Maranhão; Bahia (*Porto Seguro*); Espírito Santo (*Linhares*).

TAXONOMY – *Pyrrhopyga hephaestos* Plötz, 1879 is a variation of *J. hospita hospita* in which the blue band on the dorsal face of the hindwing is reduced to a few blue spots. Both phenotypes are sympatric, at least in **Brazil:** Rondônia (*Vilhena*) and Pará (*Óbidos*).

TYPES:

Papilio aethiops Gmelin, [1790], **praeocc.** (Esper, 1777). Lectotype male (Vane-Wright 1975) with the following labels: / 154 *Aethiops*, Gmel. Type specimen/ “and labelled accordingly by Vane-Wright”. NMI. Vane-Wright (1975) mentioned that Suriname is probably the correct type locality.

Pyrrhopyga [sic] *hospita* Butler, 1877. Lectotype male (Vane-Wright 1975) with the following labels: / Lectotype/ Type – H.48/ Peru W. Davis 77.52./ *P. hospita* Butler type/ B.M.Type No. Rh. 17306/ *Pyrrhopyga hospita* Butler det. R. I. Vane-Wright, 1972 Lectotype ♂/ NHMUK(E) #982989/ *Jemadia hospita* 184/. NHMUK.

Pyrrhopyga [sic] *ulixes* Plötz, 1879. Lectotype male (Vane-Wright 1975) with the following labels: / Type/ 4931/ Surinam Volckman/

Ulixes Plötz */ 9:16/ *Ulixes* Plötz Stett. Ent. Zeit. 1879. p. 521/ *Pyrrhopyga ulixes* Plötz det. R. I. Vane-Wright, 1972 ♂ Lectotype/. MfN.

Pyrrhopyga [**sic**] *hephaestos* Plötz, 1879. The original description mentions an unspecified number of specimens from Suriname. To ensure the correct identification of *Pyrrhopyga hephaestos*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Origin./ Surinam Prf. L.77/ 9: 25/ *Hephaestus* Möschl./ 9: 21/ *Hephaestus* Möschl./ Coll. Möschl. *E. hephastus* Möschl./ Type Verh. z. b. Ges. Wien. 1882 p. 324, Plötz. Stett. Ent. Ztg. 1879. p. 521, no: 6/ Coll. Staudinger/ Genitaluntersuchung No. 146, teste M. Hering 1931/ *Pyrrhopyga hephaestos* Plötz, det. R. I. Vane-Wright, 1972 Holotype ♀; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga hephaestos* Plötz, 1879, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN. **Syn. n.**

Jemadia zamorae Mabilles & Boulet, 1908. The name was published as a synonym of *Jemadia hephaestos* [**sic**]; (Möschler, 1883), a **nomen nudum** (ICZN, Art. 11.6).

***Jemadia hospita imitator* (Mabilles, 1891)**

Pls. 58–59, Figs. 9 (♀), 10 (lectotype ♂ of *Pyrrhopyga imitator*), 11 (♂) (d, v)

Syn.: *Jemadia paulensis* Schaus, 1902, **syn. n.**

DISTRIBUTION – Western and central Colombia to western Ecuador and northern Brazil (Amazonas).

Colombia: Boyacá (*Muzo*, *Otanche*); Caldas (*Manizales*, *Victoria*); Cauca (*Corinto*, *El Tombo*, *Pescador*); Cundinamarca (*Bogotá*); Huila (*Garzón*); Magdalena (*Ciénaga*); Putumayo; Tolima (*Honda*); Risaralda (*Pereira*); Valle del Cauca (*Bitaco*, *Buenaventura* – *Río Dagua*, *Buga*, *Calima*, *Corinto*, *Garrapatas Valley*, *Río Cali*, *Río Calima*, *Villa Carmelo*). **Ecuador:** Pichincha (*Pacto*). **Brazil:** Amazonas (*São Gabriel da Cachoeira* – *Iauareté*, *São Paulo de Olivença*?).

TAXONOMY – This subspecies is very similar to *Jemadia pseudognetus*, differing only by the dorsal outer blue band of the dorsal hindwing crossing Sc+R₁–Rs; perhaps it is a subspecies of *J. pseudognetus*. The valva of the male genitalia is very variable in *Jemadia hospita hospita* and there are no differences between *J. hospita hospita*, *J. hospita imitator* and *J. pseudognetus*.

Jemadia paulensis **syn. n.**, described from São Paulo, S. E., Brazil, probably comes from São Paulo de Olivença, Amazonas, Brazil. The original description gives São Paulo as its type locality, but on the label there is a “S. E.” [south east], probably referring to the southeastern state of São Paulo, but this seems to be incorrect. *Jemadia paulensis* **syn. n.** is considered as a new synonym of *Jemadia hospita imitator*. Evans (1951) considered this taxon as a synonym of *Jemadia hospita pseudognetus*.

TYPES:

Pyrrhopyga [**sic**] *imitator* Mabilles, 1891. The original description mentions an unspecified number of specimens from Bogotá [Cundinamarca, Colombia]. To ensure the correct identification of *Pyrrhopyga imitator*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Origin/ Coll. Staudinger/ Bogota [Cundinamarca, Colombia] Nolcken/ *Pyrr. imitator* Mab. Type/ *P. imitator* Mab./ *Hospita* Butl./ E III [??] Muzo/ 9:15.; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga imitator* Mabilles 1891, O. Mielke, Brockmann & C. Mielke det. 2019/. MfN.

Evans (1951) mentioned the type in the NHMUK, but this specimen is from Río Dagua, Buenaventura, Colombia, and has no original type label. It is a specimen of *Jemadia hospita imitator* from the collection Mabilles.

Jemadia paulensis Schaus, 1902. The original description mentions an unspecified number of types n° 5961 from São Paulo [de Olivença, Amazonas], Brazil in USNM. As in other species described by Schaus (1902), the same number is applied to more than one type specimen, so to ensure the correct identification of *Jemadia paulensis*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type No. 5961 U. S. N. M./ São Paulo S. E. Brazil/

Collection W. Schaus/ *Jemadia Paulensis* Schs Type./; two other labels will be added: / Lectotypus/ Lectotypus *Jemadia paulensis* Schaus, 1902, O. Mielke, Brockmann & C. Mielke, det. 2019/. USNM. **Syn. n.**

***Jemadia demarmelsi* Orellana, [2010]**

Pls. 58–59, Figs. 21 (holotype ♂) (d, v); Pls. 74–75, Figs. 24 (♀) (d, v)

DISTRIBUTION – Southern Venezuela.

Venezuela: Bolívar (*Gran Sabana*, *Río Surukúm*).

TAXONOMY – This species is only known from the holotype and a female specimen. The central band of the forewing is very broad, the colour of the markings is not blue like in other species of this genus, but violet.

REMARKS – Only few specimens are known.

TYPE:

Holotype male from Río Surukúm, Bolívar, Venezuela, 870 m, X-1989, F. Romero leg. (AO).

Jemadia mabilles* O. Mielke, Brockmann & C. Mielke, *sp. n.

Pls. 60–61, Figs. 1, 3–4 (paratype ♂), 2 (allotype ♀) (d, v)

DISTRIBUTION – Western, central and eastern Colombia to central-eastern Bolivia and western Venezuela.

Colombia: Boyacá (*Muzo*); Cundinamarca (*Bogotá*); Meta (*Upper Río Negro*); Putumayo (*Mocoa*); Tolima (*Río Chilli*); Valle del Cauca (*Buga*). **Ecuador:** Esmeraldas (*Chuchuvi*); Loja (*Loja*); Napo (*Muyuna*, *Tena*); Pastaza (*Mera*); Tungurahua (*Río Topo*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley* – *Pilcopata 500 m to Quebrada Quitacalzón 1,100 m, Pilcopata*); Huánuco (*Puente Cayumba, Tingo Maria*); Junín (*Chanchamayo*, *Perené*, *San Ramón*, *Satipo*, *Shimá*); Pasco (*Pozuzo*); San Martín (*Juanjui*, *Pongo de Caimarachi*, *Saposa*, *Tocache*); Ucayali (*Atalaya*). **Bolivia:** Cochabamba (*San Jacinto*); La Paz (*Apolobamba*, *Caranavi*, *Coroico*, *Río Zongo*). **Venezuela:** Táchira (*Río Frío*).

DESCRIPTION –

Jemaria [**sic**] *sosia* [misidentification]; Campos, 1927. *Rev. Col. Nac. V. Rocafuerte*, Guayaquil, 9(27–28): 98.

Jemadia sosia [misidentifications]; Bell, 1934. *Jour. N. Y. Ent. Soc.* 41: 490, figs 25 (male gen.), 37, 38 (male d, v); **syn.:** *zamorae*. – Williams & Hayward, 1944. *Acta zool. Lill.* 2: 80; **syn.:** *zamorae*, *gnetus*; Draudt. – Hayward, 1947. *Acta zool. Lill.* 4: 220; **syn.:** *zamorae*, *gnetus*; Draudt. – Evans, 1951. *Cat. Amer. Hesp.* 1, p. 52, pl. 6 (male gen.); **syn.:** *zamorae*, *hephaestus*; Plötz (female), Mabilles & Boulet. – Pallister, 1956. *Amer. Mus. Novit.* 1763: 10, 24. – Bridges, 1983. *Lep. Hesp.* 1, p. 111, 128; 2, p. 18; **syn.:** *zamorae*. – Bridges, 1988. *Cat. Hesp.* 1, p. 176, 203; 2, p. 29; **syn.:** *zamorae*. – Lamas & Grados, [1997]. *Rev. peruana Ent.* 39: 60. – [O.] Mielke, 2004. *Hesperioidea*, p. 27, in Lamas (ed.). *Checklist: Part 4A, Hesperioidea–Papilionoidea*, in Heppner (ed.). *Atlas Neotrop. Lep.* 5A; **syn.:** *zamorae*. – [O.] Mielke, 2005. *Cat. Amer. Hesperioidea* 1, p. 74; **syn.:** *zamorae*. – Orellana, [2010]. *Entomotropica* 23(3): 219, 221, figs 115, 116 (male d, v), 196 (male gen.); **syn.:** *zamorae*. – Uehara, 2016. *Konchu to Shizen* 51(11): 15.

(no genus) *sosia* [misidentification]; Beattie, 1976. *Rhop. Direct.*: 262.

This new species was recognized as *Jemadia sosia* by the above mentioned authors. *Jemadia sosia* was described from one specimen of Brazil deposited in the Musée Royal d'Histoire Naturelle, Brussels, therefore the holotype, which was examined. Evans (1951) mentioned that the type is in the BM(NH) [NHMUK], however, this specimen without data was also examined and bears a label stating that the specimen was in the Mabilles collection identified as *Pyrrhopyga* [**sic**] *sosia*. The real holotype [male] was found in the Musée Royal d'Histoire Naturelle, Brussels, now Institut Royal des Sciences Naturelles de

Belgique (IRSC), with a type label and is from Brazil. This specimen is hereby considered as the true holotype of *Jemadia hewitsonii sosia* **comb. n., stat. n.**, and *Jemadia sosia* from the abovementioned authors is described herein as *Jemadia mabillei* **sp. n.**

The description of Evans (1951) of *Jemadia sosia* is well suited for its characterization and the male specimen mentioned by him as the type of *Jemadia sosia* is herein designated as the holotype of *Jemadia mabillei* **sp. n.**

The female is very similar to the male, but has rounder wings. The forewing length has 31.3 mm. Only one female is known.

ETYMOLOGY – The specific name is in honour of Paul Mabille, one of the most important researchers of the Hesperidae.

TYPES:

The holotype male with the following labels: / Holotypus/ Type H. T./ type/ *sosial* *sosial*/ stood in Coll Mabille as *P. sosial*/ R. Oberthür Coll. Brit. Mus. 1931-186/ R. Oberthür Coll. Brit. Mus. 1931-186/ Holotypus *Jemadia mabillei* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann, C. Mielke det. 2017/. NHMUK. The first and the last labels will be added accordingly.

The allotype female with the following labels: / Allotypus/ Rio Songo [**sic**], [La Paz] Bolivia, 750 m, Coll. Fassl/ ♀/ Allotypus *Jemadia mabillei* O. Mielke, Brockmann & C. Mielke. O. Mielke, Brockmann & C. Mielke det. 2017/. ZfB. The first and the last labels will be added accordingly.

Paratypes: 3 ♂, 29-VIII-1995, 2 ♂, 1995, 1 ♂, 26-IX-1995, 1 ♂, 19-VIII-1996, Tena, Napo, Ecuador (EB); 1 ♂, 18-VIII-1997, Muyuna, Río Tena, 20 km NW of Tena, Napo, Ecuador, J. Tiefenthaler leg. (EB); 5 ♂, Tena, Napo, Ecuador, VIII-2017 (CJU); 2 ♂, 1-IX-1996, 5 ♂, 2-IX-1996, 3 ♂, 2-IV-1996, Chulva, Yacú (locality not found), Napo, Ecuador, 1000 m (EB); 1 ♂, VIII-2013, Mera, Pastaza, Ecuador, 1200 m, R. Westphal leg., OM 76.338 (OM); 11 ♂, Mera, Pastaza, Ecuador, IX-X-2012 (CJU); 1 ♂, V-2017, Chuchuvi, Esmeraldas, Ecuador, 550 m, OM 88.226 (OM); 1 ♂, 1985, Rodríguez de Mendoza, Amazonas, Peru, ex coll. Jongerling (EB); 2 ♂, Pozuzo, Pasco, Peru (EB); 28 ♂, 16-20-X-1971, 38 ♂, 27-X-1971, 27 ♂, XI-1971, Rondos, Tingo Maria, Huánuco, Peru, 850 m, Villegas leg., DZ 27.144–27.171, DZ 2.123, DZ 2.229, DZ 27.172–27.207, DZ 27.208–27.234 (DZUP); 1 ♂, XI-1964, Tingo Maria, Huánuco, Peru, DZ 26.745 (DZUP); 1 ♂, V-1972, Tingo Maria, Huánuco, Peru, 880–2200 m, DZ 26.747 (DZUP); 1 ♂, IX-1980, Pucayacu, 70 km NE Tingo Maria, Huánuco, Peru, 650 m, [collector illegible], DZ 26.745 (DZUP); 3 ♂, XI-1985, 1 ♂, 1986, 1 ♂, 14-III-1986, 1 ♂, 8-XII-1986, 1 ♂, 8-XII-1986, 1 ♂, XII-1986, 1 ♂, 15-X-1987, 1 ♂, 4-I-1987, 4 ♂, VIII-2001, Tingo Maria, Huánuco, Peru, M. Büche leg. (EB); 3 ♂, Tingo Maria, Huánuco, Peru, OM 700, OM 5.555, OM 7.663, OM 10.936 (OM); 9 ♂, XI-1964, 1 ♂, XII-1964, 5 ♂, II-1965, Tingo Maria, Huánuco, Peru, OM 763, OM 7.043, OM 7.036, OM 7.083, OM 7.039, OM 7.040, OM 7.042, OM 10.931, OM 10.935 OM 7.037, OM 7.661, OM 7.662, OM 7.664, OM 7.665, OM 7.666 (OM); 2 ♂, Tingo Maria, Huánuco, Peru, 750 m, OM 76.129, OM 77.110 (OM); 1 ♂, 12-VIII-1992, Tingo Maria, Huánuco, Peru, Büche leg., OM 35.931 (OM); 1 ♂, 23-VII-1981, Rondos, Tingo Maria, Huánuco, Peru, 650 m, [O.] Mielke & Casagrande leg., OM 37.460 (OM); 3 ♂, 19-X-1989, Río Tulumayo, 5 km SE Vitoc, San Ramón, Junín, Peru, 1150 m, [O.] Mielke & Casagrande leg., OM 23.398, OM 23.437, OM 23.512 (OM); 2 ♂, I-1981, 1 ♂, VII-1981, 1 ♂, X-1981, Río Perené, Junín, Peru, OM 34.931, OM 34.974, OM 37.464, OM 35.092 (OM); 1 ♂, X-1986, San Martín, Peru, OM 26.522 (OM); 1 ♂, VI-1989, San Martín, Peru, OM 26.899 (OM); 1 ♂, 1-IX-2012, Passaraya, Saposoa, San Martín, Peru, C. Calderón leg., OM 72.984 (OM); 1 ♂, 17-VIII-1998, 1 ♂, 1-IX-1998, 1 ♂, II-2002, Juanjuí, San Martín, Peru, ex coll. F. König (EB); 4 ♂, VI-VII-2007, Mina de Sal, Rioja, San Martín, Peru, 1900 m, ex coll. Ralph Sturm (EB); 1 ♂, Pongo de Cainarachi, San Martín, Peru (EB); 1 ♂, VIII-2017, El Afluente, Rioja, San Martín, Peru, 1300 m, 5°39'48" S, 77°41'41" W, J. Pintado leg., OM 86.585 (OM); 1 ♂, 11-XI-2012, Quebrada Quitacalzón, Cosñipata Valley, Cusco, Peru, 900–1100 m, O. Mielke leg., OM 72.974 (OM); 5 ♂, 26-29-XI-2016, 10 ♂, 4-8-XI-2017 Chontachaca, Cosñipata Valley, Cusco, Peru, 13°02' S, 71°28' W, A. Huamon V. leg., OM 82.638, OM 82.658, OM 82.668, OM 82.678,

OM 82.688, OM 84.511, OM 84.653, OM 84.663, OM 84.673, OM 84.683, OM 84.693, OM 84.703, OM 84.713, OM 84.742, OM 84.859 (OM); 1 ♂, IX-2004, Rodríguez de Mendoza, Amazonas, Peru (CJU); 1 ♂, 4-IX-1999, 1 ♂, 5-IX-2000, 6 ♂, 6-X-2014, Shimá, Junín, Peru, J. Uehara leg. (CJU); 1 ♂, VII-2013, Shimá, Junín, Peru (CJU); 4 ♂, 6-7-X-2014, Shimá, Junín, Peru, S. Kise leg. (CJU); 1 ♂, 10-V-2016, Mariposa, Junín, Peru, M. Solis leg. (CJU); 1 ♂, 29-XI-2011, Pilcopata, Cusco, Peru, J. Uehara leg. (CJU); 1 ♂, 29-XI-2011, Pilcopata, Cusco, Peru, S. Kise leg. (CJU); 1 ♂, 2003, Tingo Maria, Huánuco, Peru (CJU); 1 ♂, IX-2003, Atalaya, Ucayali, Peru (CJU); 14 ♂, IX-X-1988, Caranavi, La Paz, Bolivia, 600–1000 m, C. Tello leg., OM 20.178–20.190, OM 20.193 (OM); 17 ♂, XI-1988, Caranavi, La Paz, Bolivia, 700–1200 m (EB); 7 ♂, IX-XII-2001, Caranavi, La Paz, Bolivia, 500–1000 m (EB); 17 ♂, X-XI-2002, Caranavi, La Paz, Bolivia, 800–1700 m, C. Tello leg., OM 60.635, OM 60.522, OM 60.550, OM 60.606, OM 60.711, OM 60.747, OM 60.761, OM 60.775, OM 60.782, OM 60.789, OM 60.824, OM 60.831, OM 60.872, OM 60.921, OM 60.942, OM 60.984, OM 61.448 (OM); 1 ♂, XII-2002-I-2003, Caranavi, La Paz, Bolivia, 1100–2100 m, C. Tello leg., OM 61.250 (OM); 1 ♂, I-2003, Caranavi, La Paz, Bolivia, 900–1500 m, OM 76.659 (OM); 5 ♂, X-2004, Caranavi, La Paz, Bolivia, 1000 m, C. Tello leg., OM 60.113, OM 60.120, OM 60.147, OM 60.289, OM 66.282 (OM); 5 ♂, XI-2004, Caranavi, La Paz, Bolivia, 1000 m, C. Tello leg., OM 66.126, OM 66.133, OM 66.140, OM 66.217, OM 66.224 (OM); 1 ♂, I-2005, Caranavi, La Paz, Bolivia, 1000 m, C. Tello leg., OM 66.227 (OM); 1 ♂, 15-XI-2018, Taipiplaya, Caranavi, La Paz, Bolivia, J. Uehara leg. (CJU); 15 ♂, 16-18-XI-2018, Broncini, Caranavi, La Paz, Bolivia, J. Uehara leg. (CJU); 3 ♂, 16-XI-2018, Alianza, Caranavi, La Paz, Bolivia, J. Uehara leg. (CJU); 1 ♂, 21-XI-2018, Vagante, Coroico, La Paz, Bolivia, J. Uehara leg. (CJU); 6 ♂, San Martín, Tocache, Peru, III-2014, D. Gonzales A. leg. (EB); 1 ♂, Satipo, Peru, 10-X-2007 (EB); 3 ♂, Río Zongo, [La Paz], Bolivia, 750 m, coll. Fassl (ZfB); 4 ♂, Oberer Río Negro [upper Río Negro, Meta], Ostcolumbia [East Colombia], 800m, coll. Fassl (ZfB); 1 ♂, 1-XI-1964, Tingo Maria, Peru, coll. Hansen/Trier (ZfB).

Jematus Grishin, 2019

One species.

DISTRIBUTION – Colombia to western Ecuador, Colombia to central-eastern Bolivia, northern Venezuela to French Guiana and to southern Brazil, Paraguay and northeastern Argentina.

Jematus gnetus (Fabricius, 1781)

Three subspecies.

DISTRIBUTION – Western and central Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, northern Venezuela to French Guiana and northern Brazil.

TAXONOMY – The DNA barcode evidences the existence of two clusters with a minimum divergence of 1%, one formed by *J. gnetus gnetus* (n= 4) and the other by the remaining subspecies (n= 3).

Jematus gnetus gnetus (Fabricius, 1781)

Pls. 60–61, Figs. 5, 8–10 (♀), 6–7 (♂) (d, v)

Syn.: *Papilio vulcanus* Cramer, 1779, **praeocc.** (Fabricius, 1775)

Erycides megalesius Hübner, [1819], **nom. n.**

Jemadia suzetta Mabille & Boulet, 1908

DISTRIBUTION – Western and central Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, northern Venezuela to French Guiana and northern to southern Brazil.

Colombia: Caldas (*Victoria*); Cauca (*Popayán*); Cundinamarca (*Llanos de San Martín, Santa Maria*); East Cordillera (*east side*); Guaviare (*San José del Guaviare*); Huila (*La Plata to Tierra à Dentro*); Meta (*Puerto Balsa, Remolino, Upper Río Negro, Villavicencio*);

Putumayo (*Mocoa*). **Ecuador:** Chimborazo (*Chimbo*); Guayas (*El Salado*); Imbabura (*Lita*); Loja (*Loja*); Morona-Santiago (*Huamboya, Macas, Santiago de Méndez*); Napo (*Arosemena Tola, Cotundo, Puerto Napo, Río Challuayacu, Tena*); Pastaza (*Puyo, Sarayacu*); Pichincha (*West of Quito*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Pilcopata 550 m to Quebrada Quitacalzón 1,100 m, Marcapata, Quincemil*); Huánuco (*Río Cucharas, Tingo María, Tournavista*); Junín (*Chanchamayo, La Merced, Perené, San Ramón, Satipo, Shimá*); Loreto (*Contamana, Iquitos, Yurimaguas*); Madre de Dios (*Parque Nacional del Manu, Pebas, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Pozuzo, Puerto Bermúdez – Río Pichis*); Puno (*Río Inambari*); San Martín (*Juanjuí, Rioja, Tarapoto to Chazuta*); Ucayali (*Pucallpa*).

Bolivia: Beni (*El Cebu, Río Benicito, Río Yata*); Cochabamba (*Chapare, Tiraque*); La Paz (*Caranavi, Chulumani, Farinas, Mariapo, Río Zongo, Sarampiuni, Tumupasa*); Santa Cruz (*Buenavista*). **Venezuela:** Amazonas (*Maroa, Yavita*); Barinas (*Barinitas*); Bolívar (*Kanarakuni*).

Guyana. Suriname. French Guiana: Cayenne (*Cayenne*); Saint-Laurent-du-Maroni. **Brazil:** Amazonas (*Humaitá – 103 km East, Iranduba – Uypiranga, Manaus, Maués, São Paulo de Olivença*); Pará (*Belém, Óbidos, Santarém*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor, Porto Walter, Senador Guimard, Santa Rosa do Purus*); Rondônia (*Ariquemes, Cacaúlândia, Candeias do Jamari, Cautário, Ji-Paraná, Ouro Preto d'Oeste, Pimenta Bueno, Santo Antônio, Vilhena*); Mato Grosso (*Alta Floresta, Cáceres, Diamantino, Pantanal*); Maranhão.

The citations **Brazil:** Rio de Janeiro (*Campos*) (Bescke 1833) and (*Nova Friburgo*) (Kefenstein [1838] as *Thymele vulcanus*), **Cuba** (Lucas 1857) as *Pyrrhopyga [sic] vulcanus*, **Brazil:** Rio Grande do Sul (Weymer & Maassen 1890) as *Pyrrhopyge gnetus* are probably incorrect.

TAXONOMY – The syntype male of *Pyrrhopyga [sic] fallax* in Musée Royal d'Histoire Naturelle de Bruxelles, now Institut Royal des Sciences Naturelles de Belgique (IRSC), as mentioned in the original description, is a male of *Jematus gnetus*.

TYPES:

Papilio vulcanus Cramer, 1779. The original description mentions an unspecified number of males and females from Suriname. To ensure the correct identification of *Papilio vulcanus*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Felder Coll^N/ N° 13. *Vulcanus* Cr III 245. C. D./ NHMUK(E) #982975/ *Jemadia gnetus* syn. *vulcanus* Cramer, 1779/; two other labels will be added: / Lectotypus/ Lectotypus *Papilio vulcanus* Cramer, 1779, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK. Another syntype female is herein designated as paralectotype; this specimen with the following labels: / Calkoen Surin[ame]/ Museum Leiden/ ♀/ Museum Leiden *Jemadia suzetta* Mab. det. V. R. Altens '41/; two other labels will be added: / Paralectotypus/ Paralectotypus *Papilio vulcanus* Cramer, 1779, O. Mielke, Brockmann & C. Mielke det. 2019/. NBC.

Papilio gnetus Fabricius, 1781. The original description mentions an unspecified number of specimens from India in collection Banks, now in NHMUK. To ensure the correct identification of *Papilio gnetus*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type/ *gnetus*/ *Papilio Gnetus* Fabr. Sp. Ins. N° 624/; two other labels will be added: / Lectotypus/ Lectotypus *Papilio gnetus* Fabricius 1781, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Erycides megalisus Hübner, [1819], **nom. n.** is a new name for *Papilio vulcanus* Cramer, 1779.

Jemadia suzetta Mabilie & Boulet, 1908. Holotype (one male is mentioned) with the following labels: / Type H T – 58/ Bolivie/ *suzetta* **sp. n.**/ R. Oberthür Coll. Brit. Mus. 1931-136/ NHMUK(E) #982974/ *Jemadia gnetus* syn. *suzetta* Mabilie & Boulet, 1908. NHMUK.

Jematus gnetus brevipennis (Schaus, 1902)

Pls. 60–61, Figs. 11 (♂), 12 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil (São Paulo and Paraná), Paraguay and northeastern Argentina.

Brazil: São Paulo (*Araçatuba, Campinas, Casabranca, Mirandópolis, Rio Claro, São José do Rio Preto*); Paraná (*Jaguariaíva*).

Paraguay: Guairá (*Villarica*); Canindeyú (*Agrícola Armistício – 100 km N of Ciudad de Este*). **Argentina:** Misiones (*Pindapoy Chico*).

TAXONOMY – This subspecies, described as a species by Schaus (1902) and mentioned as a subspecies of *J. hewitsonii* by Evans (1952), received species status by Burns & Janzen (2001), and finally was combined with *Jemadia gnetus* as a subspecies by O. Mielke & Casagrande (2002). Here the subspecies status is accepted because there is allopatry, although no significant morphological differences have been found; in addition, the DNA barcoding reveals a divergence to *J. gnetus gnetus* (n= 4) of a minimum and a maximum of 1% and 1.2% (n= 2), respectively. The differences are in the reduced blue colour of the central band on the dorsal and ventral hindwing in the male to spots in the cell and between CuA₂–2A on the dorsal side and very thin band on the ventral side, forewing with a wider central semi-hyaline white spot in CuA₁–CuA₂ (longer than wider) and wider subapical and apical semi-hyaline white bands, in the nearly black palpi and the black, or nearly so, cheeks and legs; and also by a less conspicuous blue colour of the wings.

TYPE:

Jemadia brevipennis Schaus, 1902. The original description mentions an unspecified number of types n° 5962 from São Paulo, Brazil in USNM. As in other species described by Schaus (1902), the same number had been applied to more than one type specimen, so a syntype male is **herein designated as the lectotype** to ensure the correct identification of *Jemadia brevipennis*. This specimen with the following labels: / Type N° 5962 U. S. N. M./ São Paulo S. E. Brazil./ Collection W. Schaus/ *Jemadia brevipennis*, Schs type/ Genitalia X-48.47 J. M. Burns 2000/; two other labels will be added: / Lectotypus/ Lectotypus *Jemadia brevipennis* Schaus, 1902, O. Mielke, Brockmann & C. Mielke det. 2019/. USNM.

Jematus gnetus mineira O. Mielke, C. Mielke & Casagrande, **ssp. n.**

Pls. 60–61, Figs. 14 (holotype ♂), 15 (♀ allotype), 16 (paratype ♂) (d, v)

DISTRIBUTION – Central to southern Brazil.

Brazil: Distrito Federal (*Brasília*), Minas Gerais (*Belo Horizonte, Brumadinho – Casabranca, Oliveira, Rio Preto*); São Paulo (*Teodoro Sampaio – Parque Estadual do Morro do Diabo*); Paraná (*Foz do Iguaçu, Planaltina do Paraná*).

DESCRIPTION – This new subspecies is very similar to *J. gnetus gnetus*, but differing by a slightly smaller size of the forewing length: 23–25 mm – male, and 27–32 mm – female; in *J. g. gnetus* the male forewing reaches 28 mm, and in the female reaches 34 mm. The only other differences are the black, or nearly so, palpi and legs (clearly white in *J. gnetus gnetus*); the cheeks are white as in *J. gnetus gnetus*. From *J. gnetus brevipennis*, this subspecies differs by the wing pattern, being the blue and the semi-hyaline white bands similar with *J. gnetus gnetus*.

ETIMOLOGY – The subspecific name refers to Minas Gerais, the Brazilian state from where the holotype originates.

TYPES:

Holotype male with the following labels: / Holotypus/ 16-IV-2002 Casabranca, Brumadinho, MG [Minas Gerais, Brazil], 700 m, C. Mielke leg./ Holotypus *Jematus gnetus mineira* O. Mielke, C. Mielke & Casagrande. O. Mielke, C. Mielke & Casagrande det. 2019/ OM 60.837/. OM-DZUP.

Allotype female with the following labels: / Allotypus/ 10-IV-2002 Casabranca, Brumadinho, MG [Minas Gerais, Brazil], 700 m, C. Mielke leg./ BC-OM [barcoded]/ Allotypus *Jematus gnetus mineira* O. Mielke, C. Mielke & Casagrande. O. Mielke, C. Mielke & Casagrande det. 2019/ OM 60.578/. OM-DZUP.

Paratypes: 1 ♂, 13-III-1977, Fazenda Água Limpa, Brasília, Distrito Federal, Brazil, Gifford leg., DZ 24.826 (DZUP); 1 ♀, X-1977, Cabeça do Veado, Brasília, Distrito Federal, Brazil, J. Daly leg., DZ 24.836 (DZUP); 1 ♂, 26-X-1978, Cabeça do Veado, Brasília, Distrito Federal, Brazil, Gifford leg., DZ 24.846 (DZUP); 1 ♂, 23-III-2012, Córrego do Ouro, Sobradinho, Brasília, Distrito Federal, Brazil, 808 m, 5°30'50.39" S, 47°57'59.69" W, E. O. Emery leg., DZ 20.679 (DZUP); 1 ♀, no date, Belo Horizonte, Minas Gerais, F. Campos leg.,

DZ 25.095 (DZUP); 6 ♂, 16-IV-2002, Casabranca, Brumadinho, Minas Gerais, Brazil, OM 60.585, OM 60.592, OM 60.599, OM 60.613, OM 60.823, OM 60.830 (OM); 1 ♀, 16-IX-2002, Oliveira, Minas Gerais, Brazil, F. Campos leg., OM 58.386 (OM); 1 ♀, 18-XI-2017, São José do Barreiro, São Roque de Minas, Minas Gerais, Brazil, 900 m, 20°18'48.67"S, 46°33'18.55" W, C. Mielke leg., DZ 36.435 (DZUP); 1 ♂, 4-X-1928, Rio Preto, Minas Gerais, Brasil, W. Marten leg. (ZfB); 1 ♀, 22-25-X-1987, Morro do Diabo, Teodoro Sampaio, São Paulo, Brazil, 300–600 m, [O.] Mielke & Casagrande leg., OM 17.891 (OM); 1 ♀, 11-X-2009, RPPN 2 Barras, Planaltina do Paraná, Paraná, Brazil, 250 m, Carneiro, Leite, Dias & Dolibaina leg., DZ 25.440 (DZUP); 1 ♂, 9-XI-2014, Trilha do Poço Preto, Parque Nacional do Iguaçu, Foz do Iguaçu, Paraná, Brasil, R. Greve leg., DZ 36.442 (DZUP).

Jemasonia Grishin, 2019

Six species.

DISTRIBUTION – Costa Rica to western Ecuador, Colombia to French Guiana and to central-eastern Bolivia, northern to southeastern Brazil and northern Argentina.

Jemasonia suekentonmiller (Grishin, 2014)

Pls. 62–63, Figs. 1 (holotype ♂), 2 (paratype ♀) (d, v)

DISTRIBUTION – Costa Rica and Panama.

Costa Rica: Cartago (*Grano de Oro*); Guanacaste (*Área de Conservación Guanacaste*). **Panama:** Veraguas (*Calobre*).

TYPE:

Jemadia suekentonmiller Grishin, 2014. Holotype male with the following labels: / Voucher: D. H. Janzen & W. Hallwachs DB: <http://janzen.sas.upenn.edu> Area de Conservacion Guanacaste Costa Rica 04-SRNP-34396/ Legs away for DNA/ Holotype ♂ *Jemadia suekentonmiller* Grishin/. Holotype data: Costa Rica; Área de Conservación Guanacaste, Guanacaste province, Sector Pitilla, site Sendero Cuestona, 10.99455 -85.41461, 640 m, 12-VIII-204 as pupa on *Casearia arborea* (Salicaceae) by Calixto Moraga, adult eclosed 08-Sep-2004, voucher code 04-SRNP -34396. USNM.

Jemasonia ovid (Evans, 1951)

Pls. 62–63, Figs. 3 (♂), 4 (♀) (d, v)

DISTRIBUTION – Western and central Colombia and western Ecuador.

Colombia: Cauca (*Popayán*); Cundinamarca (*Bogotá*); Chocó (*Río Tamana*); Valle del Cauca (*Buenaventura* – *Río Dagua*, *Calima*). **Ecuador:** Esmeraldas (*Chuchuvi*, *Durango*, *San Francisco*); Imbabura (*Paramba*); Pichincha (*Mindo-Namibillo*, *Pacto*); Santo Domingo de los Tsáchilas (*Alluriquín*).

TYPE:

Jemadia hewitsonii ovid Evans, 1951. Holotype male with the following labels: / Type – *ovid* Evans/ Paramba [Imbabura, Ecuador] 3500'. IV.97. dry season, (Rosenberg). Crowley Bequest 1901-78./ NHMUK(E) #982977/ *Jemadia hewitsonii ovid* Evans, 1951/. NHMUK.

Jemasonia pater (Evans, 1951)

Pls. 62–63, Figs. 5–6 (♂), 7 (♀) (d, v)

DISTRIBUTION – Panama to western Ecuador, central and eastern Colombia to eastern Ecuador and northern Venezuela.

Panama: Panamá Oeste (*Cerro Campana*); Veraguas (*Santa Fe*). **Colombia:** Boyacá (*Muzo*); Caquetá (*Florencia*); Cauca (*San Andrés*); Caldas (*Manizales*); Cundinamarca (*Bogotá*); Putumayo (*Mocoa*, *Upper*

Río Mocoa); Risaralda (*Pereira*); Valle del Cauca (*Cali*, *Jiménez*); Valle del Magdalena. **Ecuador:** Manabí (*Diez de Agosto*); Napo (*Misahualli*, *Río Jondachi*); Tungurahua (*Baños*). **Venezuela:** Aragua (*Maracay?*); Sucre (*Guayana*); Táchira (*Río Doradas*, *Río Negro*, *Río Frío*); Yaracuy (*Temerla*); Zulia (*Perijá*).

The citation **Mexico** (Llorente, Luis & Vargas 1990) is probably incorrect.

TYPE:

Jemadia hewitsonii pater Evans, 1951. Holotype male with the following labels: *Jemadia hewitsonii pater* Evans, 1951. Holotype with the following labels: / Type – *pater* Evans/ Nouvelle-Grenade Région de Bogota [Cundinamarca, Colombia] reçu en décembre 1917 du frère Apollinaire-Marie/ R. Oberthür Coll. Brit. Mus. 1931-136/ BMNH(E) #982978/ *Jemadia hewitsonii pater* Evans, 1951/. NHMUK.

Jemasonia albescens (Röber, 1925), *stat. rest.*

Pls. 62–63, Figs. 8 (neotype ♂ of *Jemadia albescens*), 10, 12 (♂), 11, 13 (♀) (d, v); Text-Fig. 59 (♂ genitalia)

DISTRIBUTION – Southern Colombia, western and eastern Ecuador to central-eastern Bolivia.

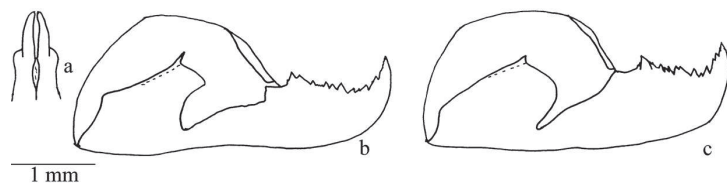
Colombia: Putumayo (*Mocoa*). **Ecuador:** Chimborazo (*Riobamba*); Esmeraldas (*Chuchuvi*); Loja (*Loja*); Morona-Santiago (*Macas*); Napo (*Río Napo*); Orellana (*Coca*); Pastaza (*Canelos*, *Río Pastaza*, *Sarayacu*); Tungurahua (*Ambato*, *Hacienda El Rosario*, *Río Topo*, *Río Verde*); Zamora-Chinchipe (*Baños*, *Parque Nacional Podocarpus*, *Zamora*). **Peru:** Amazonas (*Chachapoyas* – *Yambrasbamba*, *Rodríguez de Mendoza*); Cajamarca (*Charapi*); Cusco (*Aguas Calientes*, *Cosñipata Valley* – *Pilcopata 550 m to Mirador 1,700 m*, *Machu Picchu*, *Quincemil*); Huánuco (*Boquerón del Padre Abad*, *San Ramón*, *Tingo Maria*); Junín (*Andamarca*, *Chanchamayo*, *San Ramón*, *Satipo*, *Shimá*); Madre de Dios (*Upper Río Madre de Dios* – *Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Huancabamba*, *Pozuzo*); Puno (*Río Inambari*); San Martín (*Jorge Chávez*, *Juanjuí*, *Rioja* – *El Afluente 1,200 m to Jorge Chávez 1,450 m*, *Moyobamba*, *Santa Rosa del Mirador*, *Tocache*). **Bolivia:** Beni (*Bueyes*); Cochabamba (*Chapare*, *Cristal Mayu*, *Yungas del Palmar*); La Paz (*Apolobamba*, *Caranavi*, *Chulumani*, *Coroico*, *Río Mapiri*, *Río Zongo*).

TAXONOMY – The DNA barcode present a minimum divergence of 4.6% (n= 3) to *Jemasonia hewitsonii hewitsonii* (n= 2). The male genitalia differs from this species by the shape of the bifid uncus, being the two projections narrower and closer to each other in dorsal view, and by the valva with a well pointed terminal and dorsal projection (Fig. 59); in addition, they are sympatric at least in Tingo Maria, Huánuco, Pozuzo, Pasco, **Peru**, and Caranavi and Mapiri, La Paz, Bolivia, reason for the species reinstated status.

The wingspan of the holotype mentioned in the original description is 62 mm, this is a relative very large *Jemadia* specimen, only found by us in Tingo Maria, Huánuco, Peru or Caranavi, La Paz, Bolivia. As the holotype was not located, it is preferred to designate a neotype from Ecuador, the type locality, with a shorter wingspan (51 mm).

TYPE:

Jemadia albescens Röber, 1925. Holotype (one male is mentioned) from Macas, [Morona-Santiago], Ecuador. The holotype was not found in SMT or ZfB, where the Röber's types are deposited, or in any museum or collection mentioned above (see "Abbreviations of the collections"). To ensure the correct identification of *Jemadia albescens*, mentioned as *Jemadia hewitsonii albescens* by Evans (1951), a male is **herein designated as the neotype**; the specimen with the following labels: / Neotypus/ Coca Upp[er]. R[io]. Napo, [Ecuador] V.-VII.1899 (W. Goodfellow./ gen[itá]lia. prep[arada]., [O.] Mielke 2012/ Rothschild Bequest B.M.1939-1./ ex coll[ection]. Brit[ish] Mus[eum]. Nat[ural]. Hist[ory]./ Neotypus *Jemadia albescens* Röber, 1925 O. Mielke, Brockmann & C. Mielke det. 2015/. OM 72.148/. OM-DZUP. Specimen from the duplicate collection of NHMUK, examined by Evans (1951) and presented by R. I. Vane-Wright to OM in 1976.



Text-Fig. 59. *Jemasonia albescens* **stat. rest.** Male genitalia: a) uncus tip, dorsal view (Peru, Huánuco, Tingo Maria, Rondos, 850 m, 25-XI-1971, Villegas leg., DZ 26.934); b) right valva, internal view (same specimen); c) right valva, internal view (Ecuador, Napo, Río Coca, V-VII-1899, W. Goodfellow leg., OM 72.148 (neotype)).

Jemasonia hewitsonii (Mabille, 1878)

Two subspecies.

DISTRIBUTION – Panama to western and central Colombia, eastern Colombia to central-eastern Bolivia, northern Venezuela to French Guiana, northern and southeastern Brazil and northern Argentina.

TAXONOMY – This species is polytypic and quite variable in its blue colour, as well as in the width of both dorsal and ventral blue bands. The morphology of the genitalia is quite variable, as the harpe (Fig. 60) can end bluntly, sharply or with a dorsally folded distal point, almost the same size as in *J. albescens* **stat. rest.**, *J. ortizi* and *J. pateri*; the dorsal ornamentation of the harpe is equally variable.

Jemasonia hewitsonii hewitsonii (Mabille, 1878)

Pls. 62–63, Figs. 9, 14, 16, 18 (♂), 15, 17, 19–20 (♀) (d, v);
Text-Fig. 60 (♂ genitalia)

Syn.: *Jemadia le cerfi* Mabille & Boulet, 1908; incorr. orig. spelling (ICZN, Art. 32.5)

Jemadia lecerfi; Mabille, 1912; correct spelling.

DISTRIBUTION – Panama to western and central Colombia, eastern Colombia to southeastern Peru, Guyana to French Guiana and northern Brazil.

Panama: Veraguas (*Calobre*). **Colombia:** Caquetá (*Río Orteguaza*); Cundinamarca (*Bogotá*); Valle del Cauca (*Anchicayá*). **Ecuador:** Morona-Santiago (*Río Shankaimé*). **Peru:** Loreto (*Iquitos, Pebas, Río Marañon, Ucayali, Yurimaguas*); Madre de Dios (*Puerto Maldonado*); San Martín (*Rioja – Pedro Ruiz*); **French Guiana:** Cayenne (*Pointe Macouria, Tonate*). **Guyana.** **Suriname:** Brokopondo (*Brownsberg*); Wanica (*Domburg*). **Brazil:** Amazonas (*Benjamin Constant, Iranduba – Uypiranga, São Paulo de Olivença*); Pará (*Rio Arapiuns, Juruti, Óbidos, Santarém, Santo Antonio do Tauá*); Rondônia (*Cacaulândia*).

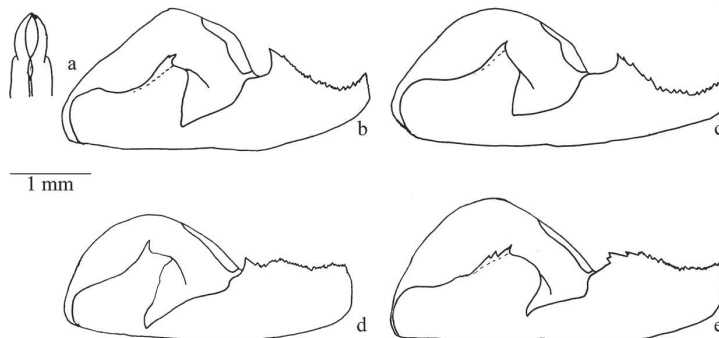
TAXONOMY – This subspecies is characterized by the narrower forewings than in the other subspecies (the ratio of forewing length to its width – from the middle of the costa to the tornus – is about 1.9, while in the other subspecies is 1.8); by the semi-transparent central band of the forewing being narrower (3–4.5 mm); by the dorsal blue submarginal band of the hindwing being generally reduced in its width and length; and by the less intense blue colour on the dorsal face.

TYPES:

Pyrrhopyga [**sic**] *hewitsonii* Mabille, 1878. This name is for the figure of *Pyrrhopyga vulcanus*; Hewitson, 1857 (misidentification), described from Brazil [Amazonas – on the label of the lectotype]. To ensure the correct identification of *Pyrrhopyga hewitsonii*, one syntype male figured by Hewitson (1857) is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H 54/ Amazons [Brazil]. Hewitson Coll. 79.-69. *Pyrrhopyga vulcanus*. 1./ 193/ R. Oberthür Coll. Brit. Mus. 1931-136/ NHMUK(E) #982978/ *Jemadia hewitsonii hewitsonii* Mabille, 1878/; two other labels will be added: / Lectotypus; Lectotypus *Pyrrhopyga hewitsonii* Mabille, 1878, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Jemadia le cerfi Mabille & Boulet, 1908. The original description mentions two males and one female from French Guiana in MNHN. To ensure the correct identification of *Jemadia le cerfi*, a male syntype is

herein designated as the lectotype; this specimen with the following labels: / Type/ Type/ Gourdonville [Cayenne] Guyane Francaise 15-9-1905 G. Le Moullet Coll. Boulet Museum Paris/ *J. Le Cerfi* Mab. et Boulet. Ann. Sc. N. 197 (1909)/; two other labels will be added: / Lectotypus/ Lectotypus *Jemadia le cerfi* Mabille & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN.



Text-Fig. 60. *Jemasonia hewitsonii hewitsonii*. Male genitalia: a) uncus tip, dorsal view (Brazil, Pará, Óbidos, II-1969, OM 35.917); b) right valva, internal view (same specimen); c) right valva, internal view (Brazil, Amazonas, São Paulo de Olivença, OM 15.009); d) right valva, internal view (Brazil, Pará, Santo Antonio do Tauá, Reserva Sonho Azul, 8-XII-2006, P. Jauffret leg., DZ 24.966); e) right valva, internal view (Brazil, São Paulo, Araçatuba, 25-XI-1950, A. Gentile leg., OM 8.555).

Jemasonia hewitsonii sosia (Mabille, 1878), **comb. n., stat. n.**

Pls. 62–63, Figs. 22 (♂), 23–24 (♀) (d, v); Pls. 64–65, Figs. 1, 5 (♂), 2 (♀) (d, v)

Syn.: *Jemadia lisetta* Mabille & Boulet, 1908, **syn. n.**

DISTRIBUTION – Eastern Colombia to central-eastern Bolivia, northern Venezuela to southeastern Brazil and northern Argentina.

Colombia: Meta (*Villavicencio*). **Ecuador:** Morona-Santiago (near *Macas, Santiago de Méndez*); Napo (*Río Anzú, Río Challuayacu*); Pastaza (*Sarayacu*); Tungurahua (*Baños, El Tópo, Lapabrenor or La Palmera, Yunguilla*). **Peru:** Cusco (*Chontachaca 950 m*); Huánuco (*Cucharas, Tingo Maria*); Junín (*Río Colorado, Río Perené, Shimá*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado, Salvación, Tambopata Reserve*); Pasco (*Pozuzo*); Puno (*Sandia*). **Bolivia:** Beni (*Trinidad*); Cochabamba (*Chapare*); La Paz (*Caranavi, Mapiri, Puente Villa*); Santa Cruz (*Buenavista, Potrerillo del Guenda, Santa Cruz de La Sierra, Sara*). **Venezuela:** Amazonas (*Campamiento Turístico Yutajé*); Bolívar (*Jabillal, Tumeremo*); Carabobo (*Puerto Cabello*). **Brazil:** Roraima (*Alto Alegre*); Amazonas (*Eirunepé, Manicoré, Maués*); Pará (*Igarapé dos Índios – Rio Cuminá, Juruti, Santo Antônio do Tauá*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor, Porto Walter, Senador Guiomard*); Rondônia (*Ariquemes, Cacaulândia, Candeias do Jamari, Pimenta Bueno, Porto Velho, Vilhena*); Mato Grosso (*Alta Floresta, Brasnorte, Chapada dos Guimarães – Buriti, Comodoro, Cuiabá, Diamantino, Pantanal*), Mato Grosso do Sul (*Bataguassú, Nioaque*); Goiás (*Chapada dos Veadeiros*); Distrito Federal (*Brasília, Planaltina*); Alagoas (*Maceió*); Minas Gerais (*Pitangui, São Roque de Minas*); Rio de Janeiro (*Casimiro de Abreu, Rio de Janeiro, Silva Jardim*); São Paulo (*Araçatuba*). **Argentina:** Santiago del Estero.

TAXONOMY – *Pyrrhopyga* [**sic**] *sosia* was described from a single specimen (holotype) which was examined in the collection of Musée Royal d'Histoire Naturelle de Bruxelles, now Institut Royal des Sciences Naturelles de Belgique (IRSC). Evans (1951) and probably all references of subsequent authors to *Jemadia sosia* are of the herein described *Jemadia mabillei* **sp. n.**

Pyrrhopyga sosia is a subspecies of *Jemasonia hewitsonii* and differs from *Jemadia hewitsonii hewitsonii* by having wider wings (the ratio of forewing length by its width – from the middle of the costa to the tornus – is about 1.8); by the semi-transparent central band of the forewing being wider (3.9–4.4 mm); by the apical spots normally being

slightly wider; by the distinct presence of a dorsal blue submarginal band on the hindwing; by the ventral marginal band on the hindwing usually being slightly wider; as well as, by the more intense blue colour. As *Pyrrhopyga hewitsonii* and *Pyrrhopyga sosia* were described by the same author and in the same publication, we hereby propose that the name *Pyrrhopyga hewitsonii* has “precedence” over *Pyrrhopyga sosia*.

Jemadia lisetta Mabilles & Boulet, 1908 is a new synonym of *Pyrrhopyga sosia* Mabilles, 1878. Evans (1951) mentioned that the type of *Jemadia lisetta* is in the BM(NH) [NHMUK], however, this specimen of the Mabilles collection, a male of *Jemasonia hospita hospita* (Butler, 1877), is not one syntype, considering that it does not have any information about locality, which is categorically mentioned in the original description.

TYPES:

Pyrrhopyga [sic] sosia Mabilles 1878. Holotype male with the following labels: / Type/ *Pyrrhopyga sosia* P. Mab. Bresill/ *P. Sosia* ? P. Mab/ *Jemadia hewitsoni [sic]* Mab. Det. Fr. J. Ball 1926/. IRSC.

Jemadia lisetta Mabilles & Boulet, 1908. The original description mentions three males from Peru and one male from Bolivia in MNHN and collection Mabilles. To ensure the correct identification of *Jemadia lisetta*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Perou 1904 O. Staudinger Coll. Boulet Museum Paris/ *J. lisetta* Mab. et Boulet. Ann. Sc. Nat. p. 197 (1909)/ *Jemadia lisetta* M. & B. det. R. I. Vane-Wright 1972 Syntype ♂/; two other labels will be added: / Lectotypus/ Lectotypus *Jemadia lisetta* Mabilles & Boulet, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. MNHN. **Syn. n.**

***Jemasonia ortizi* (Orellana, [2010])**

Pls. 62–63, Figs. 21 (♂) (d, v)

DISTRIBUTION – Central Colombia to northwestern Venezuela.

Colombia: Cundinamarca (*Bogotá*). **Venezuela:** Mérida (*Río Muchachí*); Táchira (*Campamento Siberia, Río Negro, Río Doradas – Presa Las Cuevas, Río Frío*).

TYPE:

Jemadia ortizi Orellana, [2010]. Holotype male from Campamento Siberia, Táchira, Venezuela, 1100 m, X-1998, A. Orellana & R. Manrique leg. AO.

***Nosphistia* Mabilles & Boulet, 1908**

One species.

DISTRIBUTION – Colombia to Bolivia, southern Venezuela, Suriname and northern Brazil.

***Nosphistia zonara* (Hewitson, 1866)**

Pls. 64–65, Figs. 3, 6–7 (♂), 4, 8 (♀) (d, v)

Syn.: *Pyrrhopyga [sic] perplexus* Mabilles, 1878

Jemadia gigantea Röber, 1925

DISTRIBUTION – Western, central and eastern Colombia to Bolivia, southern Venezuela, Suriname and northern Brazil.

Colombia: Cundinamarca (*Bogotá*); Meta (*Río Guyabero, Villavicencio*); Valle del Cauca. **Ecuador:** Loja; Napo (*Río Arajuno, Río Napo*); Pastaza (*El Partidero – on the Río Anzú, Mera, Puyo, Río Pastaza*); Tungurahua (*Baños, Hacienda La Mascota, La Merced – on the Río Pastaza, Palora, Río Topo*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo, La Merced, Río Paucartambo*); Loreto (*Iquitos, San Antonio*); Madre de Dios (*Parque Nacional del Manu, Tambopata Reserve*); Pasco (*Oxapampa, Puerto Inca*); San Martín (*Rioja – El Afluente 1,300 m to Jorge Chávez 1,450 m, Santa Cruz 1,200 m*). **Bolivia. Venezuela:**

Amazonas (*La Esmeralda*). **Suriname. Brazil:** Amazonas (*Manicoré, Tefé, São Paulo de Olivença, Rio Jutai, Upper Rio Uaupés*); Pará (*Óbidos*); Rondônia (*Jamari, Vilhena*).

The citations **Argentina:** Misiones (Hayward 1941, 1973, Canals 2003) are incorrect.

TYPES:

Pyrrhopyga [sic] zonara Hewitson, 1866. The original description mentions an unspecified number of males from Amazonas, [Brazil]. To ensure the correct identification of *Pyrrhopyga zonara*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type – H. 65/ Amazonas. [Brazil] Hewitson Coll. 79–69. *Pyrrhopyga zonara*. 2/ 182/ NHMUK(E) #982985/ *Nosphistia zonara* Hewitson, 1866/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga zonara* Hewitson, 1866, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Pyrrhopyga [sic] perplexus Mabilles, 1878. The original description mentions an unspecified number of males, without data in Musée Royal d'Histoire Naturelle de Bruxelles, now Institut Royal des Sciences Naturelles de Belgique (IRSC). No types were found in IRSC, but found in NHMUK and MfN. To ensure the correct identification of *Pyrrhopyga perplexus*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H. 66/ Brasilia; *Perplexus* P. Mab./ Ex musaeo P. Mabilles/ R. Oberthür Coll. Brit. Mus. 1931–136/ NHMUK(E) #982986/ *Nosphistia zonara* Hewitson, 1866/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga perplexus* Mabilles, 1878, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK. Another syntype male is herein designated as paralectotype; this specimen with the following labels: / Origin./ *perplexus* Mab. ♂/ Chanchamayo [Junín, Peru] Thamm 85./ Coll. Staudinger/; two other labels will be added; / Paralectotypus *Pyrrhopyga perplexus* Mabilles, 1878, O. Mielke, Brockmann & C. Mielke, det. 2019/. MfN.

Jemadia gigantea Röber, 1925. Holotype (one female is mentioned) with the following labels: / *gigantea*/ Holotypus *Jemadia gigantea* Roeber, 1925, O. Mielke det. 1988/ Stauding. & Bang-Hass Ankauf 1961/ Staatl. Museum für Tierkunde Dresden/ GART specimen ID: 02560 Exemplar + Etiketten dokumentiert specimen + label data documented 2003/. SMT.

MICROCERISINA Grishin, 2019

Eleven genera.

TAXONOMY – The subtribe is characterized by the genome; no morphological characters are known (Grishin in Zhang *et al.* 2019).

***Croniades* Mabilles, 1903**

Two species.

DISTRIBUTION – Colombia to central-eastern Bolivia, French Guiana to northern Brazil.

***Croniades auraria* H. H. Druce, 1908, stat. rest.**

Pls. 60–61, Figs. 13, 17 (♂), 18 (♀) (d, v); Text-Fig. 61 (♂ genitalia)

DISTRIBUTION – Central-eastern Peru to central-eastern Bolivia.

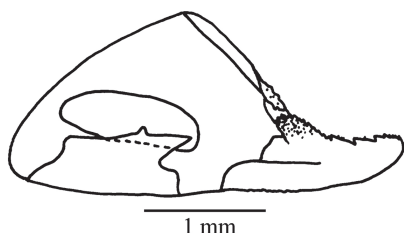
Peru: Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to Quebrada Santa Isabel 1,200 m*); Junín (*Río Perené, San Ramón*); Pasco (*Pozuzo*). **Bolivia:** La Paz (*Caranavi, Farinas, Incahuara 1,500 m, Río Zongo*).

TAXONOMY – *Croniades auraria* **stat. rest.** was considered a subspecies of *C. pieria* (Evans 1951), but here the status is reinstated considering the differences in the male genitalia: harpe with a slight dorsally curved distal tip, its basal dorsal spine thick and the sacculus with two disto-dorsal spines, while in *C. pieria* with a straight harpe, its

basal dorsal spine more slender and the sacculus with only one disto-dorsal spine.

TYPE:

Croniades auraria H. H. Druce, 1908. The original description mentions an unspecified number of males from Farinas, La Paz, Bolivia, 1500 m in collection Druce. To ensure the correct identification of *Croniades auraria*, one syntype male is **herein designated as the lectotype**; this specimen with the following labels: / Type/ Farinas La Paz Bolivia 1,500 m./ *Croniades auraria* H. H. Druce 1908, TYPE T.E.S./ ex coll. Hamilton Druce, 1919./ J.J.Joicey Coll. B.M. 1925.-451./; two other labels will be added: / Lectotypus/ Lectotypus *Croniades auraria* H. H. Druce, 1908, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.



Text-Fig. 61. *Croniades auraria* **stat. rest.** Male genitalia: right valva, internal view (Bolivia, La Paz, Río Zongo, OM 11.974).

Croniades pieria (Hewitson, 1857)

Pls. 60–61, Figs. 19–20, 24 (♀), 21–23 (♂) (d, v); Text-Fig. 62 (♂ genitalia)

Syn.: *Elbella zesta* Evans, 1951

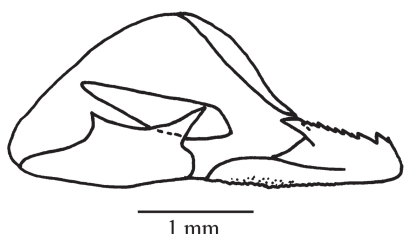
DISTRIBUTION – Southern Colombia to southeastern Peru, French Guiana to northern Brazil.

Colombia: Putumayo (*Mocoa*). **Ecuador:** Esmeraldas (*Durango*); Napo (*Archidona*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cosñipata Valley – Chontachaca 950 m*); Loreto (*Iquitos*); Madre de Dios (*Parque Nacional del Manu*); San Martin (*Moyobamba, Tocache – Polvora*). **French Guiana:** Cayenne (*Macouria – Sablance, Montagne des Chevaux*). **Brazil:** Amazonas (*Maués, São Paulo de Olivença, Tefé*); Pará (*Altamira, Belém, Juruti, Óbidos, Santarém, Rio Tapajós*); Acre (*Marechal Thaumaturgo*); Mato Grosso (*Alta Floresta*).

TYPES:

Pyrrhopyga [**sic**] *pieria* Hewitson, 1857. The original description mentions an unspecified number of females from Rio Amazonas, [Brazil] in collection Saunders & Hewitson. To ensure the correct identification of *Pyrrhopyga pieria*, one syntype female is **herein designated as the lectotype**; this specimen with the following labels: / Type H T – H.81/ Amazons. [Brazil] Hewitson Coll. 79.-69. *Pyrrhopyga pieria*. 2. – Amaz./ 284/; two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga pieria* Hewitson, 1857, O. Mielke, Brockmann & C. Mielke det. 2019/. NHMUK.

Elbella zesta Evans, 1951. Holotype female with the following labels: / Type – *zesta* Evans/ [French] Guyane/ ex coll. G. Dallier 1925/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.



Text-Fig. 62. *Croniades pieria*. Male genitalia: right valva, internal view (Brazil, Pará, Tapajós, OM 33.148).

Pseudocroniades O. Mielke, 1995

One species.

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Pseudocroniades machaon (Westwood, 1852)

Two subspecies.

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Pseudocroniades machaon machaon (Westwood, 1852)

Pls. 64–65, Figs. 9, 12 (♀), 10–11 (♂) (d, v)

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Minas Gerais (*Conceição dos Ouros, Passa Quatro, Poços de Caldas*); São Paulo (*Salesópolis, São Paulo*); Paraná (*Balsa Nova, Campina Grande do Sul, Cândói, Castro, Chopinzinho, Curitiba, Guarapuava, Guaratuba – Castelhos 750–800 m, Lapa, Mangueirinha, Morretes – Alto da Serra 800 m, Morretes – Morro Alto 300 m, Piên, Ponta Grossa, Quatro Barras, São José dos Pinhais, Tibagi, Tunas do Paraná, Turvo*); Santa Catarina (*Águas Mornas – Teresópolis, Campo Alegre, Corupá, Curitiba, Dalbergia, Joinville, Massaranduba, Mafra, Monte Castelo, São Bento do Sul, Taió*); Rio Grande do Sul (*Northeastern region of the state, Panambi, Pelotas, Vacaria, Zona Missioneira*). **Paraguay.** **Argentina:** Misiones.

TYPE:

Ericides [**sic**] *machaon* Westwood, 1852. Lectotype male (O. Mielke 1995) with the following labels: / Type/ Type H T/ Brazil/ Brazil Stevens. 51.-III./ Lectotipo/ *Ericides machaon* Westwood, 1852, Lectótipo, O. Mielke det. 1993/ *Pseudocroniades machaon machaon*, O. Mielke det. 1993/. NHMUK.

Pseudocroniades machaon seabrai O. Mielke, 1995

Pls. 64–65, Figs. 13 (holotype ♂) (d, v)

DISTRIBUTION – Southeastern Brazil (Rio de Janeiro).

Brazil: Rio de Janeiro (*Rio de Janeiro – Alto da Boa Vista*).

REMARKS – Only known from two specimens from the type locality.

TYPE:

Pseudocroniades machaon seabrai O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ 20-II-1971 Alto da Boa Vista, Rio [de Janeiro] [Estado do Rio de Janeiro, Brasil], Pe. Moure leg./ DZ 2037/ *Pseudocroniades machaon seabrai* [O.] Mielke, Holótipo, O. Mielke det. 1993/. DZUP.

Parelbella O. Mielke, 1995

Five species.

DISTRIBUTION – Mexico to central-eastern Bolivia, southern Venezuela, French Guiana, northern to southern Brazil, Paraguay and northeastern Argentina.

Parelbella ahira (Hewitson, 1866)

Two subspecies.

DISTRIBUTION – Eastern Colombia to central-eastern Bolivia, southern Venezuela, French Guiana, northern to southern Brazil, Paraguay and northeastern Argentina.

***Parelbella ahira ahira* (Hewitson, 1866)**

Pls. 64–65, Figs. 14 (lectotype ♀ of *Pyrrhopyga ahira*), 15 (♂), 16 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *jamina* Butler, 1870

Pyrrhopyga [**sic**] *zimra* Hewitson, 1871

DISTRIBUTION – Eastern Colombia to central-eastern Bolivia, southern Venezuela, French Guiana and northern Brazil.

Colombia: Guaviare (*San José del Guaviare*); Meta (*Villavicencio*). **Ecuador:** Sucumbíos (*Cerro Lumbaqui Norte*). **Peru:** Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m to San Pedro 1,400 m*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo, La Merced, Perené, Río Colorado, San Ramón, Shimá*); Loreto (*Nauta*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado*); San Martín (*Juanjui*). **Bolivia:** La Paz (*Caranavi, Yungas*); Santa Cruz (*Buenavista, Santa Cruz de La Sierra, Sara*). **Venezuela:** Bolívar (*Montecal, Suapure*). **French Guiana:** Cayenne (*Cayenne, Montagne de Kaw*); Saint-Laurent-du-Maroni. **Brazil:** Amazonas (*Manaus, São Paulo de Olivença, Tefê*); Pará (*Belém, Benevides, Óbidos, Santarém, Santo Antonio do Tauá*); Acre (*Senador Guiomard*); Rondônia (*Cacaulândia*); Mato Grosso (*Diamantino*); Maranhão (*Feira Nova do Maranhão*).

TYPES:

Pyrrhopyga [**sic**] *ahira* Hewitson, 1866. Lectotype female (O. Mielke 1995) with the following labels: / Type – H. 61/ Lectotype/ Pará/ Pará [Brazil]. Hewitson Coll. 79-69 *Pyrrhopyga ahira*. 1./ Lectotipo/ *Pyrrhopyga ahira* Hewitson, 1866, Lectótipo, O. Mielke det. 1993/ *Parelbella ahira ahira*, O. Mielke det. 1993/ NHMUK(E)# 982983/. NHMUK.

Pyrrhopyga [**sic**] *jamina* Butler, 1870. Lectotype female (O. Mielke 1995) with the following labels: / Type/ Druce Coll. Ex. Kaden Coll./ *P. Jamina* Butl. Type/ Godman-Salvin Coll. 1912.-23./ Lectotipo/ *Pyrrhopyga jamina* Butler, 1870, Lectótipo, O. Mielke det. 1993/ *Parelbella ahira ahira*, O. Mielke det. 1993/. NHMUK.

Pyrrhopyga [**sic**] *zimra* Hewitson, 1871. Lectotype female (O. Mielke 1995) with the following labels: / Type/ Type H T/ Hewitson Coll. 79.-69. *Pyrrhopyga jamina*. 1/ Lectotipo/ *Pyrrhopyga zimra* Hewitson, 1871. Lectótipo, O. Mielke det. 1993/. NHMUK.

***Parelbella ahira extrema* (Röber, 1925)**

Pls. 64–65, Figs. 17 (neotype ♂ of *Jemadia extrema*), 18 (♀) (d, v)

DISTRIBUTION – Central to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Mato Grosso (*Pantanal*); Distrito Federal (*Brasília*); Minas Gerais (*Paracatu, São Roque de Minas*); São Paulo (*Campinas, Morro Agudo, Teodoro Sampaio – Parque Estadual do Morro do Diabo and Porto Cabral*); Paraná (*Diamante do Norte, Foz do Iguaçu, Guairá, Rolândia, Terra Rica*). **Paraguay:** Cordillera (*San Bernardino*); Central (*Molinaspá, Nueva Itália*); Guairá (*Itapé, Villarica*); Paraguari (*Sapucaí*). **Argentina:** Misiones (*General Belgrano*).

TYPE:

Jemadia extrema Röber, 1925. Neotype male (O. Mielke 1995) with the following labels: / Neotipo/ 22-25-X-1987 Morro do Diabo, Teodoro Sampaio, S[ão]. P[aulo]. [Brazil] 300–500 m, [O.] Mielke & Casagrande leg./ 17.875/ *Jemadia extrema* Roeber, 1925, Neótipo, O. Mielke det. 1993/ *Parelbella ahira extrema* Roeber, 1925, O. Mielke det. 1993/. DZUP.

***Parelbella peruana* O. Mielke, 1995**

Pls. 64–65, Figs. 19 (holotype ♂), 20 (allotype ♀) (d, v); Pls. 74–75, Figs. 8 (♀) (d, v)

DISTRIBUTION – Southeastern Peru to central-eastern Bolivia.

Peru: Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m*); Junín (*Aldea, Chanchamayo, San Ramón*). **Bolivia:** La Paz (*Incahuara 1,300–1,500 m, Puente Villa*).

TYPE:

Parelbella peruana O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ Chanchamayo [Junín] E. Peru 1000 m/ gen[italia]. prep[arada]., [O.] Mielke 1993/ OM 33.182/ *Parelbella peruana* [O.] Mielke, Holótipo, O. Mielke det. 1993/. OM-DZUP.

***Parelbella polyzona* (Latreille, [1824])**

Pls. 64–65, Figs. 21 (♂), 22 (♀) (d, v)

DISTRIBUTION – Southeastern and southern Brazil.

Brazil: Espírito Santo (*Linhares*); Rio de Janeiro (*Rio de Janeiro*); São Paulo (*Caraguatatuba*); Santa Catarina (*Joinville*).

The citations **Colombia:** Amazonian area (Andrade-C. *et al.* 2015) and **Guyana** (Palo Jr. 2017) are incorrect; perhaps misidentifications of *Parelbella ahira ahira*.

TYPE:

Hesperia polyzona Latreille, [1824]. Holotype (one male is mentioned) with the following labels: / Type/ According to Olaf Mielke (*in litt.* 1970) the probable type loc. is S. Brazil, det. R. I. Vane-Wright 1976/ R. Oberthür Coll. Brit. Mus. 1931-136/ *H. polyzona*. (*scriptit.* Latreille)/ *H. polyzona* Lat. *vulcanus* Cr. Bres./ NHMUK.

***Parelbella macleannani* (Godman & Salvin, 1893)**

Pls. 64–65, Figs. 23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Southwestern and eastern Mexico to western Ecuador.

Mexico: Chiapas (*San Cristóbal*); Oaxaca (*Chimalapa, Istmo de Tehuantepec, Río Sarabia*); Veracruz (*Catemaco, Dos Amates, Los Tuxtlas*). **Guatemala:** El Petén (*Sayaxché*). **Honduras:** Cayo (*Camp Sibun*). **Nicaragua.** **Costa Rica:** Alajuela (*San Ramón*); Guanacaste (*Área de Conservación Guanacaste*); Puntarenas (*Osa*). **Panama:** Chiriquí (*Bugaba*). Colón (*Gamboa, Lion Hill*); Herrera (*Cerro Alto Higo, Chepo de La Minas*); Los Santos (*Cerro El Montoso*); Panamá (*Cerro Galera, El Llano, Madden Forest*); Panamá Oeste (*Cerro Campana*); Veraguas (*Calobre*). **Colombia.** **Ecuador:** Imbabura (*Paramba*); Pichincha (*San Miguel de los Bancos*).

TYPE:

Jemadia macleannani Godman & Salvin, 1893. Lectotype male (O. Mielke 1995) with the following labels: / Type/ Sp. figured/ Calobre, [Veraguas] Panama, Arcé/ B.C.A. Lep. Rhop. *Jemadia macleannani*, G. & S./ Godman-Salvin Coll. 1912.-23./ Lectotipo/ *Jemadia macleannani* Godman & Salvin, 1893, Lectótipo, O. Mielke det. 1993/ *Parelbella macleannani*, O. Mielke det. 1993/. NHMUK.

***Parelbella nigra* O. Mielke, Austin & Warren, 2008**

Pls. 66–67, Figs. 1 (paratype ♂), 2 (paratype ♀) (d, v)

DISTRIBUTION – Southwestern and eastern Mexico.

Mexico: Oaxaca (*San José Chiltepec, Santiago Comaltepec – Puerto Eligio, Totontepec Villa de Morelos*); Veracruz (*Presidio*).

TYPE:

Parelbella nigra O. Mielke, Austin & Warren, 2008. Holotype male with the following labels: / T. Escalante Totontepec [Villa Morelos] Oax[aca, Mexico] VII-50/ A. C. Allyn Acc 1973-48/ MGCL FLMNH Specimen no. 40755/ Genitalic Vial GTA-13765/ Holotype *Parelbella nigra* [O.] Mielke Austin & A. Warren/. MGCL.

***Ochropyge* O. Mielke, 2002**

One species.

DISTRIBUTION – Central-western to southern Brazil and northeastern Argentina.

***Ochropyge ruficauda* (Hayward, 1932)**

Pls. 66–67, Figs. 3–4 (♂) (d, v)

Syn.: *Pyrrhopyge excelsus* Bell, 1947

DISTRIBUTION – Central-western to southern Brazil and northeastern Argentina.

Brazil: Mato Grosso (*Vila Bela da Santíssima Trindade*); Paraná (*Castro, Foz do Iguaçu, Londrina, Morretes – Alto da Serra 800 m*); Santa Catarina (*Joinville*). **Argentina:** Misiones.

TYPES:

Pyrrhopyge ruficauda Hayward, 1932. Holotype male with the following labels: / Typus/ Misiones [Argentina]/ 44943/ *Pyrrhopyge* Hbn. *ruficauda* Hayw. Holotipo 3/ 4/ 32 K. J. Hayward/. MABR.

Pyrrhopyge excelsus Bell, 1947. Holotype male with the following labels: / Holotypus/ Londrina Parana, Brazil 14-9-1909/ G1755/ *Pyrrhopyge excelsus* Bell Holotype ♂/. AMNH.

***Protellabella* O. Mielke, 1995**

One species.

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia and Suriname, northern to central Brazil.

***Protellabella alburna* (Mabille, 1891)**

Two subspecies.

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia, Suriname, French Guiana and northern to central Brazil.

***Protellabella alburna alburna* (Mabille, 1891)**

Pls. 66–67, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia and Suriname.

Ecuador: Morona-Santiago (*Santiago de Méndez*); Pastaza (*Mera*). **Peru:** Amazonas (*Cordillera del Condor, Río Santiago*); Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m, Marcapata Valley*); Huánuco (*Monzón, Tingo Maria*); Junín (*Aldea, Chanchamayo, Shimá*); Madre de Dios (*Parque Nacional del Manu, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Pozuzo*); Puno (*San Juan del Oro*); San Martín (*Nueva Jordania, Tarapoto, Yumbatos*). **Bolivia:** Cochabamba (*Alto Palmar, Cristal Mayu*); La Paz (*Caranavi*). **Suriname:** Sipalivini (*Voltzberg*).

The citation **Brazil:** Paraná (Bell 1934a) from a specimen in the AMNH is incorrect.

TYPE:

Pyrrhopyga [**sic**] *alburna* Mabille, 1891. Lectotype male (O. Mielke 1995) with the following labels: / Origin./ Lectotipo/ Coll. Staudinger/ Chanchamayo [Junín, Peru] Thamm/ *Alburna* n.sp. (Mab.)/ *P. alburna* Mab./ *Alburna* Mab./ *Pyrrhopyga alburna* Mabille, Lectotipo, O. Mielke det. 1993/ *Protellabella alburna alburna*, O. Mielke det. 1993/. MFN.

***Protellabella alburna brasiliensis* O. Mielke, 1995**

Pls. 66–67, Figs. 7 (holotype ♂), 8 (allotype ♀) (d, v)

DISTRIBUTION – Northern Peru, French Guiana and northern to central Brazil.

Peru: Loreto (*Iquitos, Nauta*). **French Guiana:** Cayenne (*Montagne des Chevaux – Galion*). **Brazil:** Amazonas (*Tefé*); Acre (*Senador Guimard*); Rondônia (*Alto Paraíso, Cacauplândia, Pimenta Bueno, Vilhena*); Mato Grosso (*Diamantino, Sinop*); Goiás (*Pirenópolis*).

TYPE:

Protellabella alburna brasiliensis O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ Pimenta Bueno, Rondônia [Brazil] VIII-IX-1970/ DZ 1899/ *Protellabella alburna brasiliensis* [O.]

Mielke, Holotipo, O. Mielke det. 1993/. DZUP.

***Elbella* Evans, 1951, stat. rev.**

Six species.

DISTRIBUTION – Mexico to southern Brazil, Paraguay and northeastern Argentina.

TAXONOMY – This genus was synonymized to *Microceris* by Zhang *et al.* (2019), but is here revalidated. Although the genome of the species of *Elbella* **stat. rev.** shows some similarities with that genus, all species included here in *Elbella* **stat. rev.** remain well segregated by the morphology from the closely related *Microceris variicolor*, the only species here included. In addition, the divergence time is similar to that of *Merobella*, another genus accepted by Zhang *et al.* (2019).

All species of *Elbella* **stat. rev.** present in the male genitalia a valva with a very long and curved harpe and an unexpanded costa, and in the female genitalia the sterigma is a simple ring. In *Microceris*, the valva bears a very short harpe and a dorsally well expanded costa, and in the female genitalia the sterigma is tubular. See figures in O. Mielke (1995).

***Elbella intersecta* (Herrich-Schäffer, 1869), comb. rev.**

Five subspecies.

DISTRIBUTION – Panama to western Ecuador, eastern Colombia to central-eastern Bolivia, Venezuela, Suriname, French Guiana, northern to southern Brazil and northeastern Argentina.

***Elbella intersecta intersecta* (Herrich-Schäffer, 1869), comb. rev.**

Pls. 66–67, Figs. 9–14 (♂), 15 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *fluminis* Butler, 1873

Pyrrhopyga [**sic**] *semidentata* Mabille, 1877

Pyrrhopyga [**sic**] *tiribazus* Plötz, 1879, **nom. nud.** (ICZN, Art. 11.6)

DISTRIBUTION – Panama to western Ecuador, eastern Colombia to central-eastern Bolivia, southern Venezuela, Suriname, French Guiana and northern Brazil.

Panama: Bocas del Toro (*Chiriquí Grande*). **Colombia:** Cauca; Cesar (*Atanquez*); Meta (*Upper Río Negro, Villavicencio*); Valle del Cauca (*Buenaventura – Río Dagua*). **Ecuador:** El Oro (*El Pasaje*); Esmeraldas (*Chuchivi*); Loja (*Loja*); Morona-Santiago (*Macas, Santiago de Méndez*); Napo (*Archidona, Cotundo, Hotel Jaguar, Napo, Puerto Misahualli, Río Coca, Río Napo, Tena, Yanahurco*); Pastaza (*Mera, Sarayacu*); Tungurahua (*Hacienda La Mascota, Río Topo*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cusco (*Cashiriari, Cosñipata Valley – Pilcopata 550 m to Chontachaca 950 m, Quincemil, Río Urubamba*); Huánuco (*Cucharas, Tingo Maria*); Junín (*Chanchamayo, La Merced, Perené, Río Colorado, San Ramón, Satipo, Shimá*); Loreto (*Achinamiza, Balsapuerto, Iquitos, Pebas, Río Putumayo, Río Ucayali, Upper Río Putumayo*); Madre de Dios (*Parque Nacional del Manu, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco), Tambopata Reserve*); Pasco (*Pichis Road*); San Martín (*Juanjuí, Tarapoto, Yumbatos*). **Bolivia:** La Paz (*Caranavi, Mapi, Parque Nacional Madidi, Río Chimate*); Santa Cruz (*San Ignacio de Velasco*). **Venezuela:** Amazonas (*Cerro de la Neblina, Río Siapa*). **Suriname:** Paramaribo (*Charlesburg*). **French Guiana:** Cayenne (*Camp Patawa, Cayenne, Montagne de Kaw*); Saint-Laurent-du-Maroni (*Saint-Laurent-du-Maroni*). **Brazil:** Amazonas (*Ipixuna, Manicoré, Maués, Parintins, Rio Madeira, São Gabriel da Cachoeira, São Paulo de Olivença*); Pará (*Itaituba, Juruti, Óbidos, Santarém*); Acre (*Mâncio Lima – Parque Nacional da Serra do Divisor, Santa Rosa do Purus*); Rondônia (*Cacauplândia, Ouro Preto d'Oeste, Pimenta Bueno, Vilhena*); Mato Grosso (*Comodoro, Diamantino, Rondonópolis*); Maranhão (*Imperatriz*).

TYPES:

Myscelus intersecta Herrich-Schäffer, 1869. Lectotype male (O. Mielke 1995) with the following labels: / Origin./ Coll. H.-Sch./ *intersecta* HS/ *intersecta* HS/ *Intersecta* H.-Sch./ Coll. Staudinger/ Lectotipo/ *Myscelus intersecta* Herrich-Schäffer, 1869, Lectótipo, O. Mielke det. 1993/ *Elbella intersecta intersecta*, O. Mielke det. 1993/. MfN.

Pyrrhopyga [*sic*] *fluminis* Butler, 1873. Lectotype male (O. Mielke 1995) with the following labels: / Type/ Type/ Braz[il]. Santarem [Pará, Brazil]/ Santarem Bates 52-96/ Lectotipo/ *Pyrrhopyga fluminis* Butler, 1873, Lectótipo, O. Mielke det. 1993/ *Elbella intersecta intersecta*, O. Mielke det. 1993/. NHMUK.

Pyrrhopyga [*sic*] *semidentata* Mabilie, 1877. Lectotype male (O. Mielke 1995) with the following labels: / Type/ *Pyrrhopyga semidentata* P. Mab. Perou, Nouv. Grenade/ *semidentata* P. Mab./ [illegible]/ Lectotipo/ *Pyrrhopyga semidentata* Mabilie, 1877, Lectótipo, O. Mielke det. 1993/ *Elbella intersecta intersecta*, O. Mielke det. 1993/. IRSC.

Pyrrhopyga [*sic*] *tiribazus* Plötz, 1879. The name was published as a synonym of *Pyrrhopyga* [*sic*] *fluminis*, a *nomen nudum* (ICZN, Art. 11.6).

Elbella intersecta losca* Evans, 1951, *comb. rev.

Pls. 66–67, Figs. 17 (♂), 18 (♀) (d, v)

Syn.: *Elbella intersecta ilona* Evans, 1951, *syn. n.*

DISTRIBUTION – Central to southern Brazil and northeastern Argentina.

Brazil: Mato Grosso (*Chapada dos Guimarães*); Mato Grosso do Sul (*Corumbá*, *Nioaque*, *Pantanal*); Goiás (*Campinaçu*, *Rio Araguaia*, *Vianópolis*); Distrito Federal (*Brasília*, *Planaltina*); Minas Gerais (*Carmo do Rio Claro*, *Corinto*, *Curvelo*, *Paraopeba*, *Santana do Riacho* – *Serra do Cipó*, *São Roque de Minas*, *Sete Lagoas*); São Paulo (*Araçatuba*, *Campinas*, *Rio Claro*, *São José do Rio Preto*, *São Paulo*, *Teodoro Sampaio* – *Parque Estadual do Morro do Diabo and Porto Cabral*); Paraná (*Foz do Iguaçu*, *São Pedro do Ivaí*). **Argentina:** Misiones (*General Belgrano*).

TAXONOMY – *Elbella intersecta ilona* Evans, 1951 is a new synonymy of *Elbella intersecta losca* *comb. rev.* The differences mentioned by Evans (1951) for *E. intersecta ilona* are variations of this taxon. The red collar, patagia and a little red at the base of specimens, and the bronze colour of dorsal wings of *E. intersecta losca* *comb. rev.* are characteristic.

TYPE:

Elbella losca Evans, 1951. Holotype male with the following labels: / Type – *losca* Evans/ Jan./ Biol.C.A.Lep.Rhop. *Pyrrhopyge menecrates*, Mab./ Chapada [dos Guimarães], Matto Grosso [Brazil]. H. H. Smith./ Godman-Salvin Coll. 1912.-23./ NHMUK.

Elbella intersecta ilona Evans, 1951. Holotype male with the following labels: / Type – *ilona* Evans/ R. Araguayra [*sic*] Prov. Goyaz, [Brazil] June 1906 (G. A. Baer)/ Rothschild Bequest B.M. 1938-1/. NHMUK. **Syn. n.**

Elbella intersecta paraensis* O. Mielke, 1995, *comb. rev.

Pls. 66–67, Figs. 16 (holotype ♂) (d, v)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará.

TYPE:

Holotype male with the following labels: / Holotypus/ Pará [Brazil]/ 11.996/ gen[itália]. prep[arada]., [O.] Mielke 1986/ *Elbella intersecta paraensis* [O.] Mielke, Holótipo, O. Mielke det. 1993/. OM-DZUP.

Elbella intersecta peter* Evans, 1951, *comb. rev.

Pls. 66–67, Figs. 19 (♂), 20 (paratype ♀) (d, v)

DISTRIBUTION – Northern Brazil (Pará).

Brazil: Pará (*Belém*, *Benevides*).

TYPE: Holotype male with the following labels: / Type – *peter* Evans/ Miles Moss Coll. B.M. 1947-453/ *P. semidentata* ♂ Pará [Brazil] garden bord *Banisteria mossil*. NHMUK.

Elbella intersecta rufitegula* O. Mielke, 1995, *comb. rev.

Pls. 66–67, Figs. 21 (holotype ♂), 22 (allotype ♀) (d, v)

DISTRIBUTION – Northeastern to southeastern Brazil.

Brazil: Pernambuco (*Recife*, *São Lourenço da Mata*); Bahia (*Uruçuca*); Espírito Santo (*Conceição da Barra*, *Linhares*, *Santa Teresa*); Minas Gerais (*Marliéria*); Rio de Janeiro (*Casimiro de Abreu*).

TYPE:

Elbella intersecta rufitegula O. Mielke, 1995. Holotype male with the following labels: / Holotypus/ 8-9-III-1994 Pq. [Parque] Est[adual]. Rio Doce, Marliéria, M[inas] G[erais] [Brazil] 350 m. [O.] Mielke leg./ *Elbella intersecta rufitegula* [O.] Mielke, Holótipo, O. Mielke det. 1993/ DZ 1876/. DZUP.

Elbella intersecta strova* Evans, 1951, *comb. rev.

Pls. 66–67, Figs. 23 (holotype ♂), 24 (♀) (d, v)

DISTRIBUTION – Panama and northern and southeastern Venezuela.

Panama: Chiriquí; Bocas del Toro. **Venezuela:** Bolívar (*Río Urbana*, *Suapure*); Guárico (*Antigua Hacienda Guaruma*); Yaracuy (*Nirgua*).

TYPE:

Elbella intersecta strova Evans, 1951. Holotype male with the following labels: / Type – *strova* Evans/ Suapure [Bolívar] Venez[uela]. 2-III-[18]99 (Klages)/ Rothschild Bequest B.M. 1939-1/. NHMUK.

Elbella lamprus* (Hopffer, 1874), *comb. rev.

Two subspecies.

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Elbella lamprus lamprus* (Hopffer, 1874), *comb. rev.

Pls. 68–69, Figs. 1 (lectotype ♂ of *Pyrrhopyge lamprus*), 2 (♀) (d, v)

Syn.: *Pyrrhopyga* [*sic*] *menecrates* Mabilie, 1878

Pyrrhopyge margimmiscus Hayward, 1935

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Minas Gerais (*Belo Horizonte*); Rio de Janeiro (*Cachoeiras de Macacu*, *Rio de Janeiro*, *Xerém*); São Paulo (*Teodoro Sampaio* – *Porto Cabral*); Paraná (*Diamante do Norte*, *Foz do Iguaçu*); Santa Catarina (*Itaiópolis*, *Seara* – *Nova Teutônia*); Rio Grande do Sul (*Guarani*, *Panambi*, *Santa Rosa*, *São Luiz Gonzaga*). **Paraguay:** Alto Paraná (*Itakyri*); Guairá (*Villarica*). **Argentina:** Misiones (*Puerto Iguazú*, *Salto Encantado*).

The citations **Peru** (Mabilie 1878, Palo Jr. 2017), **Peru:** Junín (*Río Perené*) (Martin 1941), **Bolivia** (Mabilie & Boulet 1908, Mabilie 1912, Draudt 1921), **Honduras**, **Colombia**, **Venezuela**, **Peru** and **Bolivia** (Hayward 1940b, Bell 1946, Hayward 1947, 1948), **Suriname**, **Venezuela** (Evans 1951), **Brazil:** Alagoas (*Maceió*) (Cardoso 1949) and Distrito Federal (*Brasília*) (de Lesse & Brown 1971) are probably incorrect. The citation **Venezuela** (Godman & Salvin 1893), as *Pyrrhopyge menecrates*, refers to *Elbella intersecta strova* *comb. rev.* (Orellana [2010]).

TYPES:

Pyrrhopyge lamprus Hopffer, 1874. Lectotype male (O. Mielke 1995) with the following labels: / 4886/ *lamprus* Hpfr.*/ Rio [de Janeiro, Brazil] v. Lgsdf./ Lectotipo/ *Pyrrhopyge lamprus* Hopffer, 1874,

Lectótipo, O. Mielke det. 1993/ *Elbella lampira lampira*, O. Mielke det. 1993/. MfN.

Pyrrhopyga [sic] menecrates Mabilie, 1878. Lectotype female (O. Mielke 1995) with the following labels: / Type/ *menecrates* P. Mab./ *Pyrrhopyga [sic] Menecrates* P. Mabilie Perou/ Hesp. Fifiers. ♀ God.15 Cr. Pl. 199E Surinam/ *Pyrrhopyga menecrates* Mabilie, 1878, Lectótipo, O. Mielke det. 1993/ *Elbella lampira lampira*, O. Mielke det. 1993/. IRSC.

Pyrrhopyge margimmiscus Hayward, 1935. Holotype female with the following labels: / Typus/ Coll. Breyer, Rep. Argentina, Misiones/ No. 7, K. J. Hayward det./ 2511/ *Pyrrhopyge margimmiscus* Holotipo ♀/ IML.

***Elbella lampirus albociliata* O. Mielke, 1995, comb. rev.**

Pls. 68–69, Figs. 3 (holotype ♂), 4 (allotype ♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: São Paulo (*Cubatão, Santos*); Paraná (*Morretes 20 m, Paranaguá, Pontal do Paraná*); Santa Catarina (*Águas Mornas – Teresópolis, Camboriú, Florianópolis, Joinville, Massaranduba, São Bento do Sul – Rio Natal, São Francisco do Sul*).

TYPE:

Elbella lampira [sic] albociliata O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ Serrinha, Joinville, Santa Catarina, Brasil, 0–200 m, 28-XII-1977, Miers leg./ *Elbella lampira albociliata* [O.] Mielke, Holótipo, O. Mielke det. 1993/ DZ 1844/. DZUP.

***Elbella viriditas* (Skinner, 1920), comb. rev.**

Three subspecies.

DISTRIBUTION – Northern Peru to central-eastern Bolivia, northern Brazil and northeastern Argentina.

***Elbella viriditas viriditas* (Skinner, 1920), comb. rev.**

Pls. 68–69, Figs. 5 (♂), 6 (♀) (d, v)

Syn.: *Pyrrhopyge carriae* Bell, 1931

DISTRIBUTION – Central-eastern Bolivia to northeastern Argentina.

Bolivia: La Paz (*Mapiri, Muchanes*). **Paraguay:** Paraguairí (*Sapucaí*); San Pedro (*Nueva Germania, Río Jejuy*). **Argentina:** Misiones (*San Pedro*).

The citation **Honduras:** Cortés (*San Pedro Sula*) (Skinner 1920, Bell 1931, Gillham & Ehrlich 1955), **El Salvador** (Steinhauser 1975) and **Brazil** (Palo Jr. 2017) are incorrect.

TYPES:

Pyrrhopyge viriditas Skinner, 1920. Holotype female with the following labels: / San Pedro Sula [Honduras]/ Type No. 7058 *Pyrrhopyge viriditas* Skinner/ ANSP, transferred to CMP.

Pyrrhopyge carriae Bell, 1931. Holotype male with the following labels: / Sapucaí [Paraguairí] Paraguay II-11-25/ Rodolpho Heinrich/ Holotype *Pyrrhopyge carriae* Bell 7587/ Det. E. L. Bell/ Genitalia slide No. 1958/. ANSP, transferred to CMP.

***Elbella viriditas chia* Evans, 1951, comb. rev.**

Pls. 68–69, Figs. 7 (♂), 8 (♀) (d, v)

DISTRIBUTION – Northern Peru to northern Brazil (Amazonas and Pará).

Peru: Loreto (*Pebas*). **Brazil:** Amazonas (*Fonte Boa, Rio Madeira, São Paulo de Olivença, Tefê*); Pará (*Óbidos*).

The citation **Colombia** (Palo Jr. 2017) is probably incorrect.

TYPE:

Elbella intersecta chia Evans, 1951. Holotype male with the

following labels: / Brazil [Amazonas] Madeira R[iver]. Amazon R. U. Moffat. 1913-485/ Type – *chia* Evans/. NHMUK.

***Elbella viriditas boliviana* O. Mielke, 1995, comb. rev.**

Pls. 68–69, Figs. 9 (holotype ♂) (d, v)

DISTRIBUTION – Central-eastern Bolivia.

Bolivia: La Paz (*Caranavi*); Santa Cruz (*Parque Nacional Amboró*). TYPE:

Elbella viriditas boliviana O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ Caranavi, Yungas, [La Paz] Bolivia, 500 – 1000 m, II-III-1989, Tello leg./ *Elbella viriditas boliviana* [O.] Mielke, Holótipo, O. Mielke det. 1993/ OM 22.812/. OM-DZUP.

***Elbella dulcinea* (Plötz, 1879), comb. rev.**

Pls. 68–69, Figs. 23 (♂), 24 (♀) (d, v)

DISTRIBUTION – Panama to central Colombia, eastern Ecuador, Trinidad and Tobago and Venezuela.

Panama: Chiriquí (*Bugaba, Lino, Santa Cruz*); Panamá (*Ancon, Bayano, El Llano, Madden Forest*); Panamá Oeste (*Cerro Campana*); Veraguas. **Colombia:** Cundinamarca (*Bogotá*). **Ecuador:** Tungurahua (*Baños*). **Trinidad and Tobago:** Port of Spain (*Cascade – Hololo Mountain Road*). **Venezuela:** Aragua (*El Limón, Parque Nacional Henry Pittier, Río El Castaño*); Bolívar (*Montecal*); Distrito Capital (*Caracas – Puente Hierro*); Guárico (*Antigua Hacienda Guaruma*); Miranda (*Río Chico*); Táchira (*Río Frío*); Trujillo (*Valera*).

TYPE:

Pyrrhopyga [sic] dulcinea Plötz, 1879. Lectotype male (O. Mielke 1995) with the following labels: / Origin./ *Dulcinea* Mab. ♂/ *Scylla Mén. Dulcinea* Stg./ Col[ombia]. K./ *Scylla Mén.*/ Coll. Staudinger/ Lectotipo/ *Pyrrhopyga dulcinea* Plötz, 1879, Lectótipo, O. Mielke det. 1993/ *Elbella dulcinea*, O. Mielke det. 1993/. MfN.

***Elbella scylla* (Ménétriés, 1855), comb. rev.**

Pls. 70–71, Figs. 1 (♂), 2 (♀) (d, v)

DISTRIBUTION – Western and eastern Mexico to Costa Rica.

Mexico: Campeche (*Calakmul*); Catemaco (*Dos Amates*); Chiapas (*Agua Azul, Boca de Chajul, Chorcadero, Comitán de Domínguez, San Jerónimo, Ocosingo – San Quentin, Santa Rosa, Río Lacantún*); Colima (*Colima, Comala, El Jabalí, La Salada, Paso Ancho, Punta de Agua de Camotlán, Villa de Álvarez – Agua Dulce*); Durango (*Revolcaderos*); Guerrero (*Naranjo, Tierra Colorada*); Hidalgo (*Acuimantla*); Jalisco (*Ahuacapán, Boca de Tomatlán, La Galera, Mismaloya, Puerto Vallarta*); Mexico (*Malinalco, Sierra Manchititla [Temascaltepec]*); Michoacán (*Apatzingan*); Morelos (*Ayala, Progreso, Puente de Ixtla, Rancho Viejo, Sierra de Huautla, Xochitepec, Yautepec*); Nayarit (*El Guamúchil, El Refilión, Jumatán, Mecatán, Singayta, Tepic*); Oaxaca (*Candelaria Loxicha, Chimaultepec, Putla, Sierra de Juárez, Tuxtepec, Yautepec*); Puebla (*Matamoros, Zihuatéutla*); Quintana Roo (*Noh-bek, Nuevo Xcán*); San Luis Potosí (*Aquismón, Huasteca Potosina, Pago Pago*); Sinaloa (*Mazatlán, Rosario, Venado*); Tabasco (*Teapa*); Tamaulipas (*Alta Cima, Goleana Canyon, Gomez Faria, La Florida, La Pesca*); Veracruz (*Catemaco, Los Tuxtlas, Misantra, Presidio, Tierra Blanca, Xalapa*). **Belize:** Cayo (*Sibun Forest Reserve*). **Guatemala:** Alta Verapaz (*Cahabón, Polochic Valley, San Cristobal*); Baja Verapaz (*San Jerónimo*); El Petén (*Parque Nacional Tikal*); Izabal (*Morales – Cayuga, Morales – Motagua Valley*); Jutiapa (*El Paraíso*); Santa Rosa (*Guzacapan*). **Honduras:** Atlántida (*La Ceiba*); Chontales; Cortés (*San Pedro Sula*). **El Salvador:** La Libertad; San Salvador. **Nicaragua:** Chontales; Granada (*Granada*); Managua (*Managua*). **Costa Rica:** Alajuela (*Atenas, San Mateo*); Guanacaste (*Área de Conservación Guanacaste*); San José (*Colón, Cantón de Mora – El Rodeo, Mora*).

The citations **Panama, Colombia, Peru, Bolivia** and **Brazil** (Palo Jr. 2017) are incorrect.

TYPE:

Pyrrhopyga [**sic**] *scylla* Ménétériés, 1855. The original description mentions an unspecified number of specimens from Nicaragua in ZISP. To ensure the correct identification of *Pyrrhopyga scylla*, one syntype male from Nicaragua is **herein designated as the lectotype**; the labels of this specimen were not annotated during our visit to this institution in 1996, but two other labels will be added: / Lectotypus/ Lectotypus *Pyrrhopyga scylla* Ménétériés, 1855, O. Mielke, Brockmann & C. Mielke det. 2019/. ZISP.

Elbella theseus* (Bell, 1934), *comb. rev.

Pls. 72–73, Figs. 13–14 (♂), 15 (♀) (d, v)

DISTRIBUTION – Western and central Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, northwestern Venezuela and northern Brazil (Amazonas and Pará).

Colombia: Boyacá (*Muzo*); Cundinamarca (*Bogotá, Medina*); Meta (*Upper Río Negro*). **Ecuador:** Loja (*Loja*); Morona-Santiago (*Macas, Santiago de Méndez, Sináí – Tres Marías*); Napo (*Jatun Sacha*); Pastaza (*Abitagua, Mera, Pastaza*); Santo Domingo de los Tsáchilas (*Santo Domingo de los Colorados*); Tungurahua (*Yunguilla*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Rodríguez de Mendoza*); Cajamarca (*Charapí*); Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Quitacalzón 1,100 m*); Huánuco (*Tingo Maria*); Junín (*Chanchamayo, La Merced, Shimá*); Loreto (*Iquitos*); Madre de Dios (*Parque Nacional del Manu, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Pozuzo*); Puno (*Chaquimayo, Yahuarimayo*); San Martín (*Juanjuí, Rioja – El Afluente 1,300 m, Tarapoto, Tocache*). **Bolivia:** Cochabamba (*Chapare, Cochabamba*); La Paz (*Apolobamba, Mapiri, Río Zongo, Taipiplaya*); Santa Cruz. **Venezuela:** Táchira (*Río Doradas*). **Brazil:** Amazonas (*Manaus, São Paulo de Olivença*); Pará (*Itaituba, Monte Cristo*).

TYPE:

Jemadia theseus Bell, 1934. Holotype male with the following labels: / Type/ G531/ Brazil/ *Jemadia theseus* Bell Holotype ♂/. AMNH.

***Apatiella* Grishin, 2019**

One species.

DISTRIBUTION – Southeastern to southern Brazil and northeastern Argentina.

***Apatiella iphinous* (Latreille, [1824])**

Three subspecies.

DISTRIBUTION – Southeastern to southern Brazil and northeastern Argentina.

TAXONOMY – There are no differences in the morphology and the DNA barcode between *A. iphinous iphinous* (n= 3) and *A. iphinous mariae* **comb. n., stat. n.** (n= 4), despite the wing markings, which provide grounds to include the last as a subspecies of the first. The three subspecies are allopatric, but in a very small valley in Itamonte, Minas Gerais, Brazil, there occurs a hybrid population between *A. iphinous iphinous* and *A. iphinous mariae* **comb. n., stat. n.** In these specimens the red colour at the head is reduced until nearly absent, sometimes paler and almost yellow; anal tuft also varies from black to pale yellow; hindwing dorsally sometimes with some blue scales near the anal angle; and ventral hindwing sometimes also with diffuse blue scales (see Pls. 68–69, Figs. 17–22; Pls. 74–75, Figs. 22).

***Apatiella iphinous iphinous* (Latreille, [1824])**

Pls. 68–69, Figs. 13 (♂), 14 (♀) (d, v)

Syn.: *Pyrrhopyga* [**sic**] *othello* Plötz, 1879

Mimoniades mimetes Mabilie, 1909

Mimoniades othello var. *nigrita* J. Zikán, 1938

DISTRIBUTION – Southeastern Brazil.

Brazil: Minas Gerais (*Camanducaia – Monte Verde, Itamonte* (all hybrid specimens), *Passa Quatro, Poços de Caldas, Virgínia*); Rio de Janeiro (*Cachoeiras de Macacu, Casimiro de Abreu, Itatiaia, Petrópolis, Resende, Teresópolis*); São Paulo (*Campos do Jordão, Mogi das Cruzes 1,200 m, São Bento do Sapucaí, São José do Barreiro – Bocaina*).

The citations **Guyana** (Mabilie 1912, Draudt 1921) are incorrect.

TYPES:

Hesperia iphinous Latreille, [1824]. Lectotype male (O. Mielke 1995) with the following labels: / Type H T/ Lectotipo/ *iphinoë* *H. iphinous* Lat. Brésil/ *Mimoniad. mimetes* Mab. Brésil/ Ex musaeo P. Mabilie 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/ *Hesperia iphinous* Latreille, [1824], Lectótipo, O. Mielke det. 1993/ *Elbella iphinous* (Latreille, [1824]), O. Mielke det. 1993./ NHMUK.

Pyrrhopyga [**sic**] *othello* Plötz, 1879 and *Mimoniades mimetes* Mabilie, 1909. Neotype males (O. Mielke 1995; same specimen) for both taxa with the following labels: / Neotipo *mimetes, othello* P. N. Itatiaia, Resende, R[io de] J[aneiro] [Brazil]. 10-I-1973 1200 m, [O.] Mielke [leg.]/ DZ 4179/ *Mimoniades othello* Plötz, 1879, Neótipo, O. Mielke det. 1993/ *Mimoniades mimetes* Mabilie, 1909, Neótipo, O. Mielke det. 1993/ *Elbella iphinous* (Latreille, [1824]), O. Mielke det. 1993/. DZUP.

Mimoniades othello var. *nigrita* J. Zikán, 1938. Lectotype male (O. Mielke 1995) with the following labels: / Lectotipo/ ♂/ Typus/ 11.-2.-[19]26 Mar.[omba, Parque Nacional do Itatiaia, Itatiaia, Rio de Janeiro, Brazil, Zikán leg., 1.200 m]/ *Mimoniades othello* Pltz. var. *nigrita* Zik[án]./ Coleção J. F. Zikán/ *Elbella iphinous* (Latr., 1824) [O.] Mielke det./ I.O.C., Lepidop. 33.001/. IOC.

***Apatiella iphinous molinai* (Hayward, [1942]),**

comb. n.

Pls. 68–69, Figs. 10–11 (♂), 12 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil and northeastern Argentina.

Brazil: Minas Gerais (*Catas Altas – Caraça*); São Paulo (*Guapiara – Paivinha*); Paraná (*Campina Grande do Sul – Morro do Caratuva, Guaratuba – Castelhano, Guaratuba – Pontal do Itararé 900–1,200 m, Morretes 20 m, Morretes – first plateau of Paraná 800 m, Quatro Barras – Morro do Anhangava, Pontal do Paraná*); Santa Catarina (*Joinville*). **Argentina:** Misiones.

The type locality mentioned in the original description, Mato Grosso, Brazil, is wrong; the label of the holotype says Misiones [Argentina].

TYPE:

Pyrrhopyge molinai Hayward, [1942]. Holotype male with the following labels: / Type/ *Pyrrhopyge molinai* Hayw., holotype ♂, Misiones [Argentina]. K. J. Hayward det./ Prep. 590/. IML.

Apatiella iphinous mariae* (Bell, 1931), *comb. n.,

stat. n.

Pls. 68–69, Figs. 15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil.

Brazil: São Paulo (*São Paulo*); Paraná (*Balsa Nova, Castro, Curitiba, Guarapuava, Inácio Martins, Lapa, Mafra, Palmas, Piên, Ponta Grossa, Prudentópolis, São José dos Pinhais, Tibagi, Tunas do Paraná, Turvo, União da Vitória*); Santa Catarina (*Águas Mornas – Teresópolis, Alto Rio dos Cedros, Blumenau, Corupá, Curitiba, Joaçaba, Lages, Mafra, Massaranduba, Santa Cecília, São Bento do Sul – Rio Vermelho, Urubici – Santa Clara*); Rio Grande do Sul (*Bom Jesus, Gramado, Passo Fundo, Pelotas, São Francisco de Paula*).

TYPE:

Pyrrhopyge mariae Bell, 1931. Holotype male with the following labels: / Type/ 391/ Rio Grande do Sul [Brazil]/ *Pyrrhopyge mariae* Bell Holotype ♂/. AMNH.

Hegesippe Evans, 1951

Two species.

DISTRIBUTION – Central to southern Brazil and Uruguay.

Hegesippe luteizona (Mabille, 1877)

Pls. 70–71, Figs. 5 (neotype ♂ of *Pyrrhopyge xanthomargo*), 6, 10, 12 (♀), 7, 9, 11 (♂) (d, v)

Syn.: *Pyrrhopyga* [*sic*] *martii* Plötz, 1879

Pyrrhopyga [*sic*] *josepha* Plötz, 1879

Sarbia amoena Röber, 1925

Pyrrhopyge xanthomargo Hayward, 1942

DISTRIBUTION – Central to southern Brazil.

Brazil: Mato Grosso; Goiás (*Goiânia*, *Teresina de Goiás*); Distrito Federal (*Brasília*, *Planaltina*); Bahia (*Rio de Contas* – *Pico das Almas* 1,600 m); Minas Gerais (*Baependi*, *Camanducaia* – *Monte Verde*, *Catas Altas* – *Caraça*, *Nova Lima*, *Passa Quatro*, *Poços de Caldas*, *Rio das Velhas*, *Santana do Riacho* – *Serra do Cipó*, *São João del Rei*); São Paulo (*Angatuba*, *Araraquara*, *Campos do Jordão*, *Itararé*, *Rio Claro*, *São Bernardo do Campo*, *São José do Barreiro* – *Bocaina*, *São Paulo*); Paraná (*Castro*, *Tibagi* – *Guartelá and Estiva*); Rio Grande do Sul (*Central Region*, *Pelotas*).

The citations **Mexico** (Mabille 1877, Mabille 1878, Godman & Salvin 1893, Mabille 1903, Mabille & Boulet 1908, Mabille 1912, Draudt 1921, Bell 1934b, Spitz 1932, Hoffmann 1941, 1976) are incorrect.

TYPES:

Pyrrhopyga [*sic*] *luteizona* Mabille, 1877. Lectotype male (O. Mielke 1995) with the following labels: / Type/ *Sarbia luteizona* Mab. det. Fr. J. Ball 1926/ *Pyrrhopyga luteizona* P. Mabille Mexique/ Mexique Gluss leg./ Lectotipo/ *Pyrrhopyga luteizona* Mabille, 1877, Lectótipo, O. Mielke det. 1993/ *Elbella luteizona*, O. Mielke det. 1993/. IRSC.

Pyrrhopyga [*sic*] *martii* Plötz, 1879. Holotype male with the following labels: / Type/ 4915/ *Martii* Plötz*/ Brasil. Virmond/ *Martii* Plötz Stett. Ent. Zeit. 1879, p. 525/. MfN.

Pyrrhopyga [*sic*] *josepha* Plötz, 1879. Holotype male with the following labels: / Holotypus/ 4899/ 1: 40/ Brasil Besckel/ *Josepha* Plötz */ *Josepha* Plötz Stett. Ent. Zeit. 1879, p. 534/. MfN.

Sarbia amoena Röber, 1925. The holotype of *Pyrrhopyga* [*sic*] *martii* Plötz, 1879 is also the neotype (O. Mielke 1995) of *Sarbia amoena*; the following two labels were added: / Neotipo/ *Sarbia amoena* Röber, 1925, Neótipo, O. Mielke det. 1993/. MfN.

Pyrrhopyge xanthomargo Hayward, 1942. Holotype female with the following labels: / Tipus/ Angatuba, Est. de São Paulo [Brazil] III-1917, Coll. Azevedo Marques/ *Pyrrhopyge xanthomargo* Hayw. ♀ Holotipo/ No. 18.627 Proc./ *Hegesippe luteizona* (Mab., 1877) [O.] Mielke det./ MNRJ. This specimen was lost by fire (2-IX-2018). To ensure the correct identification of *Pyrrhopyge xanthomargo*, erroneously mentioned as a synonym of *Elbella carriae* (Bell, 1931) by Evans (1951) and currently a synonym of *Elbella viriditas viriditas* (Skinner, 1920), a male is **herein designated as the neotype** for *P. xanthomargo*; the specimen with the following labels: / Neotypus/ Brazil Brasília D. F. Fazenda Agua Limpa D. Gifford [leg.] 15 III 1977/ ex coleção Gifford/ *Elbella luteizona* O. Mielke det. 1993/ Neotypus *Pyrrhopyge xanthomargo* Hayward, 1942 O. Mielke, Brockmann & C. Mielke det. 2019/ DZ 4175/. DZUP. This specimen, although a male, is very similar to the destroyed holotype female of *P. xanthomargo*. This dark phenotype is known from Brasília (Distrito Federal) to Castro (Paraná) in Brazil.

Hegesippe hegesippe (Mabille & Boulet, 1908)

Pls. 70–71, Figs. 13 (♂), 14 (♀) (d, v)

DISTRIBUTION – Southern Brazil and Uruguay.

Brazil: Paraná (*Campina Grande do Sul*, *Castro*, *Curitiba*, *Lapa*, *Palmas*, *São José dos Pinhais*, *Tibagi*, *Tijucas do Sul*); Santa Catarina

(*Curitibanos*, *Lages*, *Massaranduba*, *Otacílio Costa*, *São Bento do Sul* – *Rio Vermelho*, *Urubici*); Rio Grande do Sul (*Alto Feliz*, *Bom Jesus*, *Pelotas*, *Vacaria*). **Uruguay:** Montevideo (*Montevideo*).

The citations **Bolivia** (Mabille & Boulet 1908, Evans 1951, O. Mielke 1995, Gareca *et al.* 2006) and **Guyana** (Mabille 1912, Draudt 1921) are incorrect.

TYPE:

Sarbia hegesippe Mabille & Boulet, 1908. Holotype male (one male was mentioned) with the following labels: / Type H T/ *hegesippe* ♂ ? *pertyi* Pl./ *pertyi* Pl. am.[America] m.[meridional]/ Ex musaeo P. Mabille 1923/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Microceris Watson, 1893

One species.

DISTRIBUTION – Central Brazil.

Microceris variicolor (Ménétriés, 1855)

Pls. 70–71, Figs. 15 (♂), 16 (♀) (d, v)

DISTRIBUTION – Central Brazil.

Brazil: Mato Grosso (*Chapada dos Guimarães* – *Buriti*, *Diamantino*, *Pantanal*, *Rondonópolis*, *Tombador*); Mato Grosso do Sul (*Nioaque*, *Sapezal* – *Rio Juruena*); Maranhão (*Balsas*, *Imperatriz*); Tocantins (*Taguatinga*); Goiás (*Anápolis*, *Cavalcante*, *Chapada dos Veadeiros*, *Goiânia*, *Jatá*, *Parque Nacional das Emas*); Distrito Federal (*Brasília*, *Planaltina*); Minas Gerais (*Araguari*, *Uberlândia*); São Paulo (*Brotas* – *Campo Alegre*, *Castilho*).

There is a male from French Guiana in MNHN, probably mislabelled.

REMARKS – A typical species of the Brazilian savannah (= Cerrado) in open areas.

TYPE:

Pyrrhopyga [*sic*] *variicolor* Ménétriés, 1855. Lectotype male (O. Mielke 1995) from Minas Gerais, Brazil, to which were added the following two labels: / Lectotipo/ *Pyrrhopyga variicolor* Ménétriés, 1855, Lectótipo, O. Mielke det. 1994/. ZISP.

Blubella Grishin, 2019

Eleven species.

DISTRIBUTION – Mexico to southern Brazil, Paraguay and northeastern Argentina.

Blubella adonis (Bell, 1931)

Pls. 70–71, Figs. 3 (♂), 4, 8 (♀) (d, v)

DISTRIBUTION – Southeastern to southern Brazil, Paraguay and northeastern Argentina.

Brazil: Minas Gerais (*Passa Quatro*, *Poços de Caldas*); Rio de Janeiro (*Cachoeiras de Macacu*, *Itatiaia*); São Paulo (*Campos do Jordão*, *São Paulo*); Paraná (*Balsa Nova*, *Castro*, *Curitiba*, *Foz do Iguaçu*, *Guarapuava*, *Guaratuba*, *Inácio Martins*, *Londrina*, *Matinhos*, *Morretes*, *Palmas*, *Pontal do Paraná*, *Rolândia*, *São José dos Pinhais*, *Turvo*); Santa Catarina (*Águas Mornas* – *Teresópolis*, *Blumenau*, *Dalbérgia* (as *Neu Bremen*), *Joaçaba*, *Joinville*, *Massaranduba*, *Santa Cecília*, *São Bento do Sul*, *Seara* – *Nova Teutônia*, *Taió*); Rio Grande do Sul (*Cerro Largo*, *Pelotas*, *Porto Alegre*, *São Francisco de Paula*, *São Luiz Gonzaga*). **Paraguay:** Alto Paraná (*Itakyry*); Guairá (*Villarica*). **Argentina:** Misiones (*General Belgrano*, *Puerto Iguazú*, *Puerto Libertad*).

TYPE:

Pyrrhopyge adonis Bell, 1931. Holotype male with the following labels: / Type/ Villarica Nov[ember]. [Guairá] Paraguay/ *Pyrrhopyge adonis* Bell Holotype ♂/. AMNH.

***Blubella rondonia* (O. Mielke, 1995)**

Pls. 70–71, Figs. 17 (holotype ♂), 18 (allotype ♀) (d, v)

DISTRIBUTION – Central-eastern Peru and central-northern Brazil (Rondônia and Mato Grosso).

Peru: Cusco (*Cosñipata Valley – Pilcopata 550 m*); Huánuco (*Tingo Maria*); Junín (*La Merced, Río Perené, San Ramón, Satipo, Shimá*). **Brazil:** Rondônia (*Cacaulândia, Pimenta Bueno*); Mato Grosso (*Diamantino*).

TYPE:

Elbella rondonia O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ 28-I-1978 Alto Rio Arinos, Faz[enda]. São João, Diamantino, MT [Mato Grosso,] [Brazil], Furtado leg./ *Elbella rondonia* [O.] Mielke, Holótipo, O. Mielke det. 1993/ gen[itália]. prep[arada]. [O.] Mielke 1987/ OM 32.893/. OM-DZUP.

***Blubella lustra* (Evans, 1951)**

Pls. 70–71, Figs. 19 (♂), 20 (♀) (d, v)

DISTRIBUTION – Central Colombia to western Ecuador, eastern Colombia and northwestern Venezuela.

Colombia: Boyacá (*Otanche*); Cundinamarca (*Bogotá*); Tolima (*Mariquita*). **Ecuador:** Imbabura (*Paramba*). **Venezuela:** Táchira (*Río Frío*).

TYPE:

Elbella azeta lustra Evans, 1951. Holotype male with the following labels: / Type – *lustra* Evans/ Bogota [Cundinamarca, Colombia]/ Rothschild Bequest B.M. 1939-1./ NHMUK.

***Blubella miodesmiata* (Röber, 1925)**

Pls. 70–71, Figs. 21 (♂) (d, v)

DISTRIBUTION – Southern Mexico, Costa Rica to western Ecuador and central Colombia.

Mexico: Oaxaca (*Comaltepec*). **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste*). **Panama:** Colón (*Gamboa, Piña*). **Colombia:** Boyacá (*Muzo*); Cauca (*Popayán*); Cundinamarca (*Bogotá*); Department?: (*Río Magdalena*). **Ecuador:** Esmeraldas (*San Lorenzo*).

TYPE:

Jemadia miodesmiata Röber, 1925. Neotype male (O. Mielke 1995) with the following labels: / Neotipo/ Colombia/ Rothschild Bequest B.M. 1939-1./ *Jemadia miodesmiata* Röber, 1925. Neótipo. O. Mielke det. 1993/ *Elbella miodesmiata* (Röber, 1925. O. Mielke det. 1993/. NHMUK.

***Blubella etna* (Evans, 1951)**

Pls. 70–71, Figs. 22 (♂), 23 (allotype ♀ of *Elbella madeira*) (d, v)

Syn.: *Elbella etna moda* Evans, 1951

DISTRIBUTION – Central and eastern Colombia to central-eastern Bolivia, Trinidad and Tobago, French Guiana, northern Brazil and northwestern Argentina.

Colombia: Amazonas (*Letícia*); Boyacá (*Otanche*); Cundinamarca (*Bogotá*); Guaviare (*San José del Guaviare*). **Ecuador:** Pastaza (*Río Pastaza*). **Peru:** Huánuco (*Cucharas – Río Huallaga, Tingo Maria*); Junín (*La Merced, Satipo, Shimá*); Loreto (*Iquitos, Pebas*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado, Shintuya*); San Martín (*Juanjuí, Sapasoa*). **Bolivia:** Cochabamba (*Chapare*); La Paz (*Caranavi*); Santa Cruz (*Sara, Trinidad*). **Trinidad and Tobago:** Diego Martin (*Cumberland Hill*). **French Guiana.** **Brazil:** Amazonas (*Benjamin Constant, Parintins, Tefé*); Pará (*Belém, Óbidos*); Acre (*Cruzeiro do Sul, Santa Rosa do Purus*); Rondônia (*Cacaulândia, Pimenta Bueno*); Mato Grosso (*Diamantino*). **Argentina:** Santiago del Estero.

TAXONOMY – The figured female specimen (Pls. 70–71, Fig. 23)

is the allotype of *Elbella madeira* O. Mielke, 1995, a misidentification.

TYPES:

Elbella etna etna Evans, 1951. Holotype male with the following labels: / Type – *etna* Evans/ Tefé (Ega) Amazonas [Brazil] M. de Mathan/ 7^{brc} 8^{brc} 1879/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

Elbella etna moda Evans, 1951. Holotype male with the following labels: / Type – *moda* Evans/ Peru. [Junín] La Merced. 2500ft. V & VI. [19]03. Watkins & Tomlinson. 1904-133/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Blubella madeira* (O. Mielke, 1995)**

Pls. 70–71, Figs. 24 (holotype ♂) (d, v)

DISTRIBUTION – Central-eastern Peru and northern to central-northern Brazil (Amazonas and Mato Grosso).

Peru: Cusco (*Cosñipata Valley – San Pedro 1,400 m*); Junín (*Río Paucartambo*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado*). **Brazil:** Amazonas (*São Paulo de Olivença*); Mato Grosso (*Diamantino*).

TYPE:

Elbella madeira O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ 23-IV-1978 Alto Rio Arinos, Faz[enda]. S[ão]. João, Diamantino, MT [Mato Grosso] [Brazil], Furtado leg./ gen[itália]. prep[arada]. [O.] Mielke 1987/ OM 33.030/ *Elbella madeira* [O.] Mielke, Holótipo, O. Mielke det. 1993/. OM-DZUP.

***Blubella bicuspis* (de Jong, 1983)**

Pls. 72–73, Figs. 1 (holotype ♂) (d, v)

DISTRIBUTION – Suriname and northern Brazil (Amazonas).

Suriname: Paramaribo (*Paramaribo*). **Brazil:** Amazonas (*Manaus – 60 km N, São Paulo de Olivença*).

TYPE:

Elbella bicuspis de Jong, 1983. Holotype male with the following labels: / Museum Leiden Holotype *Elbella bicuspis* R. de Jong/ Suriname Paramaribo E. H. Jonkers/ Museum Leiden Coll. E. H. Jonkers Acc. 1980-03/. NBC.

***Blubella patroclus* (Plötz, 1879)**

Two subspecies.

DISTRIBUTION – Honduras, Panama to northern Argentina and to northwestern Venezuela.

***Blubella patroclus patroclus* (Plötz, 1879)**

Pls. 72–73, Figs. 2 (♂), 3 (♀) (d, v)

Syn.: *Jemadia umbrata* Mabilie & Boulet, 1908

DISTRIBUTION – Eastern Ecuador to central-eastern Bolivia and northern Argentina.

Ecuador: Chimborazo (*Riobamba*); Loja (*Loja*); Napo (*Cotundo*); Pastaza (*Mera, Río Pastaza*); Tungurahua (*Ambato*); Zamora-Chinchipe (*Zamora*). **Peru:** Amazonas (*Cordillera del Condor, Huambo, Rodríguez de Mendoza*); Cajamarca (*Charapi*); Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Quitacazón 1,100 m, Salvación*); Huánuco (*Cucharas – Río Huallaga, Tingo Maria*); Junín (*Chanchamayo, Río Perené, Shimá*); Loreto (*Pebas*); Madre de Dios (*Tambopata Reserve*); Pasco (*Pozuzo*); Puno (*Carabaya*); San Martín (*Huambo, Rioja – El Afluente 1,300 m*). **Bolivia:** Cochabamba (*Chapare, Cochabamba*); La Paz (*Caranavi, Río Zongo, Taipiplaya*); Santa Cruz (*Santa Cruz de la Sierra, Sara*). **Argentina:** Santiago del Estero.

TYPES:

Pyrrhopyga [sic] patroclus Plötz, 1879. Holotype male with the following labels: / Holotypus/ Type/ ? Ecuador, ? Peru Schmettr/

18262/ 9: 26/ 9: 7 + 9: 26/ *patroclus* Plötz type Pl. gibt [??] 18362/ gen[itália]. prep[arada]., [O.] Mielke 1979/ Genital-Unters. Nr. 4.719, Zool. Mus. Berlin/. MfN.

Jemadia umbrata Mabilie & Boullet, 1908. Neotype male (O. Mielke 1995) with the following labels: / Neotipo/ Caranavi, [La Paz] Bolivia, 600-1000 m, IX-X-1988. Tello leg./ *Jemadia umbrata* Mabilie & Boullet, 1908, Neótipo, O. Mielke det. 1993/ *Elbella patroclus patroclus* (Plötz), O. Mielke det. 1993/ gen[itália]. prep[arada]., [O.] Mielke 1993/ 20.253/. OM-DZUP.

***Blubella patroclus acala* (Evans, 1951)**

Pls. 72–73, Figs. 4 (♂) (d, v); Pls. 74–75, Figs. 4 (♀) (d, v)

DISTRIBUTION – Honduras, Panama to western, central and eastern Colombia and northwestern Venezuela.

Honduras. Panama: Chiriquí (*Bugaba*). **Colombia:** Amazonian area; Antioquia (*Valdivia*); Boyacá (*Muzo*); Cesar (*Manauare*); Cundinamarca (*Bogotá, Guayabetal*); Meta (*Aama de Cabarral, Upper Río Negro*). **Venezuela:** Táchira (*Río Frío*).

TYPE:

Elbella umbrata acala Evans, 1951. Holotype male with the following labels: / Type/ B.C.A.Lep. Rhop. *Jemadia patrobas*, Hew./ Godman-Salvin Coll. 1912.-23./ Manauare, [Cesar] N. Colombia, F. Simons/ *Elbella umbrata acala* Evans, 1951/ Holotype, O. Mielke det. 1993/. NHMUK.

***Blubella azeta* (Hewitson, 1866)**

Two subspecies.

DISTRIBUTION – Northern and eastern Colombia to central-eastern Bolivia, southern Venezuela to French Guiana, northern and central Brazil, Paraguay and northeastern Argentina.

***Blubella azeta azeta* (Hewitson, 1866)**

Pls. 72–73, Figs. 5 (♂), 6 (♀) (d, v)

DISTRIBUTION – Northern and eastern Colombia to central-eastern Bolivia, southern Venezuela to French Guiana, northern Brazil and Paraguay.

Colombia: Cesar (*Manauare*); Guaviare (*San José del Guaviare*); Meta (*Villavicencio*). **Ecuador:** Loja (*Loja*); Morona-Santiago (*Macas, Santiago de Méndez*); Napo (*Tena*); Pastaza (*Puyo*); Tungurahua (*Yungilla*). **Peru:** Cusco (*Cosñipata Valley – Chontachaca 950 m to Quebrada Santa Isabel 1,200 m, Quincemil*); Huánuco (*Tingo Maria*); Junín (*Río Perené, Satipo, Shimá*); Loreto (*Pebas, Río Putumayo*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); Pasco (*Pozuzo, Puerto Bermudez – Río Pichis*); San Martín. **Bolivia:** Cochabamba (*Chapare*); La Paz (*Caranavi, Río Zongo, Sarampiuni*); Santa Cruz (*Buenavista, Parque Nacional Amboró*). **Venezuela:** Bolívar (*El Caura*). **Guyana:** Potaro-Siparuni (*Anundabaru*). **French Guiana:** Cayenne (*Matoury*). **Brazil:** Amazonas (*Barcelos, Manaus, Manicoré, Maués, São Paulo de Olivença*); Pará (*Rio Arapiuns, Juruti, Monte Cristo, Óbidos, Santarém*); Acre (*Rio Branco*); Rondônia (*Vilhena*); Rondônia (*Ariquemes*); Mato Grosso (*Barra do Bugres, Diamantino*). **Paraguay:** Paraguairí (*Sapucaí*).

TYPE:

Pyrrhopyga [sic] azeta Hewitson, 1866. Lectotype male (O. Mielke 1995) with the following labels: / Type / St. Paulo [de Olivença, Amazonas, Brazil] Hewitson Coll. 79.-89. *Pyrrhopyga azeta* 2./ Lectotipo/ *Pyrrhopyga azeta* Hewitson, 1866. Lectótipo, O. Mielke det 1993/ *Elbella azeta azeta*, O. Mielke det. 1993/. NHMUK.

***Blubella azeta giffordi* (O. Mielke, 1995)**

Pls. 72–73, Figs. 7 (holotype ♂), 8 (allotype ♀) (d, v)

DISTRIBUTION – Central Brazil to northeastern Argentina.

Brazil: Distrito Federal (*Brasília*); Minas Gerais (*Betim, Pedro Leopoldo, Rio Acima*); São Paulo (*Rancharia*); Paraná (*Diamante*

do Norte, Foz do Iguacu, Terra Rica). **Argentina:** Misiones (*General Belgrano, Puerto Iguazú*).

TYPE:

Elbella azeta giffordi O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ 29-XI-1980 ESAF, Brasília, D[istrito] F[ederal] [Brazil]. Ferreira leg./ DZ 1882/ gen[itália] prep[arada] [O.] Mielke 1993/ *Elbella azeta giffordi* [O.] Mielke, Holótipo, O. Mielke det. 1993/. DZUP.

***Blubella blanda* (Evans, 1951)**

Pls. 72–73, Figs. 11 (♂), 12 (♀) (d, v)

DISTRIBUTION – Western and central Colombia to western Ecuador, eastern Colombia to central-eastern Bolivia, Guyana to French Guiana, northern and southeastern Brazil, Paraguay and northeastern Argentina.

Colombia: Boyacá (*Otanche*); Caquetá (*Río Orteguasa*). **Ecuador:** Esmeraldas (*Chuchuví*); Imbabura (*Paramba*); Morona-Santiago (*Macas, Santiago de Méndez*). **Peru:** Cusco (*Cosñipata Valley – Quebrada Santa Isabel 1,200 m, Quincemil*); Huánuco (*Tingo Maria*); Junín (*Río Colorado, Satipo, Shimá*); Loreto (*Iquitos, Lower Río Tapiche, Pebas, San Antonio*); Madre de Dios (*Parque Nacional del Manu, Puerto Maldonado, Tambopata Reserve*); San Martín (*Achinamiza, Rioja, Yumbatos*). **Bolivia:** Beni (*Reyes*); La Paz (*Caranavi, Muchanes, Río Zongo*); Santa Cruz (*Santa Cruz de la Sierra, Sara*). **Guyana.** Suriname: Wanica (*Domburg*). **French Guiana:** Cayenne (*Cayenne, Montagne de Kaw, Montagne des Singes*); Saint-Laurent-du-Maroni (*Saint-Laurent-du-Maroni*). **Brazil:** Amazonas (*Benjamin Constant, Manaus, São Paulo de Olivença, Tonantins*); Pará (*Belém, Colares Island, Óbidos*); Acre (*Alto Juruá, Marechal Thaumaturgo*); Rondônia (*Cacaulândia, Candeias do Jamari, Pimenta Bueno, Vilhena*); Mato Grosso (*Diamantino, Pantanal, Vera*); Maranhão; Espírito Santo (*Baixo Guandu, Linhares, Santa Teresa*); Rio de Janeiro; São Paulo (*Campinas, Indiana, Teodoro Sampaio – Parque Estadual do Morro do Diabo*); Paraná (*Castro, Diamante do Norte, São Jorge do Ivaí, Terra Rica*). **Paraguay:** Guairá (*Villarica*); Paraguairí (*Sapucaí*). **Argentina:** Misiones (*General Belgrano*).

TYPE:

Elbella patrobas blanda Evans, 1951. Holotype male with the following labels: / Type – *blanda* Evans/ [Peru, Loreto] Pebas Amazonas [River] M. de Mathan fin X^{brc}. & 1^{er}. Tr. 1880/ R. Oberthür Coll. Brit. Mus. 1931-136/. NHMUK.

***Blubella patrobas* (Hewitson, 1857)**

Six subspecies.

DISTRIBUTION – Southern and eastern Mexico to western and central Colombia, eastern Colombia to eastern Ecuador, northwestern Venezuela and northern Brazil.

***Blubella patrobas patrobas* (Hewitson, 1857)**

Pls. 72–73, Figs. 18 (♂), 19 (♀) (d, v)

DISTRIBUTION – Belize to western and central Colombia, eastern Colombia to central-eastern Bolivia and northwestern Venezuela.

Belize: Corozal (*Copper Bank*). **Guatemala:** Alta Verapaz (*Polochic Valley*); El Petén (*Sayaxché*). **Honduras:** Atlántida (*La Ceiba*); Cortés (*San Pedro Sula*); Olancho (*Petuc*). **El Salvador.** **Nicaragua.** **Costa Rica:** Guanacaste (*Área de Conservación Guanacaste*). **Panama:** Chiriquí (*David*); Colón (*Gamboa, Gatun*); Panamá (*El Llano, Madden Forest*); Panamá Oeste (*Cerro Campana*); Veraguas (*Santa Fe*). **Colombia:** Antioquia (*Valdivia*); Meta (*Río Negro*); Tolima (*Río Chili*). **Ecuador:** Napo (*Yasuni Research Station*). **Venezuela:** Zulia (*El Tucuco*).

TYPE:

Pyrrhopyga [sic] patrobas Hewitson 1857. Lectotype male (O. Mielke 1995) with the following labels: / Type/ Type/ N. Granada [Colombia]. Hewitson Coll. 79-69. *Pyrrhopyga [sic] azeta*. 5./

Lectotipo, *Pyrrhopyga patrobas* Hewitson, 1857, Lectótipo. O. Mielke det. 1993/ *Elbella patrobas patrobas*, O. Mielke det. 1993/. NHMUK.

***Blubella patrobas mexicana* (O. Mielke, 1995)**

Pls. 72–73, Figs. 16 (holotype ♂), 17 (♀) (d, v)

DISTRIBUTION – Southern and eastern Mexico.

Mexico: Chiapas (*Boca de Chajul, Muste, Ocosingo – San Quentin*); Oaxaca (*Chimaloa, Chimalapa, Metates, Palomares, Río Sarabia, Sierra de Juárez, Santa María Tlalixtac*); Quintana Roo (*Nuevo Xcán*); Tabasco (*Tenosique*); Veracruz (*Catemaco, Dos Amates, Los Tuxtlas, Presidio, Santa Rosa, Sierra Blanca, Tamaulipas*).

TYPE:

Elbella patrobas mexicana O. Mielke, 1995. Holotype male with the following labels: / Holotypus/ 3-X-1972 San Quentin, [Ocosingo,] Chiapas, México, Wind leg./ *Elbella patrobas mexicana* [O.] Mielke, Holótipo, O. Mielke det. 1993/ OM 33.023/. OM-DZUP.

***Blubella patrobas melanina* (Mabille & Boullet, 1908)**

Pls. 72–73, Figs. 20 (lectotype ♂ of *Jemadia azeta* var. *melanina*), 21 (♀) (d, v)

DISTRIBUTION – Western and eastern Colombia to central-eastern Bolivia.

Colombia: Amazonian area; Valle del Cauca (*Matangá*). **Ecuador** (?). **Peru.** **Bolivia:** Beni (*Trinidad*).

TYPE:

Jemadia azeta var. *melanina* Mabille & Boullet, 1908. Lectotype male (O. Mielke 1995) with the following labels: / Type/ Type/ Perou 1904 O. Staudinger, Coll. Boullet Museu Paris/ gen[italia]. prep[arada]., [O.] Mielke, 1993/ *J. Azeta* var. *melanina* Mb. & Boull. Ann. Sc. Nat., p. 194 (1908)/ Lectotipo/ *Jemadia azeta* var. *melanina* Mabille & Boullet, 1908, Lectótipo, O. Mielke det. 1993/. MNHN.

***Blubella patrobas evansi* (O. Mielke, 1995)**

Pls. 72–73, Figs. 22–24 (♂) (d, v)

DISTRIBUTION – Western and central Colombia to western Ecuador and northwestern Venezuela.

Colombia: Boyacá (*Otanche*); Caldas (*Manzanares*); Cauca (*Caloto*); Cundinamarca (*Bogotá*); Chocó (*El Tigre, Río Tamandá*); Valle del Cauca (*Buenaventura – Río Dagua, Jiménez*). **Ecuador:** Esmeraldas (*Río Durango*); Imbabura (*Lita*). **Venezuela:** Táchira (*Río Frío, San Cristóbal*).

TYPE:

Elbella patrobas evansi O. Mielke, 1995. Holotype male with the following labels: / Holotipo/ R. Dagua [Valle], Colombia, W. Rosenberg/ Rothschild Bequest B.M. 1939.-1./ *Elbella patrobas evansi* [O.] Mielke, Holótipo, O. Mielke det. 1993/. NHMUK.

***Blubella patrobas amazonica* (O. Mielke, 1995)**

Pls. 74–75, Figs. 1 (holotype ♂), 2 (allotype ♀) (d, v)

DISTRIBUTION – Northern Brazil (Amazonas).

Brazil: Amazonas (*Maués, São Paulo de Olivença*).

TYPE:

Elbella patrobas amazonica O. Mielke, 1995. Holotype male with the following labels: / Holotypus/ São Paulo de Olivença, E. do Amazonas, [Brazil], Coll. E. May-Rio/ gen[italia]. prep[arada]., [O.] Mielke 1986/ 11.883/ *Elbella patrobas amazonica* [O.] Mielke, Holótipo, O. Mielke det. 1993/. OM-DZUP.

***Blubella patrobas tingo* (O. Mielke, 1995)**

Pls. 74–75, Figs. 3 (holotype ♂) (d, v)

DISTRIBUTION – Central-eastern to southeastern Peru and Brazil (Rondônia).

Peru: Cusco (*Cosñipata Valley 600–1,000 m*); Huánuco (*Tingo*

Maria); Junín (*San Ramón, Satipo*); Madre de Dios (*Parque Nacional del Manu, Tambopata Reserve*); San Martín (*Juanjui*). **Brazil:** Rondônia (*Cacaulândia, Pimenta Bueno*).

TYPE:

Elbella patrobas tingo O. Mielke, 1995. Holotype male with the following labels: / Holotypus/ Pimenta Bueno, Rondônia, [Brazil] VIII-IX-1970/ gen[italia]. prep[arada]., [O.] Mielke 1986/ *Elbella patrobas tingo* [O.] Mielke, Holótipo, O. Mielke det. 1993/ DZ 2993/. DZUP.

***Merobella* Grishin, 2019**

One species.

DISTRIBUTION – Costa Rica to central-eastern Bolivia, northwestern Venezuela, French Guiana and northern Brazil.

***Merobella merops* (Bell, 1934)**

Pls. 72–73, Figs. 9 (♂), 10 (♀) (d, v)

DISTRIBUTION – Costa Rica, western and central Colombia, eastern Colombia to central-eastern Bolivia, northwestern Venezuela, French Guiana and northern Brazil.

Costa Rica: Guanacaste (*Área de Conservación Guanacaste*).

Colombia: Boyacá (*Otanche*); Cundinamarca (*Bogotá*); Guaviare (*San José del Guaviare*); Meta (*Villavicencio*); Tolima (*Río Chilí*). **Peru:** Cusco (*Cosñipata Valley – Quebrada Quitacalzón 1,100 m, Quincemil*); Huánuco (*Tingo Maria*); Loreto (*Iquitos*); Madre de Dios (*Parque Nacional del Manu, Tambopata Reserve, Upper Río Madre de Dios – Amazonia Lodge 500 m (near Atalaya, Cusco)*); San Martín (*Tarapoto*). **Bolivia:** La Paz (*Caranavi*); Santa Cruz. **Venezuela:** Táchira (*Parque Nacional El Tamá, Presa Las Cuevas, Río Frío, San Joaquín – Sibéria*); **French Guiana:** Cayenne (*Cayenne*). **Brazil:** Amazonas (*Eirunepé, Parintins (as Vila Nova), Rio Tacana*); Pará (*Óbidos*); Acre (*Porto Walter*); Rondônia (*Alto Paraíso*); Mato Grosso (*Diamantino*).

TYPE:

Jemadia merops Bell, 1934. Holotype male with the following labels: / sp. Columbia/ G 1178/ *Jemadia merops* Bell Holotype ♂/. AMNH.

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Table 1: Barcoded specimens data with collection voucher numbers, GenBank accession numbers, if DNA processed by BOLD or by Nick Grishin (=NG), base pairs, collection depository and country of origin.

Taxon	Specimen number	GenBank number	Bold	NG	Bold - bp	Collection	Country/ State
<i>Agara assaricus mapirica</i>	EB 00 314	GU661983	X		658	EB	Brazil, Amazonas
<i>Agara assaricus mapirica</i>	EB 00 316	GU661984	X		658	EB	Peru, Huánuco
<i>Agara assaricus mapirica</i>	EB 01 889	JF852060	X		658	EB	Peru, Huánuco
<i>Agara assaricus mapirica</i>	EB 01 890	JF852061	X		658	EB	Peru, San Martín
<i>Agara assaricus mapirica</i>	EB 01 891	JF852062	X		658	EB	Peru, San Martín
<i>Agara belti</i> – var.	EB 00 315	GU661985	X		658	EB	Peru, Amazonas
<i>Agara belti belti</i>	03-SNRP-3151	DQ292652	X		617	USNM	Costa Rica, Guanacaste
<i>Agara belti belti</i>	04-SNRP-23918	GU149750	X		658	USNM	Costa Rica, Guanacaste
<i>Agara belti belti</i>	04-SNRP-31804	GU149745	X		658	USNM	Costa Rica, Guanacaste
<i>Agara belti belti</i>	04-SNRP-33998	GU149746	X		658	USNM	Costa Rica, Guanacaste
<i>Agara belti belti</i>	04-SNRP-3418	GU149748	X		658	USNM	Costa Rica, Alajuela
<i>Agara belti belti</i>	04-SNRP-34960	GU149747	X		658	USNM	Costa Rica, Guanacaste
<i>Agara belti belti</i>	04-SRNP-23919	GU149749	X		658	USNM	Costa Rica, Guanacaste
<i>Agara belti belti</i>	05-SNRP-2091	GU150559	X		658	USNM	Costa Rica, Alajuela
<i>Agara belti belti</i>	EB 01 276	MT885225	X		307	USNM	Mexico, Veracruz
<i>Amenis pionia pedro</i>	DZ 8.704	GU661452	X		658	DZUP	Brazil, Bahia
<i>Amenis pionia pedro</i>	DZ 8.720	GU661451	X		658	DZUP	Brazil, Bahia
<i>Amenis pionia pedro</i>	OM 67.482	GU661453	X		658	OM	Brazil, Ceará
<i>Amenis pionia pionia</i>	OM 41.053	HQ941742	X		658	OM	Paraguay, Alto Paraguay
<i>Amenis pionia pionia</i>	OM 51.450	GU661450	X		658	OM	Argentina, Salta
<i>Apatiella iphinous iphinous</i>	EB 00 346	MT885215	X		658	EB	Brazil, São Paulo
<i>Apatiella iphinous iphinous</i>	EB 00 347	GU662036	X		658	EB	Brazil, São Paulo
<i>Apatiella iphinous iphinous</i>	EB 00 348	GU662037	X		658	EB	Brazil, São Paulo
<i>Apatiella iphinous mariae</i>	EB 00 215	GU662276	X		658	EB	Brazil, Paraná
<i>Apatiella iphinous mariae</i>	EB 00 216	GU662273	X		658	EB	Brazil, Paraná
<i>Apatiella iphinous mariae</i>	EB 00 217	GU662274	X		658	EB	Brazil, Santa Catarina
<i>Apatiella iphinous mariae</i>	EB 00 218	GU662272	X		658	EB	Brazil, Santa Catarina
<i>Apyrrothrix araxes araxes</i>	EB 01 258	HQ567052	X		658	EB	Mexico, Durango
<i>Apyrrothrix araxes araxes</i>	EB 01 259	HQ567053	X		658	EB	Mexico, Nayarit
<i>Apyrrothrix araxes araxes</i>	EB 01 260	HQ567054	X		658	EB	Mexico, Sonora
<i>Apyrrothrix araxes araxes</i>	EB 01 261	HQ567055	X		658	EB	Mexico, Sonora
<i>Apyrrothrix araxes araxes</i>	EB 02 987	MT885209	X		658	EB	Mexico, Jalisco
<i>Apyrrothrix araxes cyrillus</i>	No number, female	MT992640	----	X	----	MfN	Mexico, Oaxaca
<i>Apyrrothrix. araxes cyrillus</i>	Holotype male	MT992639	----	X	----	MfN	Mexico, Oaxaca
<i>Aspitha agenoria agenoria</i>	EB 00 024	GU661971	X		658	EB	Peru, Huánuco
<i>Aspitha agenoria agenoria</i>	EB 00 025	GU661972	X		658	EB	Peru, Huánuco
<i>Aspitha agenoria agenoria</i>	EB 00 026	GU661970	X		658	EB	Peru, Huánuco
<i>Aspitha agenoria agenoria</i>	EB 00 032	GU661969	X		658	EB	Preu, Huánuco
<i>Aspitha agenoria agenoria</i>	EB 00 035	GU661968	X		658	EB	Peru, Huánuco
<i>Croniades auraria</i>	EB 00 374	HM422904	X		658	EB	Peru, Amazonas
<i>Croniades pieria</i>	EB 00 373	HM422903	X		658	EB	Brazil, Amazonas
<i>Gunayan rubricollis flemingi</i>	OM 76.805	MT885213	X		658	OM	Colombia, Meta
<i>Gunayan rubricollis rubricollis</i>	DZ 16.098	G661335	X		658	DZUP	Brazil, Acre
<i>Gunayan rubricollis rubricollis</i>	DZ 16.456	GU661331	X		658	DZUP	Brazil, Pará
<i>Gunayan rubricollis rubricollis</i>	EB 00 375	GU662025	X		658	EB	Peru, Loreto
<i>Gunayan rubricollis rubricollis</i>	EB 01 529	MT885212	X		658	EB	Peru, Huánuco
<i>Gunayan rubricollis rubricollis</i>	EB 01 530	MT885214	X		658	EB	Peru, Amazonas
<i>Gunayan rubricollis rubricollis</i>	EB 02 238	JN278054	X		658	EB	Brazil, Pará
<i>Gunayan rubricollis rubricollis</i>	DZ 16.049	GU661336	X		658	DZUP	Brazil, Acre
<i>Jemasonia albesens</i>	EB 00 401	GU662087	X		658	EB	Peru, San Martín
<i>Jemasonia albesens</i>	EB 00 402	GU662084	X		658	EB	Peru, San Martín
<i>Jemasonia albesens</i>	EB 02 773	MT885219	X		572	EB	Ecuador
<i>Jemasonia hewitsonii hewitsonii</i>	EB 00 401	GU 662087	X		658	EB	Peru, San Martín
<i>Jemasonia hewitsonii hewitsonii</i>	EB 00 402	GU 662084	X		658	EB	Peru, San Martín
<i>Jematus gnetus brevipennis</i>	EB 00 523	HM394438	X		658	EB	Paraguay
<i>Jematus gnetus brevipennis</i>	EB 00 524	HM394439	X		658	OM	Brazil, Minas Gerais
<i>Jematus gnetus gnetus</i>	EB 00 099	HM422896	X		658	EB	Peru, Junín
<i>Jematus gnetus gnetus</i>	OM 53.924	JX216188	X		658	EB	Brazil, Rondônia

Taxon	Specimen number	GenBank number	Bold	NG	Bold - bp	Collection	Country/ State
<i>Mahotis eupheme</i>	EB 01 545	MT885210	X		658	EB	Peru, Cusco
<i>Mahotis eupheme</i>	EB 01 546	MT885202	X		658	EB	Peru, Cusco
<i>Mahotis eupheme</i>	EB 01 547	HQ567331	X		658	EB	Argentina, Salta
<i>Mahotis eupheme</i>	EB 01 548	MT885224	X		658	EB	Argentina, Salta
<i>Mahotis versicolor</i>	EB 00 413	GU662291	X		658	EB	Brazil, Minas Gerais
<i>Mahotis versicolor</i>	EB 00 414	GU662292	X		658	EB	Brazil, Paraná
<i>Mahotis versicolor</i>	EB 00 415	GU662058	X		595	EB	Brazil, Minas Gerais
<i>Mimardaris periphema</i>	EB 00 381	GU662085	X		658	EB	Bolivia, Cochabamba
<i>Mimardaris periphema</i>	EB 00 382	GU662082	X		658	EB	Bolivia, Cochabamba
<i>Mimardaris periphema</i>	EB 00 399	GU662289	X		658	EB	Bolivia, La Paz
<i>Mimardaris sela peruviana</i>	EB 00 247	GU662263	X		658	EB	Peru, San Martín
<i>Mimardaris sela peruviana</i>	EB 00 392	GU662079	X		658	EB	Peru, San Martín
<i>Mimardaris sela peruviana</i>	EB 00 395	GU662119	X		589	EB	Peru, Amazonas
<i>Mimardaris sela peruviana</i>	EB 02 592	MT885206	X		658	EB	Peru, Huánuco
<i>Myscelus amystis hages</i>	00-SRNP-20049	DQ292644	X		583	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	00-SRNP-6023	DQ292643	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	04-SRNP-21290	DQ292645	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	04-SRNP-24102	GU161671	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	04-SRNP-26567	GU150557	X		674	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20288	GU150558	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20426	GU150556	X		373	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20428	GU150554	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20512	GU150555	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20514	GU150552	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20589	GU150553	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	05-SRNP-20706	GU150551	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis hages</i>	96-SRNP-365	DQ292642	X		658	USNM	Costa Rica, Guanacaste
<i>Myscelus amystis mysus</i>	EB 00 318	GU661988	X		658	EB	Peru, San Martín
<i>Myscelus amystis mysus</i>	EB 00 319	GU661989	X		658	EB	Peru, San Martín
<i>Myscelus amystis mysus</i>	EB 01 277	MT885200	X		307	EB	Peru, Loreto
<i>Myscelus epigona</i>	EB 00 211	HM422897	X		658	EB	Brazil, Paraná
<i>Myscelus epigona</i>	EB 00 583	HM394491	X		640	EB	Argentina, Misiones
<i>Myscelus epigona</i>	EB 00 584	HM394492	X		658	EB	Brazil, Santa Catarina
<i>Myscelus epigona</i>	EB 01 892	JF852063	X		658	EB	Brazil, Santa Catarina
<i>Myscelus epigona</i>	EB 01 893	JF852064	X		658	EB	Brazil, Santa Catarina
<i>Myscelus epigona</i>	EB 02 709	MT885229	X		658	EB	Brazil, Santa Catarina
<i>Myscelus phoronis phoronis</i>	EB 00 307	HM422901	X		658	EB	Peru, San Martín
<i>Myscelus phoronis phoronis</i>	EB 00 322	MT885220	X		658	EB	Peru, Amazonas
<i>Myscelus phoronis phoronis</i>	EB 00 323	MT885221	X		658	EB	Ecuador, Tungurahua
<i>Myscelus phoronis phoronis</i>	EB 01 008	HQ567203	X		658	EB	Bolivia, La Paz
<i>Myscelus phoronis phoronis</i>	EB 01 009	HQ567204	X		658	EB	Bolivia, La Paz
<i>Myscelus phoronis phoronis</i>	EB 02 603	MT885228	X		658	EB	Ecuador, Morona-Santiago
<i>Myscelus phoronis phoronis</i>	EB 02 604	MT885218	X		658	EB	Ecuador, Morona-Santiago
<i>Myscelus santhilarius</i>	EB 00 321	GU661986	X		658	EB	Brazil, Santa Catarina
<i>Pyrrhopyge amythaon podina</i>	OM 60.579	GU661376	X		658	OM	Bolivia, La Paz
<i>Pyrrhopyge amythaon podina</i>	OM 60.999	GU661481	X		658	OM	Bolivia, La Paz
<i>Pyrrhopyge amythaon podina</i>	OM 61.138	GU661492	X		658	OM	Bolivia, La Paz
<i>Pyrrhopyge amythaon podina</i>	OM 61.452	GU661488	X		658	OM	Bolivia, La Paz
<i>Pyrrhopyge attis attis</i>	OM 61.475	GU661370	X		658	OM	Bolivia, LaPaz
<i>Pyrrhopyge attis lexos</i>	DZ 16.119	GU661340	X		658	DZUP	Brazil, Acre
<i>Pyrrhopyge attis lexos</i>	EB 01 233	HQ567041	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge attis lexos</i>	EB 01 236	HQ567044	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge attis lexos</i>	EB 01 313	HQ567083	X		658	EB	Peru, Loreto
<i>Pyrrhopyge attis lexos</i>	EB 01 313	HQ567083	X		658	EB	Peru, Loreto
<i>Pyrrhopyge aziza araethyrea</i>	EB 01 106	HM394798	X		658	EB	Ecuador, Tungurahua
<i>Pyrrhopyge aziza araethyrea</i>	OM 26.851	GU661497	X		658	OM	Peru, San Martín
<i>Pyrrhopyge croceimargo</i>	EB 00 295	GU662016	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge croceimargo</i>	EB 01 089	HM394787	X		658	EB	Peru, Cusco
<i>Pyrrhopyge croceimargo</i>	EB 01 091	HM394789	X		658	EB	Peru, Cusco
<i>Pyrrhopyge draudti bruno</i>	OM 77.877	MT885216	X		658	OM	Colombia, Meta
<i>Pyrrhopyge draudti bruno</i>	OM 37.102	MT992641	----	X	----	OM	Colombia, Meta

Taxon	Specimen number	GenBank number	Bold	NG	Bold - bp	Collection	Country/ State
<i>Pyrrhopyge hadassa</i>	EB 00 432	GU662100	X		604	EB	Peru, Amazonas
<i>Pyrrhopyge hadassa</i>	EB 00 433	GU662295	X		658	EB	Ecuador, Tungurahua
<i>Pyrrhopyge halma</i>	EB 00 294	GU662002	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge halma</i>	EB 00 302	GU662001	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge hanga</i>	EB 01 829	JF852013	X		658	EB	Peru, Junín
<i>Pyrrhopyge hanga</i>	EB 01 830	JF852014	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge jola</i>	OM 53.929	GU661372	X		658	OM	Peru, Cusco
<i>Pyrrhopyge jola</i>	OM 54.097	GU661363	X		658	OM	Peru, Cusco
<i>Pyrrhopyge jola</i>	OM 54.245	GU661364	X		658	OM	Peru, Cusco
<i>Pyrrhopyge jola</i>	OM 76.570	MT885203	X		635	OM	Peru, Cusco
<i>Pyrrhopyge josephina josephina</i>	OM 60.489	GU661356	X		658	OM	Peru, Amazonas
<i>Pyrrhopyge leticia</i>	EB 01 076	HM394775	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge leticia</i>	EB 01 077	HM394776	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge mendax</i>	EB 01 073	HM394772	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge mendax</i>	EB 01 074	HM394773	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge mendax</i>	EB 01 078	HM394777	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 00 248 Paratype	GU662277	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 00 249 Paratype	GU662278	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 00 457 Paratype	GU662099	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 01 112 Paratype	HM394801	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge michael</i>	EB 01 118 Paratype	HM394807	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge michael</i>	EB 01 120 Paratype	HM394808	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge michael</i>	EB 01 246 Paratype	HQ567047	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge michael</i>	EB 01 256 Paratype	HQ567051	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 02 569 Paratype	MT885201	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 02 570 Paratype	MT885226	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 02 572 Paratype	MT885227	X		552	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 02 573 Paratype	JN278058	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 14 046 Paratype	HQ566926	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 14 050 Paratype	HQ566929	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 14 054 Paratype	HQ566932	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 14 055 Paratype	HQ566933	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 14 057 Paratype	HQ566935	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 14 065 Paratype	HQ566941	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 14 069 Paratype	HQ566943	X		658	EB	Peru, San Martín
<i>Pyrrhopyge michael</i>	EB 14 085 Paratype	HQ566949	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 14 087 Paratype	HQ566950	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	EB 14 088 Paratype	HQ566951	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge michael</i>	OM 47.078 Allotype	GU661357	X		658	OM	Peru, Huánuco
<i>Pyrrhopyge michael</i>	OM 60.266 Paratype	GU661372	X		658	OM	Peru, San Martín
<i>Pyrrhopyge mina ganus</i>	Holotype	MT992644	----	X	----	AMNH	Guyana
<i>Pyrrhopyge mina mina</i>	DZ 15.583	GU661346	X		658	DZUP	Brazil, Mato Grosso
<i>Pyrrhopyge mina mina</i>	DZ 16.112	GU661341	X		658	DZUP	Brazil, Acre
<i>Pyrrhopyge mina mina</i>	EB 14 017	HQ566909	X		658	EB	Peru, Madre de Dios
<i>Pyrrhopyge mina mina</i>	OM 77.902	MT885207	X		658	OM	Peru, Cusco
<i>Pyrrhopyge mina mina</i>	OM 78.195	MT885205	X		658	OM	Peru, Cusco
<i>Pyrrhopyge mina mina</i>	Holotype	MT992642	----	X	----	AMNH	Bolivia
<i>Pyrrhopyge mina mina</i>	Holotype - P. s. bolius	MT992643	----	X	----	AMNH	Bolivia, La Paz
<i>Pyrrhopyge mina previsto</i>	EB 01 119 Holotype	MT885211	X		307	EB	Peru, Ucayali
<i>Pyrrhopyge olivacea</i>	EB 00 254	GU662239	X		658	EB	Peru, San Martín
<i>Pyrrhopyge olivacea</i>	EB 00 440	GU662101	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge olivacea</i>	EB 00 441	GU662098	X		658	EB	Peru, San Martín
<i>Pyrrhopyge olivacea</i>	EB 01 836	JF852017	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge phaeax</i>	EB 01 081	HM394780	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge phaeax</i>	EB 01 082	HM394781	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge phaeax</i>	EB 01 084	HM394783	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge phaeax</i>	EB 01 088	HM394786	X		658	EB	Peru, Amazonas
<i>Pyrrhopyge phidias leucoloma</i>	OM 23.468	GU661359	X		658	OM	Peru, Junín
<i>Pyrrhopyge phidias phidias</i>	OM 54.090	GU661362	X		658	OM	Peru, Cusco
<i>Pyrrhopyge phidias phidias</i>	OM 61.014	GU661489	X		658	OM	Bolivia, La Paz
<i>Pyrrhopyge phidias phidias</i>	OM 61.035	GU661491	X		658	OM	Bolivia, La Paz

Taxon	Specimen number	GenBank number	Bold	NG	Bold - bp	Collection	Country/ State
<i>Pyrrhopyge phidias phidias</i>	OM 61.182	GU661486	X		658	OM	Bolivia, La Paz
<i>Pyrrhopyge phidias rusca</i>	EB 01 223	HQ567036	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge phidias rusca</i>	EB02 563	MT885223	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge phylleia</i>	EB 14 002	HQ566898	X		569	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 003	HQ566899	X		618	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 005	HQ566901	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 009	HQ566903	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 010	HQ566904	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 012	HQ566906	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 013	HQ566907	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 00 277	HM394390	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 00 278	GU662280	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 01 016	HQ567211	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 01 018	HQ567213	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 001	HQ566897	X		636	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 004	HQ566900	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 006	HQ566902	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge phylleia</i>	EB 14 011	HQ566905	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge pseudohadassa</i>	EB 00 481	HM394398	X		658	EB	Peru, Junín
<i>Pyrrhopyge pseudohadassa</i>	EB 01 835	JF852016	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge rafael</i>	EB 01 107	HM394799	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge rafael</i>	EB 01 110	MT885217	X		307	EB	Peru, Huánuco
<i>Pyrrhopyge rafael</i>	EB 01 113	HM394802	X		658	EB	Peru, Huánuco
<i>Pyrrhopyge rafael</i>	OM 35.949	GU661368	X		658	OM	Peru, Huánuco
<i>Pyrrhopyge sergius andronicus</i>	EB 00 229	GU662281	X		619	EB	Ecuador, Napo
<i>Pyrrhopyge sergius andronicus</i>	EB 02 787	MT885222	X		658	EB	Ecuador, Morona-Santiago
<i>Pyrrhopyge sergius andronicus</i>	EB 14 035	HQ566919	X		658	EB	Ecuador, Morona-Santiago
<i>Pyrrhopyge sergius andronicus</i>	Holotype	MT992645	-----	X	-----	AMNH	Ecuador, Morona-Santiago
<i>Pyrrhopyge sergius sergius</i>	15 081G12	MT992646	-----	X	----	EB	Peru, San Martín
<i>Pyrrhopyge sergius sergius</i>	15 081H01	MT992647	-----	X	----	EB	Peru, San Martín
<i>Pyrrhopyge sergius sergius</i>	15 081H02	MT992648	-----	X	----	EB	Peru, San Martín
<i>Pyrrhopyge sergius sergius</i>	Lectotype	MT992649	-----	X	----	MfN	Peru, Junín
<i>Pyrrhopyge sergius sergius</i>	OM 23.585	GU661485	X		658	OM	Peru, Junín
<i>Pyrrhopyge shiva</i>	EB 00 285	HM394395	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge shiva</i>	EB 00 336	GU662010	X		658	EB	Bolivia, La Paz
<i>Pyrrhopyge telassa</i>	EB 00 468	GU662102	X		658	EB	Ecuador, Tungurahua
<i>Pyrrhopyge telassa</i>	EB 02 101	IN278050	X		658	EB	Ecuador
<i>Pyrrhopyge telassina</i>	EB 00 253	GU662249	X		658	EB	Peru, San Martín
<i>Pyrrhopyge telassina</i>	EB 00 507	HM394423	X		658	EB	Bolivia, Beni
<i>Yanguna aspilos lara</i>	EB 01 097	MT885208	X		307	EB	Peru, Huánuco
<i>Yanguna aspilos lara</i>	EB 01 098	HM394793	X		658	EB	Peru, Huánuco
<i>Yanguna aspilos lara</i>	EB 01 099	HM394794	X		658	EB	Peru, Huánuco
<i>Yanguna aspilos lara</i>	EB 01 100	HM394795	X		658	EB	Peru, Huánuco
<i>Yanguna blossomiae blossomiae</i>	DZ 29.386	MT992651	-----	X	-----	DZUP	Brazil, Acre
<i>Yanguna blossomiae blossomiae</i>	Holotype	MT992650	-----	X	-----	CMP	Ecuador, Chimborazo
<i>Yanguna blossomiae blossomiae</i>	OM 79.356	MT992652	-----	X	-----	OM	Peru, Loreto
<i>Yanguna blossomiae galion</i>	OM 71.417 Allotype	MT992654	-----	X	-----	OM	French Guiana, Cayenne
<i>Yanguna blossomiae galion</i>	OM 71.427 Holotype	MT992653	-----	X	-----	OM	French Guiana, Cayenne
<i>Yanguna moa guiana</i>	OM 71.407 Holotype	MT992657	-----	X	-----	OM	French Guiana, Cayenne
<i>Yanguna moa moa</i>	OM 36.018 Holotype	MT992655	-----	X	-----	OM	Peru, Huánuco
<i>Yanguna moa moa</i>	OM 36.048 Paratype	MT992656	-----	X	-----	OM	Peru, Huánuco
<i>Yanguna spatiosa spatiosa</i>	EB 00 469	GU662302	X		658	EB	Peru, San Martín
<i>Yanguna spatiosa spatiosa</i>	EB 00 471	GU662303	X		658	EB	Ecuador, Tungurahua
<i>Yanguna spatiosa spatiosa</i>	EB 00 472	GU662120	X		637	EB	Peru, Amazonas
<i>Yanguna spatiosa spatiosa</i>	EB 02 090	JN278046	X		658	EB	Ecuador
<i>Yanguna spatiosa spatiosa</i>	EB 02 091	JN278047	X		658	EB	Ecuador
<i>Yanguna spatiosa spatiosa</i>	EB 02 775	MT885204	X		623	EB	Ecuador, Morona-Santiago
<i>Yanguna thelersa</i>	DZ 28.784	MT992658	-----	X	-----	DZUP	Brazil, Acre
<i>Yanguna thelersa</i>	EB	MT992661	-----	X	-----	EB	Peru, Loreto
<i>Yanguna thelersa</i>	OM 21.709	MT992659	-----	X	-----	OM	Brazil, Pará
<i>Yanguna thelersa</i>	OM 71.487	MT992660	-----	X	-----	OM	French Guiana, Cayenne

Checklist (alphabetical)

At the right column, the type locality is given: country and state, department or province (or similar) and the name bearing type.

Hesperiidae (Latreille, 1809) Stephens, 1828

Pyrrhopyginae (Mabille, 1877) Watson, 1893

Azonaxini Grishin, 2019

Azonax Godman & Salvin, 1893

guiana O. Mielke, Brockmann & C. Mielke, **sp. n.**

French Guiana (Cayenne) - HT

typhaon (Hewitson, 1877) (*Myscelus*)

[Nicaragua] - LT

urupa O. Mielke, Brockmann & C. Mielke, **sp. n.**

Brazil (Rondônia) - HT

Jerini Grishin, 2019

Jera Lindsey, 1925

tricuspidata (Mabille, 1902) (*Dichelura*)

Ecuador - HT

Oxynetrini O. Mielke, 2001

Olafia Nemésio, 2005

= *Cyclopyge* O. Mielke, 2002, **praeocc.** (Hafle & Corda, 1847) (Trilobita)

roscius (Hopffer, 1874) (*Pyrrhopyge*)

a) *roscius roscius* (Hopffer, 1874) (*Pyrrhopyge*)

Brazil - LT

= *rufocincta* (Hayward, 1932) (*Oxynetra*)

Argentina (Misiones) - HT

= *flavomaculata* (Bell, 1937) (*Oxynetra*), **syn. n.**

Brazil (Santa Catarina) - HT

b) *roscius iphimedia* (Plötz, 1886) (*Pyrrhopyga* [**sic**])

Brazil (São Paulo) - NT

= *erythrosoma* (Mabille, 1891) (*Pyrrhopyga* [**sic**])

Brazil (São Paulo) - LT

c) *roscius koikei* O. Mielke, Brockmann & C. Mielke, **ssp. n.**

Brazil (Rio de Janeiro) - HT

Oxynetra C. Felder & R. Felder, 1862

confusa Staudinger, 1888

a) *confusa confusa* Staudinger, 1888

Peru (Junín) - LT

b) *confusa franciscana* Salazar & Vargas, [2017]

Panama (Darién) - HT

semihyalina C. Felder & R. Felder, 1862

[Peru] - HT

= *felderi* Hopffer, 1874

Peru (Junín) - LT

Dis Mabille, 1889

aureopecta (Warren & Grishin, 2017) (*Oxynetra*)

Mexico (Hidalgo) - HT

hopfferi (Staudinger, 1888) (*Oxynetra*)

Panama (Chiriquí) - HT

= *annulatus* Mabille, 1889

Panama (Chiriquí) - HT

stangelandi (Grishin & Burns, 2013) (*Oxynetra*)

Costa Rica (Alajuela) - HT

Passovini O. Mielke, 2001

Agara Mabille & Boulet, 1908

assaricus (Cramer, 1779) (*Papilio*)

a) *assaricus assaricus* (Cramer, 1779) (*Papilio*)

Suriname - LT

= *janthibaris* (Butler, [1870]) (*Papilio*), **nom. nud.** (ICZN. Art. 11.6)

Brazil

b) *assaricus mapirica* Strand, 1921

Bolivia (La Paz) - HT

belti (Godman & Salvin, 1879) (*Myscelus*)

a) *belti belti* (Godman & Salvin, 1879) (*Myscelus*)

Nicaragua (Chontales) - LT

b) *belti caucaensis* (Constantino, Salazar & Vargas, 2013) (*Myscelus*)

Colombia (Caldas) - HT

draudti (Riley, 1926) (*Myscelus*)

Bolivia (La Paz) - HT

epimachia (Herrich-Schäffer, 1869) (*Myscelus*)

a) *epimachia epimachia* (Herrich-Schäffer, 1869) (*Myscelus*)

Peru (Huánuco) - NT

b) *epimachia edix* (Evans, 1951) (*Myscelus*)

Paraguay (Sapucay) - HT

c) *epimachia hermeri* O. Mielke, Brockmann & C. Mielke, **ssp. n.**

French Guiana (Cayenne) - HT

d) <i>epimachia nevada</i> O. Mielke & Casagrande, ssp. n.	Colombia (Magdalena) - HT
e) <i>epimachia verina</i> (Evans, 1951) (<i>Myscelus</i>)	Ecuador (Zamora-Chinchi) - HT
<i>gallardi</i> O. Mielke, Brockmann & C. Mielke, sp. n.	French Guiana (Saint-Laurent-du-Maroni) - HT
<i>michaeli</i> (Nicolay, 1975) (<i>Myscelus</i>)	Panama (Panamá) - HT
<i>pardalina</i> (C. Felder & R. Felder, 1867) (<i>Tamyris</i>)	
a) <i>pardalina pardalina</i> (C. Felder & R. Felder, 1967) (<i>Tamyris</i>)	Colombia (Cundinamarca) - LT
= <i>aurora</i> Röber, 1925	Ecuador (Morona-Santiago) - HT
b) <i>pardalina guarea</i> (Evans, 1951) (<i>Myscelus</i>)	Brazil (Pará) - HT
c) <i>pardalina kesselringi</i> (O. Mielke & Casagrande, 2011) (<i>Myscelus</i>)	Peru (Huánuco) - HT
d) <i>pardalina yacutinga</i> (O. Mielke & Casagrande, 2011) (<i>Myscelus</i>)	Argentina (Misiones) - HT
<i>pegasus</i> (Mabille, 1903) (<i>Myscelus</i>)	French Guiana (Cayenne) - LT
<i>perissodora</i> (Dyar, 1914) (<i>Myscelus</i>)	Mexico (Veracruz) - LT
<i>santhilarius</i> (Latreille, [1824]) (<i>Hesperia</i>)	Brazil - LT
<i>Aspitha</i> Evans, 1951	
<i>agenoria</i> (Hewitson, 1876) (<i>Pyrrhopyga</i> [sic])	
a) <i>agenoria agenoria</i> (Hewitson, 1876) (<i>Pyrrhopyga</i> [sic])	Peru (Junín) - LT
= <i>sanies</i> (H. H. Druce, 1908) (<i>Pyrrhopyge</i>), syn. n.	Bolivia (La Paz) - LT
b) <i>agenoria cruor</i> (H. H. Druce, 1908) (<i>Pyrrhopyge</i>)	Peru (Pasco) - LT
<i>aspitha</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	
a) <i>aspitha aspitha</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
b) <i>aspitha parima</i> (Plötz, 1886) (<i>Pyrrhopyga</i> [sic])	Suriname - NT
c) <i>aspitha rufescens</i> (Riley, 1919) (<i>Yanguna</i>)	Brazil (Rondônia) - HT
<i>bassleri</i> (Bell, 1940) (<i>Pyrrhopyge</i>)	Peru (San Martín) - HT
<i>leander</i> (Boullet, 1912) (<i>Yanguna</i>)	Colombia (Boyacá) - HT
<i>teffa</i> Evans, 1951, stat. rev.	Brazil (Amazonas) - HT
<i>Granila</i> Mabille, 1903	
<i>paseas</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic])	Brazil - LT
= <i>paseas</i> var. <i>albimacula</i> Mabille & Boullet, 1908	Brazil - HT
= <i>paseas</i> var. <i>pullata</i> J. Zikán, 1938	Brazil (Rio de Janeiro) - HT
<i>Myscelus</i> Hübner, [1819]	
<i>amystis</i> (Hewitson, 1867) (<i>Erycides</i>)	
a) <i>amystis amystis</i> (Hewitson, 1867) (<i>Erycides</i>)	Colombia - LT
= <i>rogersi</i> Kaye, 1914	Trinidad and Tobago (Siparia) - LT
= <i>flavicollis</i> Röber, 1925	Venezuela (Zulia) - NT
b) <i>amystis distinctus</i> Röber, 1925	Ecuador (Napó) - NT
c) <i>amystis hages</i> Godman & Salvin, 1893	Panama (Chiriquí) - LT
d) <i>amystis muttra</i> Evans, 1951	Colombia (Valle del Cauca) - HT
e) <i>amystis mysus</i> Evans, 1951	Peru (San Martín) - HT
<i>epigona</i> (Herrich-Schäffer, 1869), stat. rest.	Brazil (Santa Catarina) - NT
= <i>orthrus</i> Hewitson, 1877	[Brazil] - LT
= <i>sothis</i> (Mabille, 1883) (<i>Pyrrhopyga</i> [sic])	Brazil - LT
= <i>orbis</i> (Mabille, 1883) (<i>Pyrrhopyga</i> [sic])	Brazil - LT
= <i>nobilis</i> var. (ssp.) <i>meridionalis</i> Röber, 1925	Brazil (Santa Catarina) - LT
<i>illustris</i> Mabille, 1903, stat. rev.	Bolivia - LT
<i>nobilis</i> (Cramer, 1777) (<i>Papilio</i>)	no locality - LT
= <i>salus</i> (Fabricius, 1781) (<i>Papilio</i>), nom. n.	Suriname - LT
<i>phoronis</i> (Hewitson, 1867) (<i>Erycides</i>)	
a) <i>phoronis phoronis</i> (Hewitson, 1867) (<i>Erycides</i>)	Colombia - LT
b) <i>phoronis caucanus</i> Staudinger, 1888	Colombia (Valle del Cauca) - LT
= <i>persela</i> (Mabille, 1891) (<i>Pyrrhopyga</i> [sic])	Colombia (Cauca) - LT
<i>Passova</i> Evans, 1951	
<i>ganymedes</i> (Bell, 1931) (<i>Pyrrhopyge</i>)	
a) <i>ganymedes ganymedes</i> (Bell, 1931) (<i>Pyrrhopyge</i>)	Colombia - HT
b) <i>ganymedes gelina</i> Evans, 1951	Peru (Junín) - HT
c) <i>ganymedes gulia</i> Evans, 1951	Ecuador (Loja) - HT
<i>gazera</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT

<i>gellias</i> (Godman & Salvin, 1893) (<i>Pyrrhopyge</i>)	Costa Rica - HT
<i>glacia</i> Evans, 1951	French Guiana - HT
<i>greta</i> Evans, 1951	Peru (Pasco) - HT
<i>nigrocephala</i> (Bell, 1934) (<i>Pyrrhopyge</i>)	Colombia - HT
<i>passova</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	
a) <i>passova passova</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
b) <i>passova diamantino</i> O. Mielke, C. Mielke & Casagrande, ssp. n.	Brazil (Mato Grosso) - HT
c) <i>passova gortyna</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
d) <i>passova practa</i> Evans, 1951	Brazil (Paraná) - HT
e) <i>passova rudex</i> Evans, 1951	Peru (Junín) - HT
f) <i>passova stacer</i> Evans, 1951	Guyana - HT
g) <i>passova styx</i> (Möschler, 1879) (<i>Pyrrhopyga</i> [sic])	Colombia - LT
= <i>anina</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), nom. nud. (ICZN, Art. 11.6.)	Colombia
<i>polemon</i> (Hopffer, 1874) (<i>Pyrrhopyge</i>)	Brazil ([Bahia]) - LT
<i>vilna</i> Evans, 1951	
a) <i>vilna vilna</i> Evans, 1951	Bolivia (Beni) - HT
b) <i>vilna vanta</i> Evans, 1951	Bolivia - HT
Pyrrhopygini Mabille, 1877	
Apyrrothrixina Grishin, 2019	
<i>Aesculapyge</i> Grishin, 2019	
<i>aesculapus</i> (Staudinger, 1876) (<i>Pyrrhopyge</i>)	Panama (Chiriquí) - LT
= <i>variegaticeps</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic])	Costa Rica - LT
<i>Apyrrothrix</i> Lindsey, 1921	
<i>araxes</i> (Hewitson, 1867) (<i>Erycides</i>)	
a) <i>araxes araxes</i> (Hewitson, 1867) (<i>Erycides</i>)	Mexico – LT
b) <i>araxes cyrillus</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), stat. rev.	Mexico (Oaxaca) - HT
<i>arizonae</i> (Godman & Salvin, 1893) (<i>Pyrrhopyge</i>)	United States of America (Arizona) - LT
<i>Chalypyge</i> O. Mielke, 2002	
<i>chalybea</i> (Scudder, 1872) (<i>Pyrrhopyga</i> [sic])	
a) <i>chalybea chalybea</i> (Scudder, 1872) (<i>Pyrrhopyga</i> [sic])	Mexico - LT
b) <i>chalybea chloris</i> (Evans, 1951) (<i>Pyrrhopyge</i>)	Mexico (Guerrero) - HT
<i>rufinucha</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic]), stat. rest.	Bolivia - LT
= <i>apollo</i> (Röber, 1925) (<i>Pyrrhopyge</i>)	Bolivia (La Paz) - LT
<i>zereda</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	
a) <i>zereda zereda</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Ecuador - LT
b) <i>zereda hygieia</i> (C. Felder & R. Felder, 1867) (<i>Tamyris</i>)	Colombia (Cundinamarca) - LT
= <i>bogotana</i> (Reakirt, 1868) (<i>Pyrrhopyga</i> [sic])	Colombia (Cundinamarca) - LT
= <i>phoebus</i> (Röber, 1925) (<i>Pyrrhopyge</i>)	Colombia (Valle del Cauca) - NT
c) <i>zereda insana</i> (Staudinger, 1876) (<i>Pyrrhopyge</i>)	Peru [Junín] - LT
= <i>fassli</i> (Boullet, 1910) (<i>Pyrrhopyge</i>)	Peru (Pasco) - LT
d) <i>zereda rufipectus</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic])	Ecuador (Tungurahua) - LT
e) <i>zereda zepha</i> (Evans, 1951) (<i>Pyrrhopyge</i>)	Ecuador (Zamora-Chinchipe) - HT
<i>Melanopyge</i> O. Mielke, 2002	
<i>burnsi</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Panama (Chiriquí) - HT
<i>cossea</i> (H. Druce, 1875) (<i>Pyrrhopyga</i> [sic])	Colombia - LT
<i>erythrosticta</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic])	Nicaragua (Chontales) - LT
<i>hoffmanni</i> (Freeman, 1977) (<i>Pyrrhopyge</i>)	Mexico (Tabasco) - HT
<i>maculosa</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Colombia (Cundinamarca) - LT
= <i>agathon</i> (C. Felder & R. Felder, 1867) (<i>Tamyris</i>)	Colombia (Cundinamarca) - LT
<i>mulleri</i> (Bell, 1934) (<i>Apyrrothrix</i>)	Central America - HT
= <i>plancartei</i> (C. Hoffmann, 1941) (<i>Apyrrothrix</i>)	Mexico (Veracruz) - HT
<i>Creonpyge</i> O. Mielke, 2002	
<i>creon</i> (H. Druce, 1874) (<i>Pyrrhopyga</i> [sic])	
a) <i>creon creon</i> (H. Druce, 1874) (<i>Pyrrhopyga</i> [sic])	Panama (Veraguas) - LT
= <i>cyclops</i> (Staudinger, 1876) (<i>Pyrrhopyge</i> [sic])	Panama (Chiriquí) - LT

b) <i>creon lilliana</i> (Nicolay & Small, 1969) (<i>Pyrrhopyge</i>)	Panama (Panamá) - HT
c) <i>creon taylori</i> (Nicolay & Small, 1981) (<i>Pyrrhopyge</i>)	Costa Rica (Puntarenas) - HT
<i>Cyanopyge</i> O. Mielke, 2002, stat. rev.	
<i>sangaris</i> (Skinner, 1921) (<i>Pyrrhopyge</i>), comb. rest.	Colombia (Magdalena) - HT
<i>Jonaspyge</i> O. Mielke, 2002	
<i>elizabethae</i> Gallardo & Grishin, 2021	Honduras - HT
<i>jonas</i> (C. Felder & R. Felder, 1859) (<i>Pyrrhopyga</i> [sic])	Mexico - LT
= <i>cydonia</i> (H. Druce, 1874) (<i>Pyrrhopyga</i> [sic])	Mexico (Oaxaca) - LT
<i>tzotzili</i> (Freeman, 1969) (<i>Pyrrhopyge</i>)	Mexico (Chiapas) - HT
Microcerisina Grishin, 2019	
<i>Apatiella</i> Grishin, 2019	
<i>iphinous</i> (Latreille, [1824]) (<i>Hesperia</i>)	
a) <i>iphinous iphinous</i> (Latreille, [1824]) (<i>Hesperia</i>)	Brazil - LT
= <i>othello</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Brazil (Rio de Janeiro) - NT
= <i>mimetes</i> (Mabille, 1909) (<i>Mimoniades</i>)	Brazil (Rio de Janeiro) - NT
= <i>othello</i> var. <i>negrita</i> (J. Zikán, 1938) (<i>Mimoniades</i>)	Brazil (Rio de Janeiro) - LT
b) <i>iphinous mariae</i> (Bell, 1931) (<i>Pyrrhopyge</i>), comb. n., stat. n.	Brazil (Rio Grande do Sul) - HT
c) <i>iphinous molinai</i> (Hayward, [1942]) (<i>Pyrrhopyge</i>), comb. n.	Argentina (Misiones) - HT
<i>Blubella</i> Grishin, 2019	
<i>adonis</i> (Bell, 1931) (<i>Pyrrhopyge</i>)	Paraguay (Guairá) - HT
<i>azeta</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	
a) <i>azeta azeta</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
b) <i>azeta giffordi</i> (O. Mielke, 1995) (<i>Elbella</i>)	Brazil (Distrito Federal) - HT
<i>bicuspis</i> (de Jong, 1983) (<i>Elbella</i>)	Suriname (Paramaribo) - HT
<i>blanda</i> (Evans, 1951) (<i>Elbella</i>)	Peru (Loreto) - HT
<i>etna</i> (Evans, 1951) (<i>Elbella</i>)	Brazil (Amazonas) - HT
= <i>etna moda</i> (Evans, 1951) (<i>Elbella</i>)	Peru (Junín) - HT
<i>lustra</i> (Evans, 1951) (<i>Elbella</i>)	Colombia (Cundinamarca) - HT
<i>madeira</i> (O. Mielke, 1995) (<i>Elbella</i>)	Brazil (Mato Grosso) - HT
<i>miodesmiata</i> (Röber, 1925) (<i>Jemadia</i>)	Colombia - NT
<i>patrobas</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic])	
a) <i>patrobas patrobas</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic])	Colombia - LT
b) <i>patrobas amazonica</i> (O. Mielke, 1995) (<i>Elbella</i>)	Brazil (Amazonas) - HT
c) <i>patrobas evansi</i> (O. Mielke, 1995) (<i>Elbella</i>)	Colombia (Valle del Cauca) - HT
d) <i>patrobas melanina</i> (Mabille & Boulet, 1908) (<i>Jemadia</i>)	Peru - LT
e) <i>patrobas mexicana</i> (O. Mielke, 1995) (<i>Elbella</i>)	Mexico (Chiapas) - HT
f) <i>patrobas tingo</i> (O. Mielke, 1995) (<i>Elbella</i>)	Brazil (Rondônia) - HT
<i>patroclus</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	
a) <i>patroclus patroclus</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Peru - HT
= <i>umbrata</i> (Mabille & Boulet, 1908) (<i>Jemadia</i>)	Bolivia (La Paz) - NT
b) <i>patroclus acala</i> (Evans, 1951) (<i>Elbella</i>)	Colombia (Cesar) - HT
<i>rondonia</i> (O. Mielke, 1995) (<i>Elbella</i>)	Brazil (Mato Grosso) - HT
<i>Croniades</i> Mabille, 1903	
<i>auraria</i> H. H. Druce, 1908, stat. rest.	Bolivia (La Paz) - LT
<i>pieria</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas River) - LT
= <i>zesta</i> (Evans, 1951) (<i>Elbella</i>)	French Guiana - HT
<i>Elbella</i> Evans, 1951, stat. rev.	
<i>dulcinea</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), comb. rev.	[Colombia] - LT
<i>intersecta</i> (Herrich-Schäffer, 1869) (<i>Myscelus</i>), comb. rev.	
a) <i>intersecta intersecta</i> (Herrich-Schäffer, 1869) (<i>Myscelus</i>), comb. rev.	no locality - LT
= <i>fluminis</i> (Butler, 1873) (<i>Pyrrhopyga</i> [sic])	Brazil (Pará) - LT
= <i>semidentata</i> (Mabille, 1877) (<i>Pyrrhopyga</i> [sic])	Peru - LT
= <i>tiribazus</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), nom. nud. (ICZN, Art. 11.6)	Brazil (Pará)
b) <i>intersecta losca</i> Evans, 1951, comb. rev.	Brazil (Mato Grosso) - HT
= <i>intersecta ilona</i> Evans, 1951, syn. n.	Brazil (Goiás) - HT

c) <i>intersecta paraensis</i> O. Mielke, 1995, comb. rev.	Brazil (Pará) - HT
d) <i>intersecta peter</i> Evans, 1951, comb. rev.	Brazil (Pará) - HT
e) <i>intersecta rufitegula</i> O. Mielke, 1995, comb. rev.	Brazil (Minas Gerais) - HT
f) <i>intersecta strova</i> Evans, 1951, comb. rev.	Venezuela (Bolívar) - HT
<i>lamprus</i> (Hopffer, 1874) (<i>Pyrrhopyge</i>), comb. rev.	
a) <i>lamprus lamprus</i> (Hopffer, 1874) (<i>Pyrrhopyge</i>), comb. rev.	Brazil (Rio de Janeiro) - LT
= <i>menecrates</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	“Peru” - LT
= <i>margimmiscus</i> (Hayward, 1935) (<i>Pyrrhopyge</i>)	Argentina (Misiones) - HT
b) <i>lamprus albociliata</i> O. Mielke, 1995, comb. rev.	Brazil (Santa Catarina) - HT
<i>scylla</i> (Ménétriés, 1855) (<i>Pyrrhopyga</i> [sic]), comb. rev.	Nicaragua - LT
<i>theseus</i> (Bell, 1934) (<i>Jemadia</i>), comb. rev.	Brazil - HT
<i>viriditas</i> (Skinner, 1920) (<i>Pyrrhopyge</i>), comb. rev.	
a) <i>viriditas viriditas</i> (Skinner, 1920) (<i>Pyrrhopyge</i>), comb. rev.	“Honduras (Cortés)” - HT
= <i>carriæ</i> (Bell, 1931) (<i>Pyrrhopyge</i>)	Paraguay (Paraguari) - HT
b) <i>viriditas boliviana</i> O. Mielke, 1995, comb. rev.	Bolivia (La Paz) - HT
c) <i>viriditas chia</i> Evans, 1951, comb. rev.	Brazil (Amazonas) - HT
<i>Hegesippe</i> Evans, 1951	
<i>hegesippe</i> (Mabille & Boulet, 1908) (<i>Sarbia</i>)	South America - HT
<i>luteizona</i> (Mabille, 1877) (<i>Pyrrhopyga</i> [sic])	“Mexico” - LT
= <i>martii</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Brazil - HT
= <i>josepha</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Brazil - HT
= <i>amoena</i> (Röber, 1925) (<i>Sarbia</i>)	Brazil - NT
= <i>xanthomargo</i> (Hayward, 1942) (<i>Pyrrhopyge</i>)	Brazil (São Paulo) - NT
<i>Merobella</i> Grishin, 2019	
<i>merops</i> (Bell, 1934) (<i>Jemadia</i>)	Colombia - HT
<i>Microceris</i> Watson, 1893	
<i>varicolor</i> (Ménétriés, 1855) (<i>Pyrrhopyga</i> [sic])	Brazil (Minas Gerais) - LT
<i>Ochropyge</i> O. Mielke, 2002	
<i>ruficauda</i> (Hayward, 1932) (<i>Pyrrhopyge</i>)	Argentina (Misiones) - HT
= <i>excelsus</i> (Bell, 1947) (<i>Pyrrhopyge</i>)	Brazil (Paraná) - HT
<i>Parelbella</i> O. Mielke, 1995	
<i>ahira</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	
a) <i>ahira ahira</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Pará) - LT
= <i>jamina</i> (Butler, 1870) (<i>Pyrrhopyga</i> [sic])	no locality - LT
= <i>zimra</i> (Hewitson, 1871) (<i>Pyrrhopyga</i> [sic])	no locality - LT
b) <i>ahira extrema</i> (Röber, 1925) (<i>Jemadia</i>)	Brazil (São Paulo) - NT
<i>macleanani</i> (Godman & Salvin, 1893) (<i>Jemadia</i>)	Panama (Veraguas) - LT
<i>nigra</i> O. Mielke, Austin & Warren, 2008	Mexico (Oaxaca) - HT
<i>peruana</i> O. Mielke, 1995	Peru (Junín) - HT
<i>polyzona</i> (Latreille, [1824]) (<i>Hesperia</i>)	Brazil - HT
<i>Protellabella</i> O. Mielke, 1995	
<i>alburna</i> (Mabille, 1891) (<i>Pyrrhopyga</i> [sic])	
a) <i>alburna alburna</i> (Mabille, 1891) (<i>Pyrrhopyga</i> [sic])	Peru (Junín) - LT
b) <i>alburna brasiliensis</i> O. Mielke, 1995	Brazil (Rondônia) - HT
<i>Pseudocroniades</i> O. Mielke, 1995	
<i>machaon</i> (Westwood, 1852) (<i>Ericides</i> [sic])	
a) <i>machaon machaon</i> (Westwood, 1852) (<i>Ericides</i> [sic])	Brazil - LT
b) <i>machaon seabrai</i> O. Mielke, 1995	Brazil (Rio de Janeiro) - HT
Mimoniadina Grishin, 2019	
<i>Aerardaris</i> O. Mielke, Brockmann & C. Mielke, gen. n.	
<i>aerata</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic]), comb. n.	Colombia (Magdalena) - HT
<i>Amenis</i> Watson, 1893	
<i>picia</i> Evans, 1951, stat. n.	Colombia (Cundinamarca) - HT
<i>pionia</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic])	
a) <i>pionia pionia</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic])	Colombia - LT

	= <i>similis</i> Röber, 1925	Venezuela - LT
	b) <i>pionia pedro</i> O. Mielke & Casagrande, ssp. n.	Brazil (Bahia) - HT
	c) <i>pionia sandra</i> Orellana, [2010]	Venezuela (Táchira) - HT
	<i>ponina</i> (Herrich-Schäffer, 1869) (<i>Pyrrhopyga</i> [sic])	Venezuela (Aragua) – NT
	<i>rogeri</i> Orellana, [2010]	Venezuela (Mérida) - HT
<i>Amysoria</i> O. Mielke, 2002		
	<i>galgala</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Venezuela - LT
	= <i>strigifera</i> (C. Felder & R. Felder, 1867) (<i>Tamyris</i>)	Colombia (Cundinamarca) - LT
<i>Ardaris</i> Watson, 1893		
	<i>eximia</i> (Hewitson, 1871) (<i>Pyrrhopyga</i> [sic])	Venezuela - LT
	<i>hantra</i> Evans, 1951	Venezuela (Mérida) - HT
<i>Jemadia</i> Watson, 1893		
	<i>demarmelsi</i> Orellana, [2010]	Venezuela (Bolívar) – HT
	<i>hospita</i> (Butler, 1877) (<i>Pyrrhopyga</i> [sic])	
	a) <i>hospita hospita</i> (Butler, 1877) (<i>Pyrrhopyga</i> [sic])	Peru - LT
	= <i>aethiops</i> (Gmelin, [1790]) (<i>Papilio</i>), praeocc. (Esper, 1877)	[Suriname?] - LT
	= <i>ulixes</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Suriname - LT
	= <i>hephaestos</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), syn. n.	Suriname - LT
	= <i>zamorae</i> Mabille & Boulet, 1908, nom. nud. (ICZN, Art. 11.6)	Suriname
	b) <i>hospita imitator</i> (Mabille, 1891) (<i>Pyrrhopyga</i> [sic])	Colombia (Cundinamarca) - LT
	= <i>paulensis</i> Schaus, 1902, syn. n.	Brazil (Amazonas) - LT
	<i>mabillei</i> O. Mielke, Brockmann & C. Mielke, sp. n.	no locality - HT
	<i>pseudognetus</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	Colombia - LT
	= <i>dorylas</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Venezuela (Carabobo) - HT
<i>Jemasonia</i> Grishin, 2019		
	<i>albescens</i> (Röber, 1925) (<i>Jemadia</i>), stat. rest.	Ecuador (Napo) – NT
	<i>hewitsonii</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	
	a) <i>hewitsonii hewitsonii</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	Brazil ([Amazonas]) - LT
	= <i>le cerfi</i> [sic] (Mabille & Boulet, 1908) (<i>Jemadia</i>)	French Guiana (Cayenne) - LT
	b) <i>hewitsonii sosia</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic]), comb. n., stat. n.	Brazil - HT
	= <i>lisetta</i> (Mabille & Boulet, 1908) (<i>Jemadia</i>), syn. n.	Peru – LT
	<i>ortizi</i> (Orellana, [2010]) (<i>Jemadia</i>)	Venezuela (Táchira) - HT
	<i>ovid</i> (Evans, 1951) (<i>Jemadia</i>)	Ecuador (Imbabura) - HT
	<i>pater</i> (Evans, 1951) (<i>Jemadia</i>)	Colombia (Cundinamarca) - HT
	<i>suekentonmiller</i> (Grishin, 2014) (<i>Jemadia</i>)	Costa Rica (Guanacaste) - HT
<i>Jematus</i> Grishin, 2019		
	<i>gnetus</i> (Fabricius, 1781) (<i>Papilio</i>)	
	a) <i>gnetus gnetus</i> (Fabricius, 1781) (<i>Papilio</i>)	“India” - LT
	= <i>vulcanus</i> (Cramer, 1779) (<i>Papilio</i>), praeocc. (Fabricius, 1775)	Suriname - LT
	= <i>megalesius</i> (Hübner, [1819]) (<i>Erycides</i>), nom. n.	Suriname
	= <i>suzetta</i> (Mabille & Boulet, 1908) (<i>Jemadia</i>)	Bolivia - HT
	b) <i>gnetus brevipennis</i> (Schaus, 1902) (<i>Jemadia</i>)	Brazil (São Paulo) - LT
	c) <i>gnetus mineira</i> O. Mielke, C. Mielke & Casagrande, ssp. n.	Brazil (Minas Gerais) - HT
<i>Jember</i> Grishin, 2019		
	<i>menechmus</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	
	a) <i>menechmus menechmus</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	no locality - LT
	b) <i>menechmus desousai</i> (Orellana, [2010]) (<i>Jemadia</i>)	Venezuela (Amazonas) - HT
	<i>scomber</i> (H. H. Druce, 1908) (<i>Jemadia</i>)	Peru (Pasco) - LT
<i>Mahotis</i> Watson, 1893		
	<i>baroni</i> (Godman & Salvin, 1895) (<i>Amenis</i>)	Peru (Cajamarca) - LT
	<i>eupheme</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic]), stat. rest.	Peru (Cusco) - LT
	<i>montana</i> (Zikán, 1938) (<i>Mimoniades</i>)	Brazil (Espírito Santo) - LT
	<i>nurscia</i> (Swainson, 1821) (<i>Tamyris</i>)	
	a) <i>nurscia nurscia</i> (Swainson, 1821) (<i>Tamyris</i>)	Peru (Huánuco) - NT
	b) <i>nurscia amans</i> (Skinner, 1920) (<i>Mimoniades</i>)	Colombia (Huila) - HT
	c) <i>nurscia malis</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic])	Colombia (Antioquia) - LT

<i>versicolor</i> (Latreille, [1824]) (<i>Hesperia</i>)	Brazil - LT
= <i>mulcifer</i> (Hübner, [1831]) (<i>Mimoniades</i>)	Brazil (Minas Gerais) - NT
<i>Mimadia</i> Grishin, 2019	
<i>fallax</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	
a) <i>fallax fallax</i> (Mabille, 1878) (<i>Pyrrhopyga</i> [sic])	South America - LT
b) <i>fallax fida</i> (Evans, 1951) (<i>Jemadia</i>)	Peru (Puno) - HT
c) <i>fallax fiska</i> (Evans, 1951) (<i>Jemadia</i>)	Colombia (Cundinamarca) - HT
d) <i>fallax solaris</i> (Hayward, 1942) (<i>Jemadia</i>)	Brazil (Amazonas) - NT
= <i>fallax filta</i> (Evans, 1951) (<i>Jemadia</i>)	Peru (Loreto) - HT
<i>Mimardaris</i> O. Mielke, 2002, stat. rev.	
<i>lomax</i> (Evans, 1951) (<i>Mimoniades</i>), comb. rev.	Peru (Cusco) - HT
<i>minthe</i> (Godman & Salvin, 1879) (<i>Pyrrhopyga</i> [sic]), comb. rev.	Ecuador (Tungurahua) - LT
= <i>angustifascia</i> (Röber, 1925) (<i>Mimoniades</i>)	Ecuador (Morona-Santiago) - HT
<i>montra</i> (Evans, 1951) (<i>Mimoniades</i>), comb. rev.	Peru (Pasco) - HT
<i>periphema</i> (Hewitson, [1875]) (<i>Pyrrhopyga</i> [sic]), stat. rest., comb. rev.	Bolivia - LT
= <i>inaequalis</i> (Röber, 1925) (<i>Mimoniades</i>)	Bolivia - LT
<i>pityusa</i> (Hewitson, 1857) (<i>Pyrrhopyga</i> [sic]), comb. rev.	[Ecuador] - LT
= <i>aemulus</i> (Skinner, 1920) (<i>Mimoniades</i>)	Ecuador (Tungurahua) - HT
= <i>hemitaenia</i> (Röber, 1925) (<i>Mimoniades</i>)	Colombia (Magdalena) - LT
<i>porus</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), comb. rev.	
a) <i>porus porus</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic]), comb. rev.	Colombia - LT
= <i>punctiger</i> (Mabille & Boullet, 1908) (<i>Mimoniades</i>)	Peru - LT
= <i>minthe</i> var. <i>egena</i> (Mabille & Boullet, 1908) (<i>Mimoniades</i>)	Peru
b) <i>porus mortis</i> (Evans, 1951) (<i>Mimoniades</i>), comb. rev.	Peru (Amazonas) - HT
<i>sela</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic]), comb. rev.	
a) <i>sela sela</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic]), comb. rev.	Colombia - LT
b) <i>sela aequatorea</i> (Röber, 1925) (<i>Mimoniades</i>), comb. rev.	Ecuador (Morona-Santiago) - HT
c) <i>sela peruviana</i> (Draudt, 1921) (<i>Mimoniades</i>), comb. rev.	Peru - LT
= <i>pityusa</i> ab. <i>chanchamayonis</i> (Strand, 1921) (<i>Mimoniades</i>), nom. nud. (ICZN, Art. 45.6.2) Peru (Junín)	
= <i>pityusa</i> form <i>chanchamayonis</i> (Draudt, 1924) (<i>Mimoniades</i>), syn. n.	Peru (Junín) - HT
<i>Mimoniades</i> Hübner, 1823	
<i>ocyalus</i> Hübner, 1823	Brazil (Minas Gerais) - NT
<i>Mysarbia</i> O. Mielke, 2002	
<i>sejanus</i> (Hopffer, 1874) (<i>Pyrrhopyge</i>)	
a) <i>sejanus sejanus</i> (Hopffer, 1874) (<i>Pyrrhopyge</i>)	Peru - LT
b) <i>sejanus erythrostigma</i> (Röber, 1925) (<i>Mysoria</i>)	Bolivia (La Paz) - LT
c) <i>sejanus stollii</i> O. Mielke, 2002	Suriname - LT
= <i>thasus</i> (Stoll, 1782) (<i>Papilio</i>), praeocc. (Stoll, 1780)	Suriname - LT
<i>Mysoria</i> Watson, 1893	
<i>affinis</i> (Herrich-Schäffer, 1869) (<i>Pyrrhopyga</i> [sic])	Tropical America - LT
= <i>wilsoni</i> Freeman, 1969	Mexico (Guerrero) - HT
<i>ambigua</i> (Mabille & Boullet, 1908) (<i>Amenis</i>)	[Honduras (Cortés)] - HT
= <i>proxima</i> (Mabille & Boullet, 1908) (<i>Amenis</i>)	Mexico (Jalisco) - LT
<i>amra</i> (Hewitson, 1871) (<i>Pyrrhopyga</i> [sic])	Mexico - LT
= <i>brasiliensis</i> (Mabille, 1903) (<i>Amenis</i>)	Mexico (Oaxaca) - NT
<i>barcastus</i> (Sepp, [1851]) (<i>Papilio</i>)	
a) <i>barcastus barcastus</i> (Sepp, [1851]) (<i>Papilio</i>)	Suriname (Brokopondo) - NT
= <i>verbena</i> (Butler, 1869) (<i>Pyrrhopyga</i> [sic])	South America - LT
b) <i>barcastus alta</i> Evans, 1951	Trinidad and Tobago (Port of Spain) - HT
c) <i>barcastus antila</i> Evans, 1951	Brazil (Pará) - HT
d) <i>barcastus barta</i> Evans, 1951	Paraguay - HT
e) <i>barcastus pallens</i> (Mabille, 1891) (<i>Pyrrhopyga</i> [sic])	[Colombia] - LT
f) <i>barcastus venezuelae</i> (Scudder, 1872) (<i>Pyrrhopyga</i> [sic])	Venezuela - LT
= <i>acastus</i> (Cramer, 1775) (<i>Papilio</i>), praeocc. (Linnaeus, 1758)	Suriname (Paramaribo) - NT
= <i>decolor</i> Mabille & Boullet, 1908	[Colombia (Cundinamarca)] - LT
= <i>cayennae</i> Mabille & Boullet, 1908	French Guiana (Saint-Laurent-du-Maroni) - LT

= <i>caucana</i> Röber, 1925	Colombia ([Valle del] Cauca) - NT
<i>Nosphistia</i> Mabilie & Boulet, 1908	
<i>zonara</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
= <i>perplexus</i> (Mabilie, 1878) (<i>Pyrrhopyga</i> [sic])	Brazil - LT
= <i>gigantea</i> (Röber, 1925) (<i>Jemadia</i>)	no locality - HT
<i>Santea</i> Grishin, 2019	
<i>antias</i> (C. Felder & R. Felder, 1859) (<i>Pyrrhopyga</i> [sic])	South Brazil - LT
<i>Sarbia</i> Watson, 1893	
= <i>Metardaris</i> Mabilie, 1903	
<i>cosinga</i> (Hewitson, 1874) (<i>Pyrrhopyga</i> [sic])	
a) <i>cosinga cosinga</i> (Hewitson, 1874) (<i>Pyrrhopyga</i> [sic])	Bolivia - LT
= <i>coninga</i> [sic] var. <i>obscura</i> (Mabilie, 1903) (<i>Metardaris</i>)	Bolivia - LT
= <i>coninga</i> [sic] var. <i>sanguinea</i> (Mabilie, 1903) (<i>Metardaris</i>)	Bolivia - LT
= <i>stygiata</i> (Mabilie & Boulet, 1908) (<i>Metardaris</i>), nom. nud. (ICZN, Art. 11.6)	[Bolivia]
= <i>territa</i> (Mabilie & Boulet, 1908) (<i>Metardaris</i>), nom. nud. (ICZN, Art. 11.6)	[Bolivia]
b) <i>cosinga catana</i> (Evans, 1951) (<i>Metardaris</i>)	Peru (Puno) - HT
c) <i>cosinga cedra</i> (Evans, 1951) (<i>Metardaris</i>)	Bolivia (Cochabamba) - HT
d) <i>cosinga circe</i> (Evans, 1951) (<i>Metardaris</i>)	Bolivia (La Paz) - HT
<i>curitiba</i> O. Mielke & Casagrande, 2002	Brazil (Paraná) - HT
<i>damippe</i> Mabilie & Boulet, 1908	South Brazil - LT
<i>oneka</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	“Venezuela” - LT
<i>pertyi</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Brazil (Santa Catarina) - NT
<i>soza</i> Evans, 1951	Brazil (Paraná) - HT
<i>xanthippe</i> (Latreille, [1824]) (<i>Hesperia</i>)	
a) <i>xanthippe xanthippe</i> (Latreille, [1824]) (<i>Hesperia</i>)	Brazil - LT
b) <i>xanthippe spixii</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Brazil (Santa Catarina) - LT
<i>Sarbieta</i> Grishin, 2019	
<i>catomelaena</i> (Mabilie & Boulet, 1908) (<i>Sarbia</i>)	
a) <i>catomelaena catomelaena</i> (Mabilie & Boulet, 1908) (<i>Sarbia</i>)	Brazil (Minas Gerais) - LT
= <i>mello-leitaoi</i> [sic] (Hayward, 1942) (<i>Sarbia</i>)	Brazil (Minas Gerais) – NT
b) <i>catomelaena pico</i> O. Mielke & Casagrande, ssp. n.	Brazil (Bahia) - HT
Pyrrhopygina (Mabilie, 1877) Warren, Ogawa & Brower, 2008	
<i>Gunayan</i> O. Mielke, 2002	
<i>rhacia</i> (Hewitson, [1875]) (<i>Pyrrhopyga</i> [sic])	Brazil (Minas Gerais) - LT
<i>rubricollis</i> (Sepp, [1841]) (<i>Papilio</i>)	
a) <i>rubricollis rubricollis</i> (Sepp, [1841]) (<i>Papilio</i>)	Suriname - NT
= <i>hadora</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
b) <i>rubricollis flemingi</i> (Bell, 1947) (<i>Yanguna</i>), stat. rev., stat. n.	Venezuela (Monagas) - HT
<i>timaeus</i> (Bell, 1931) (<i>Yanguna</i>)	Peru (Loreto) - HT
<i>Pyrrhopyge</i> Hübner, [1819]	
= <i>Tamyris</i> Swainson, 1821	
= <i>Pachyrhopala</i> Wallengren, 1858	
<i>afluente</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (San Martín) - HT
<i>amyclas</i> (Cramer, 1779) (<i>Papilio</i>)	
a) <i>amyclas amyclas</i> (Cramer, 1779) (<i>Papilio</i>)	Suriname (Paramaribo) - NT
= <i>amiatus</i> (Fabricius, 1793) (<i>Hesperia</i>)	America - LT
= <i>laonome</i> (Swainson, 1821) (<i>Tamyris</i>)	Suriname (Paramaribo) - NT
b) <i>amyclas denticulata</i> Herrich-Schäffer, 1869 (<i>Pyrrhopyga</i> [sic])	Brazil (Roraima) - NT
<i>amythaon</i> Bell, 1931	
a) <i>amythaon amythaon</i> Bell, 1931	Brazil (Pernambuco) - HT
b) <i>amythaon gradens</i> Evans, 1951	Brazil (Pará) - HT
c) <i>amythaon peron</i> Bell, 1947	Peru (Loreto) - HT
d) <i>amythaon podina</i> Evans, 1951	Bolivia - HT
<i>arax</i> Evans, 1951	Bolivia - HT
<i>arinas</i> (Cramer, 1777) (<i>Papilio</i>)	

a) <i>arinas arinas</i> (Cramer, 1777) (<i>Papilio</i>)	Suriname - LT
= <i>pedaia</i> Hewitson, 1866 (<i>Pyrrhopyga</i> [<i>sic</i>])	Brazil (Amazonas) - LT
b) <i>arinas erica</i> O. Mielke, Brockmann & C. Mielke, <i>ssp. n.</i>	Peru (Junín) - HT
c) <i>arinas temenos</i> (Bell, 1931) (<i>Yanguna</i>)	Peru (Junín) - HT
<i>attis</i> Bell, 1931, <i>stat. rest.</i>	
a) <i>attis attis</i> Bell, 1931, <i>stat. rest.</i>	Bolivia (Santa Cruz) - HT
= <i>denieri</i> Hayward, 1938	Bolivia (La Paz) - HT
b) <i>attis arbor</i> Evans, 1951, <i>comb. n.</i>	Venezuela - HT
c) <i>attis cardus</i> Mabilille, 1891 (<i>Pyrrhopyga</i> [<i>sic</i>]), <i>comb. n.</i>	Brazil ([Pará]) - LT
d) <i>attis lexis</i> Evans, 1951, <i>comb. n.</i>	Guyana (Barima-Waini) - HT
e) <i>attis miersi</i> O. Mielke, Brockmann & C. Mielke, <i>ssp. n.</i>	Brazil (Santa Catarina) - HT
f) <i>attis subnubilus</i> Hayward, 1935, <i>comb. n.</i>	Argentina (Misiones) - HT
<i>aziza</i> Hewitson, 1866 (<i>Pyrrhopyga</i> [<i>sic</i>])	
a) <i>aziza aziza</i> Hewitson, 1866 (<i>Pyrrhopyga</i> [<i>sic</i>])	Colombia - LT
b) <i>aziza araethyrea</i> Hewitson, 1870 (<i>Pyrrhopyga</i> [<i>sic</i>])	Ecuador (Tungurahua) - LT
c) <i>aziza troja</i> Evans, 1951	Peru (Junín) - HT
<i>boulleti</i> Le Cerf, 1922	Colombia (Norte de Santander) - HT
<i>cacaulandia</i> O. Mielke, Brockmann & C. Mielke, <i>sp. n.</i>	Brazil (Rondônia) - HT
<i>caranavi</i> O. Mielke, Brockmann & C. Mielke, <i>sp. n.</i>	Bolivia (La Paz) - HT
<i>caribe</i> Orellana, [2010]	
a) <i>caribe caribe</i> Orellana, [2010]	Venezuela (Aragua) - HT
b) <i>caribe camachoi</i> Orellana, [2010]	Venezuela (Bolívar) - HT
<i>charybdis</i> Westwood, 1852	
a) <i>charybdis charybdis</i> Westwood, 1852 (<i>Pyrrhopyga</i> [<i>sic</i>])	[Brazil (Minas Gerais)] - LT
b) <i>charybdis semita</i> Evans, 1951	Brazil (Rio de Janeiro) - HT
<i>cosnipata</i> O. Mielke, Brockmann & C. Mielke, <i>sp. n.</i>	Peru (Cusco) - HT
<i>creona</i> H. Druce, 1874 (<i>Pyrrhopyga</i> [<i>sic</i>])	Peru - LT
= <i>aurora</i> Mabilille, 1891 (<i>Pyrrhopyga</i> [<i>sic</i>])	"Brazil" - LT
<i>cressoni</i> Bell, 1932	Bolivia (Santa Cruz) - HT
<i>creusae</i> (Bell, 1931) (<i>Yanguna</i>)	French Guiana (Cayenne) - HT
<i>crida</i> Hewitson, 1871 (<i>Pyrrhopyga</i> [<i>sic</i>])	Nicaragua (Chontales) - LT
<i>crista</i> Evans, 1951	Bolivia (La Paz) - HT
<i>croceimargo</i> Mabilille & Boullet, 1908, <i>stat. rest.</i>	Bolivia - LT
<i>decipiens</i> Mabilille, 1903	Bolivia - LT
<i>divisor</i> O. Mielke & Casagrande, <i>sp. n.</i>	Brazil (Acre) - HT
<i>draudti</i> Bell, 1931, <i>stat. rest.</i>	
a) <i>draudti draudti</i> Bell, 1931, <i>stat. rest.</i>	Bolivia (Santa Cruz) - HT
b) <i>draudti bruno</i> O. Mielke, Brockmann & C. Mielke, <i>ssp. n.</i>	Colombia (Meta) - HT
c) <i>draudti cintra</i> Evans, 1951, <i>comb. n.</i>	Brazil (Amazonas) - HT
d) <i>draudti lina</i> Bell, 1947, <i>comb. rest.</i>	Brazil (Pará) - HT
<i>eduardo</i> O. Mielke, Brockmann & C. Mielke, <i>sp. n.</i>	Bolivia (La Paz) - HT
<i>erazoe</i> Orellana, [2010]	Venezuela (Bolívar) - HT
<i>evansi</i> Bell, 1947	
a) <i>evansi evansi</i> Bell, 1947	Colombia (Boyacá) - HT
b) <i>evansi borburata</i> Orellana, [2010]	Venezuela (Carabobo) - HT
<i>frona</i> Evans, 1951	Peru (Junín) - HT
<i>guianae</i> Bell, 1932	French Guiana - HT
<i>hadassa</i> Hewitson, 1866 (<i>Pyrrhopyga</i> [<i>sic</i>])	[Ecuador] - LT
= <i>rubricor</i> Mabilille & Boullet, 1908	Ecuador (Loja) - LT
<i>haemon</i> Godman & Salvin, 1893	Costa Rica - HT
<i>halma</i> Evans, 1951, <i>stat. n.</i>	Bolivia (La Paz) - HT
<i>hanga</i> Evans, 1951, <i>stat. n.</i>	Peru (Pasco) - HT
<i>infantilis</i> H. H. Druce, 1908	
a) <i>infantilis infantilis</i> H. H. Druce, 1908	Peru - LT
b) <i>infantilis agala</i> Evans, 1951	Bolivia (Cochabamba) - HT
<i>iquitos</i> O. Mielke, Brockmann & C. Mielke, <i>sp. n.</i>	Peru (Loreto) - HT

<i>jola</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (Cusco) - HT
<i>josephina</i> Draudt, 1920, stat. rest.	
a) <i>josephina josephina</i> Draudt, 1920, stat. rest.	Bolivia (La Paz) - HT
= <i>amythaon perula</i> Evans, 1951, syn. n.	Peru (Junín) - HT
b) <i>josephina orino</i> Evans, 1951, comb. n.	Colombia (Vichada) - HT
c) <i>josephina polka</i> Evans, 1951, comb. n.	French Guiana (Saint-Laurent-du-Maroni) - HT
d) <i>josephina pollio</i> Evans, 1951, comb. n.	Peru (Puno) - HT
<i>kelita</i> Hewitson, [1869] (<i>Pyrrhopyga</i> [sic])	Bolivia (La Paz) - LT
= <i>kelita</i> var. <i>tristis</i> Mabilille & Bouillet, 1908	Bolivia (La Paz) - LT
<i>leticia</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (Junín) - HT
<i>martena</i> Hewitson, [1869] (<i>Pyrrhopyga</i> [sic])	Ecuador - LT
<i>melanomerus</i> Mabilille & Bouillet, 1908	
a) <i>melanomerus melanomerus</i> Mabilille & Bouillet, 1908	Peru (Junín) - NT
= <i>tarapotoensis</i> Bell, 1931	Peru (San Martín) - HT
b) <i>melanomerus patma</i> Evans, 1951	Ecuador (Zamora-Chinchipe) - HT
<i>mendax</i> Mabilille & Bouillet, 1908, stat. rest.	Peru - HT
= <i>telassina tagra</i> Evans, 1951, syn. n.	Peru (Amazonas) - HT
<i>michael</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (Huánuco) - HT
<i>mina</i> Bell, 1931, stat. rest.	
a) <i>mina mina</i> Bell, 1931, stat. rest.	Bolivia - HT
= <i>sergius bolius</i> Bell, 1947	Bolivia (La Paz) - HT
b) <i>mina ganus</i> Bell, 1947, comb. n.	Guyana - HT
c) <i>mina previsto</i> O. Mielke, Brockmann & C. Mielke, ssp. n.	Peru (Ucayali) - HT
<i>mopsus</i> (Bell, 1931) (<i>Yanguna</i>)	
a) <i>mopsus mopsus</i> (Bell, 1931) (<i>Yanguna</i>)	Peru (Junín) - HT
b) <i>mopsus elisa</i> O. Mielke, Brockmann & C. Mielke, ssp. n.	Peru (Madre de Dios) - HT
<i>olivacea</i> Röber, 1925, stat. rev.	Peru - LT
= <i>hadassa henna</i> Evans, 1951, syn. n.	Peru (Amazonas) - HT
<i>papius</i> Hopffer, 1874	
a) <i>papius papius</i> Hopffer, 1874	Bolivia (Beni) - LT
b) <i>papius pasca</i> Evans, 1951	Colombia (Risaralda) - HT
<i>pelota</i> Plötz, 1879 (<i>Pyrrhopyga</i> [sic])	Brazil - LT
<i>placeta</i> Evans, 1951	Brazil (Espírito Santo) - HT
<i>phaeae</i> Hopffer, 1874, stat. rest.	Peru (Junín) - LT
= <i>telassa</i> var. <i>tenuis</i> Staudinger, 1888	Peru (Junín) - LT
<i>phidias</i> (Linnaeus, 1758) (<i>Papilio</i>)	
a) <i>phidias phidias</i> (Linnaeus, 1758) (<i>Papilio</i>)	Colombia (Meta) - NT
= <i>zeleucus</i> (Fabricius, 1793) (<i>Hesperia</i>)	Colombia (Meta) - NT
= <i>williamsi</i> Bell, 1931	Peru - HT
b) <i>phidias aldo</i> O. Mielke, Brockmann & C. Mielke, ssp. n.	Brazil (Pernambuco) - HT
c) <i>phidias bixae</i> (Linnaeus, 1758) (<i>Papilio</i>)	Suriname - LT
= <i>maenas</i> (Fabricius, 1787) (<i>Papilio</i>)	America - LT
= <i>meriani</i> (Kirby, 1871) (<i>Telegonus</i>), nom. n.	Suriname - LT
d) <i>phidias garata</i> Hewitson, 1866 (<i>Pyrrhopyga</i> [sic])	Suriname - LT
e) <i>phidias hyperici</i> Hübner, 1823	Brazil (Pará) - NT
= <i>hyperici</i> Hübner, [1819], nom. nud. (ICZN, Art. 11.1)	Brazil (Pará)
= <i>hyleus</i> Mabilille, 1891 (<i>Pyrrhopyga</i> [sic]), syn. n.	Brazil (Pará) - LT
f) <i>phidias latifasciata</i> Butler, 1873	Colombia (Cundinamarca) - LT
g) <i>phidias leucoloma</i> Erschoff, 1875	Peru (Cusco) - NT
h) <i>phidias rusca</i> Evans, 1951	Peru (Amazonas) - HT
<i>phylleia</i> Hewitson, 1874 (<i>Pyrrhopyga</i> [sic])	Bolivia - LT
= <i>hades</i> Mabilille, 1903	Bolivia - LT
= <i>fleximargo</i> Mabilille & Bouillet, 1908	Peru - HT
= <i>punctata</i> Röber, 1925, syn. n.	Bolivia (La Paz) - LT
= <i>phylleia delos</i> Evans, 1951, syn. n.	Peru (Puno) - HT
<i>proculus</i> Hopffer, 1874	

a) <i>proculus proculus</i> Hopffer, 1874	Guyana - LT
= <i>thericles grinda</i> Evans, 1953, syn. n.	Guyana - HT
b) <i>proculus ponicia</i> Evans, 1951, comb. n.	French Guiana (Cayenne) - HT
<i>pseudohadassa</i> Mabilles & Boulet, 1908, stat. rest.	Peru (Upper Amazon) - LT
= <i>tenuis</i> Mabilles & Boulet, 1908, praeocc. (Staudinger, 1888), syn. n.	Peru (Junín) - LT
<i>pusca</i> Evans, 1951	Peru (Junín) - HT
<i>rafael</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (Huánuco) - HT
<i>sabrina</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (Junín) - HT
<i>sadia</i> Evans, 1951	Ecuador (Pastaza) - HT
<i>sanpedro</i> O. Mielke & Lamas, sp. n.	Peru (Cusco) - HT
<i>sarpedon</i> (Bell, 1931) (<i>Yanguna</i>)	Peru (Loreto) - HT
<i>schausi</i> Bell, 1931	Ecuador - HT
<i>sergius</i> Hopffer, 1874	
a) <i>sergius sergius</i> Hopffer, 1874	Peru (Junín) - LT
b) <i>sergius andronicus</i> Bell, 1931	Ecuador (Morona-Santiago) - HT
c) <i>sergius andros</i> Evans, 1951	Colombia - HT
d) <i>sergius selina</i> Evans, 1951	Brazil (Pará) - HT
e) <i>sergius semana</i> Evans, 1951	French Guiana (Saint-Laurent-du-Maroni) - HT
<i>shiva</i> Evans, 1951, stat. n.	Bolivia (La Paz) - HT
<i>steyermarki</i> Orellana, Costa & Grishin, 2021	Venezuela (Bolívar) - HT
<i>tatei</i> (Bell, 1932) (<i>Yanguna</i>)	Venezuela (Amazonas) - HT
<i>telassa</i> Hewitson, 1866 (<i>Pyrrhopyga</i> [sic])	[Ecuador] - LT
<i>telassina</i> Staudinger, 1888	Peru (Junín) - LT
= <i>cleopas</i> Mabilles, 1903, nom. nud. (ICZN, Art. 12.1)	Peru (Pasco)
= <i>cleopas</i> Mabilles & Boulet, 1908, syn. n.	Peru (Pasco) - LT
= <i>telassa silex</i> Evans, 1951, syn. n.	Peru (Junín) - HT
<i>terra</i> Evans, 1951	Bolivia - HT
<i>thericles</i> Mabilles, 1891	
a) <i>thericles thericles</i> Mabilles, 1891 (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT
b) <i>thericles fola</i> Evans, 1951	Colombia (Cundinamarca) - HT
c) <i>thericles orientis</i> Bell, 1947	Brazil (Pará) - HT
d) <i>thericles pseudophidias</i> Bell, 1931	Colombia (Boyacá) - HT
e) <i>thericles raymondi</i> Orellana, [2010]	Venezuela (Guárico) - HT
f) <i>thericles rileyi</i> Bell, 1931	Bolivia (Santa Cruz) - HT
g) <i>thericles ronda</i> Evans, 1953	Trinidad and Tobago (Port of Spain) - HT
<i>tingo</i> O. Mielke, Brockmann & C. Mielke, sp. n.	Peru (Huánuco) - HT
<i>zenodorus</i> Godman & Salvin, 1893	Guatemala (Alta Verapaz) - LT
<i>Yanguna</i> Watson, 1893	
<i>blossomiae</i> Williams, 1931, stat. rev.	
a) <i>blossomiae blossomiae</i> Williams, 1931, stat. rev.	Ecuador (Chimborazo) - HT
b) <i>blossomiae galion</i> O. Mielke, Brockmann & C. Mielke, ssp. n.	French Guiana (Cayenne) - HT
<i>aspilos</i> Mabilles & Boulet, 1908, stat. rest.	
a) <i>aspilos aspilos</i> Mabilles & Boulet, 1908, stat. rest.	[South America] - LT
b) <i>aspilos lara</i> O. Mielke, Brockmann & C. Mielke, ssp. n.	Peru (Huánuco) - HT
c) <i>aspilos mabillei</i> H. H. Druce, 1909, comb. n.	Peru (Pasco) - LT
<i>cometes</i> (Cramer, 1779) (<i>Papilio</i>)	
a) <i>cometes cometes</i> (Cramer, 1779) (<i>Papilio</i>)	French Guiana (Saint-Laurent-du-Maroni) - NT
b) <i>cometes angeli</i> Orellana, [2010]	Venezuela (Táchira) - HT
c) <i>cometes cometides</i> Mabilles & Boulet, 1908	Bolivia - HT
d) <i>cometes staudingeri</i> (Plötz, 1879) (<i>Pyrrhopyga</i> [sic])	Peru (Huánuco) - NT
<i>cosyra</i> (H. Druce, 1875) (<i>Pyrrhopyga</i> [sic])	Panama (Chiriquí) - LT
<i>erebus</i> (Plötz, 1879) (<i>Erycides</i>)	Brazil (Bahia) - HT
<i>moa</i> O. Mielke, Casagrande & Brockmann, sp. n.	
a) <i>moa moa</i> O. Mielke, Casagrande & Brockmann, sp. n.	Peru (Huánuco) - HT
b) <i>moa guiana</i> O. Mielke, Casagrande & Brockmann, ssp. n.	French Guiana (Cayenne) - HT
<i>spatiosa</i> (Hewitson, 1870) (<i>Pyrrhopyga</i> [sic])	

a) <i>spatiosa spatiosa</i> (Hewitson, 1870) (<i>Pyrrhopyga</i> [sic])	Ecuador (Tungurahua) - LT
b) <i>spatiosa julian</i> O. Mielke, Brockmann & C. Mielke, ssp. n.	Colombia (Valle del Cauca) - HT
<i>tetricus</i> Bell, 1931	Guyana (Cuyuni Mazaruni – Roraima) - HT
<i>thelersa</i> (Hewitson, 1866) (<i>Pyrrhopyga</i> [sic])	Brazil (Amazonas) - LT

Zoniini O. Mielke, 2001

Zonia Evans, 1951

<i>zonias</i> Evans, 1951	
a) <i>zonias zonia</i> Evans, 1951	Brazil (Amazonas) - HT
b) <i>zonias diabo</i> O. Mielke & Casagrande, 1998	Brazil (São Paulo) - HT
c) <i>zonias eberti</i> O. Mielke, C. Mielke & Casagrande, ssp. n.	Brazil (Rio de Janeiro) - HT
d) <i>zonias panamensis</i> Nicolay, 1975	Panama (Panamá) - HT

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